

*Etablissement de la polarité neuronale...*

**Synapses  
& Synaptogénèse**

*Master 2 - 2013*

*... et trafic membranaire*

**Lydia Danglot**

INSTITUT JACQUES MONOD

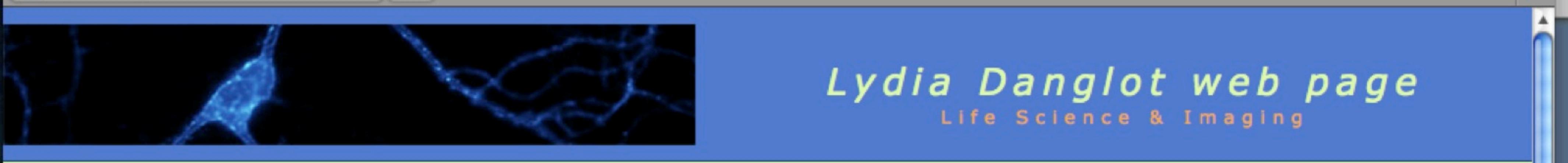


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free Lydia Danglot - Cours de neurosci... +



Novembre 2, 2009

Thème de recherche

Publications

Enseignement

Liens favoris

CONTACT

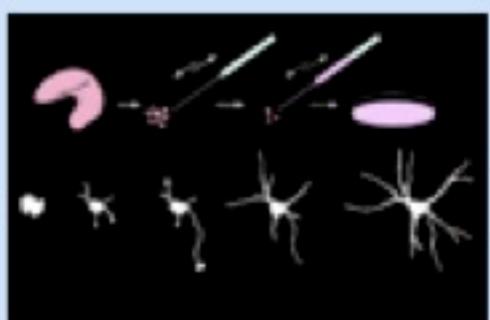
[French](#)[English](#)

## Enseignement

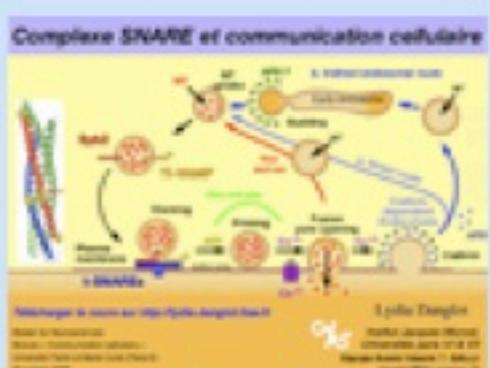
### Cours

- [Master2 de Neurosciences - UE Synapse et synaptogenèse \(code UE : MBIP5019\) - Université Pierre et Marie Curie \(Paris 6\):](#)  
Planning [Neuritogenèse et polarité neuronale](#).
- [Master2 de Neurosciences - UE Communication Cellulaire](#)  
(code UE : MBIP5003) - Université Pierre et Marie Curie (Paris 6):  
[Les protéines SNARE et l'exocytose](#) : classification des SNAREs, voie de recyclage des VS, comment mesurer l'exocytose, comment mesurer le recyclage, les protéines régulant l'assemblage des SNARE (Munc18, munc13, Syt, complexine), souris KO Syb2, souris mocha,...
- [Master2 de Génétique - Université Paris Diderot \(Paris 7\)](#), UE Neurobiologie cellulaire et développementale.  
[Développement de l'hippocampe et synaptogenèse](#): Neuroanatomie générale, présentation du SNC, présentation du télencéphale et de l'hippocampe, développement de l'hippocampe, migration des neurones excitateurs et inhibiteurs, modèle des neurones dissociés d'hippocampe en culture, polarité neuronale, formation des synapses.
- [Ecole doctorale Frontières du Vivant \(Universités Paris V, VI, VII\)](#)  
Club Neurobiologie & Optique: [Diversité et usage des protéines fluorescentes en Neurosciences](#).

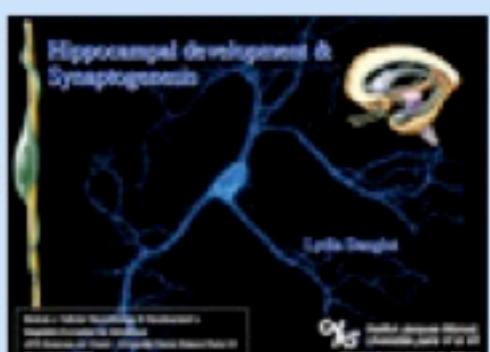
### MANUEL de cours



Master2- Paris 6  
Neuritogenèse et polarité neuronale.

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Master2- Paris 6  
Complexe SNARE et communication cellulaire.

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Master2- Paris 7  
Développement de l'hippocampe et synaptogenèse

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# Plan

1. *Le neurone: unité fonctionnelle polarisée*
2. *D'où proviennent les neurones*
3. *Polarité neuronale: les domaines*
4. *Rôle du trafic membranaire dans la neuritogenèse*
5. *Polarité et cytosquelette*
6. *La métaphore (imparfaite) épithéliale*
7. *Formation de la Synapse*

# 1. Le Neuron: unité fonctionnelle polarisée

- 1800's: les techniques de microscopies permettent l'examen de tissus nerveux

Refinement of the microscope allows scientists to examine brain tissue

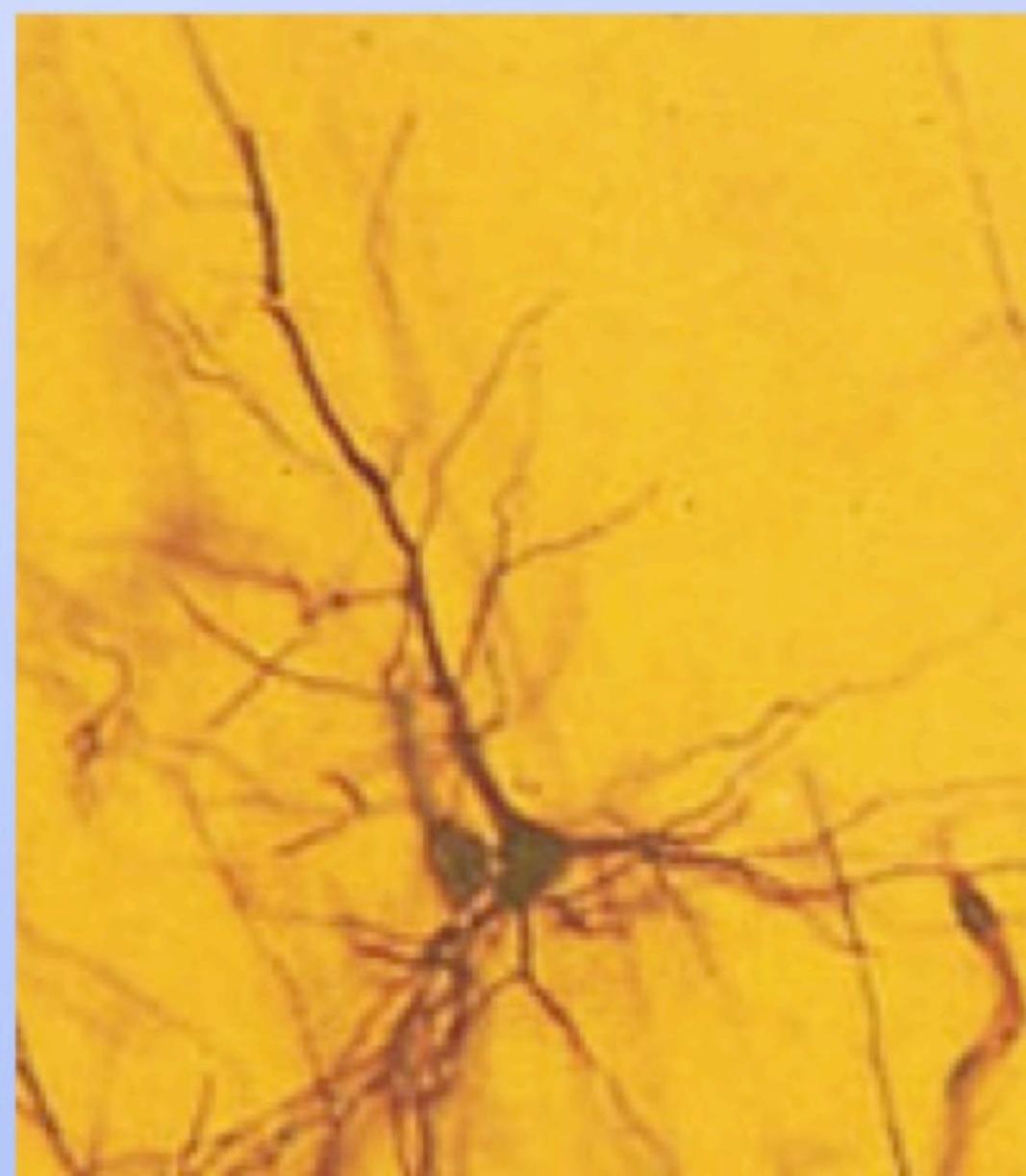


- Les tissus sont composés d'unités nommées: les cellules

All tissues are composed of microscopic units called cells

- Les cellules du cerveau sont nommés neurones

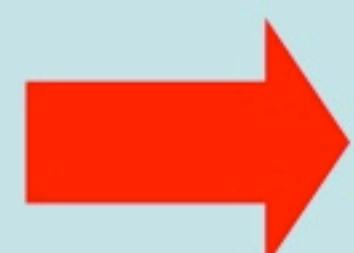
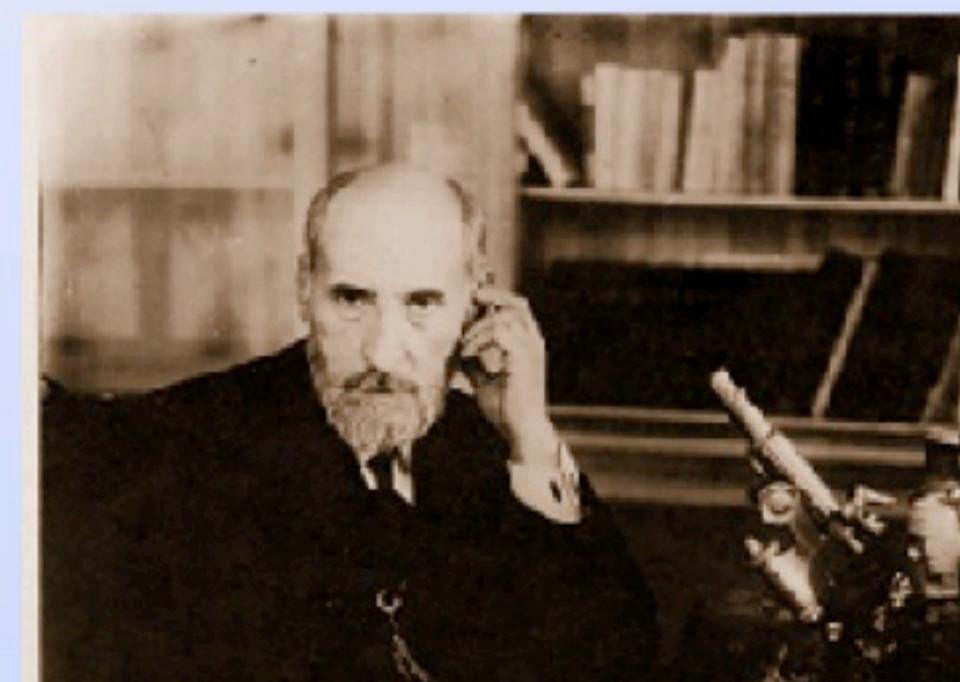
Cells in the brain are termed “neurons”



# Un peu d'histoire...

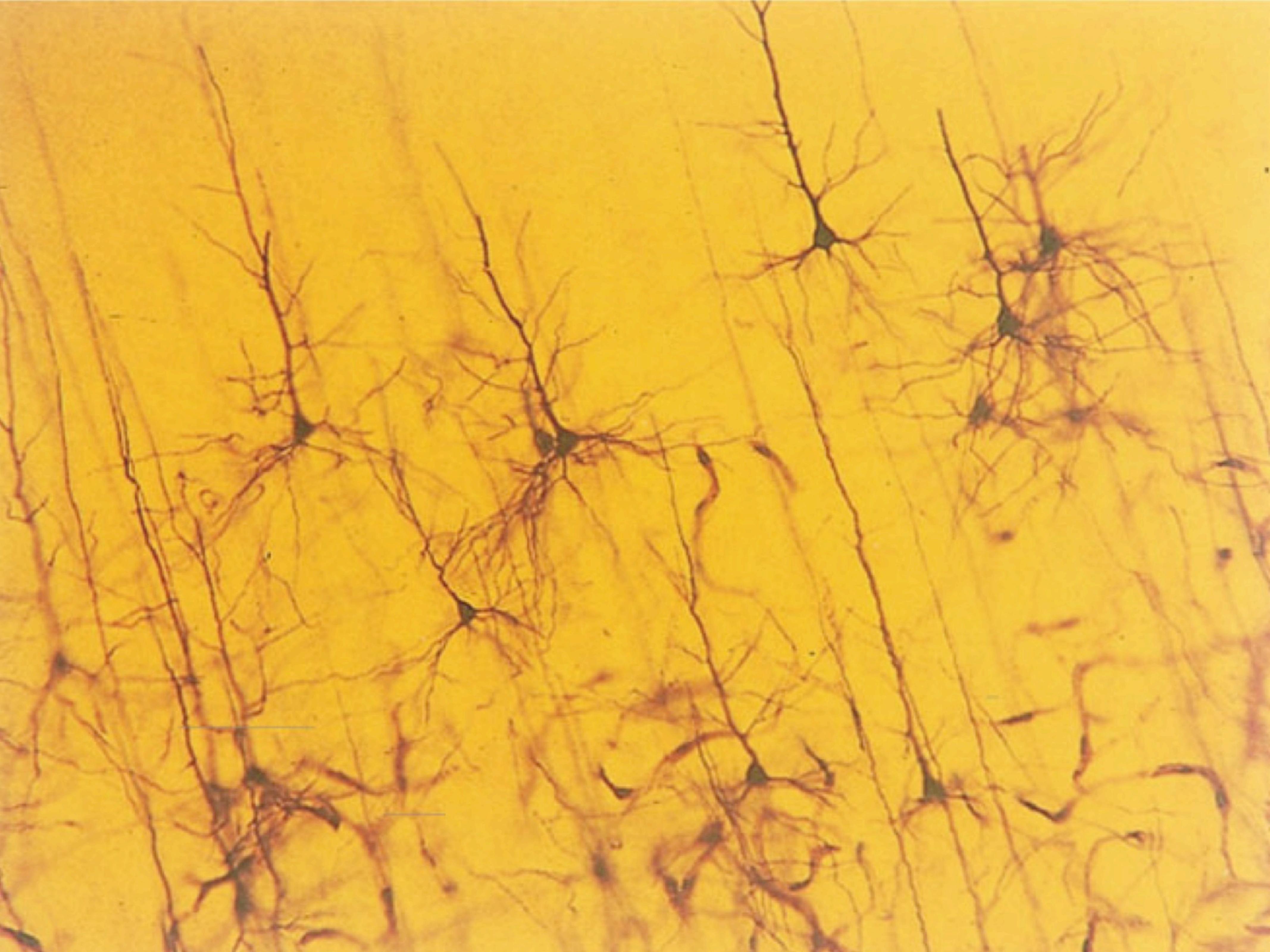


- Théorie réticulaire (C. Golgi): Il existe une continuité cytoplasmique entre les neurones (tunnel).
- Théorie du neurone (Cajal): Les neurones sont bel et bien séparé, sans continuité, leur membrane sont complètes.

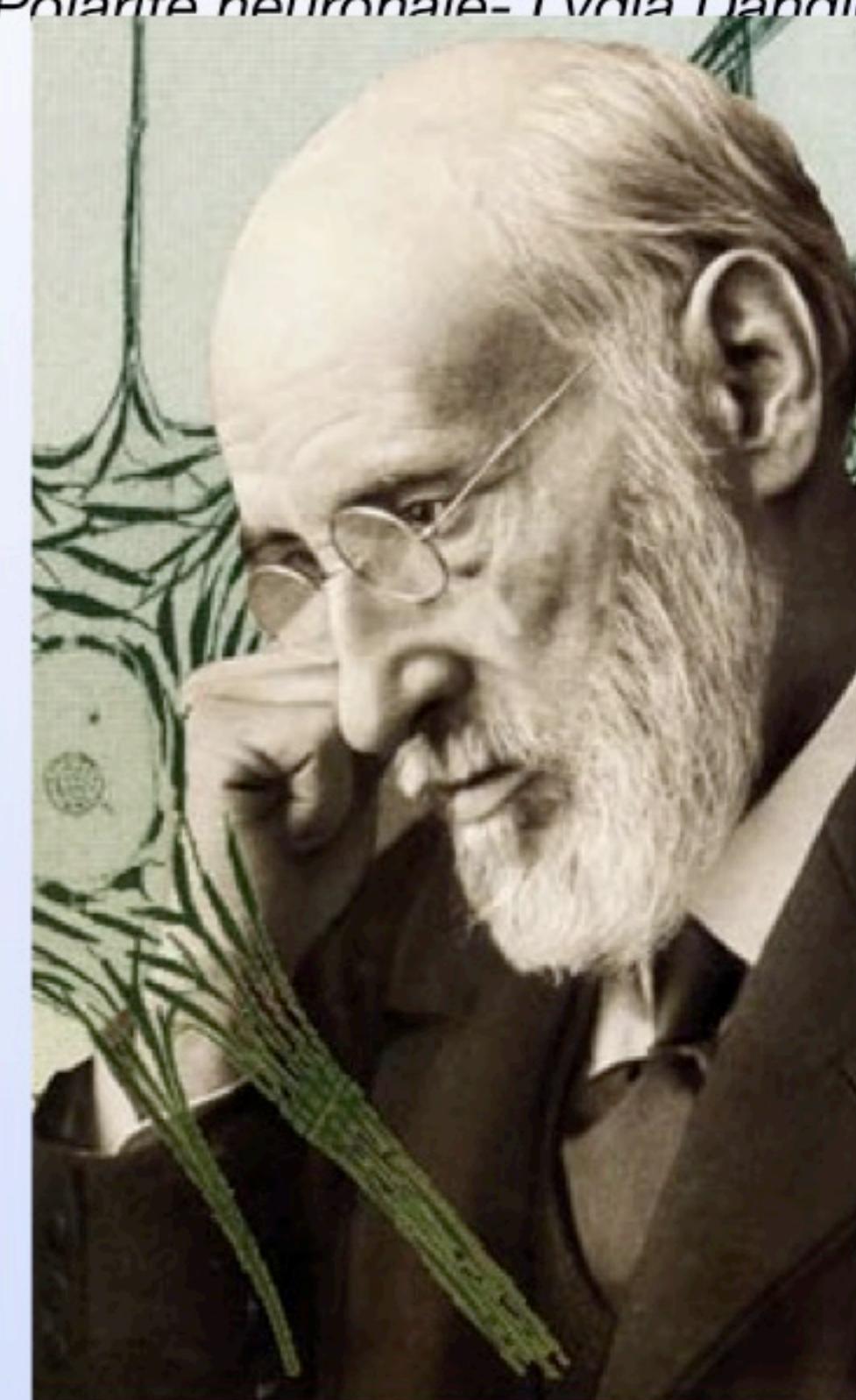
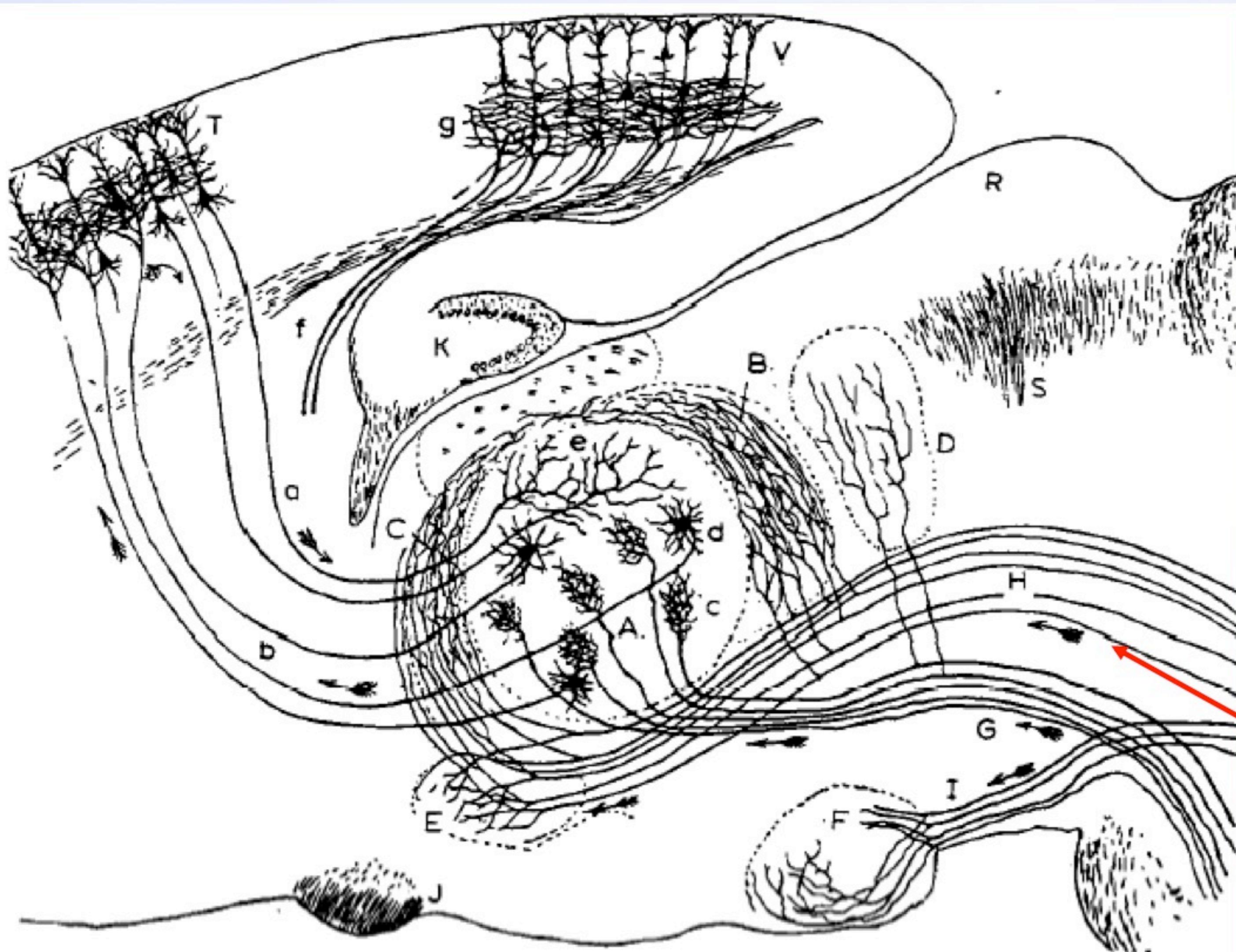


Aujourd'hui, les scientifiques donnent raison à Cajal

Ironie: Cajal a réussi à convaincre la communauté scientifique grâce à une technique histologique inventée par Golgi !  
(Les deux ont reçu le prix Nobel en 1906)

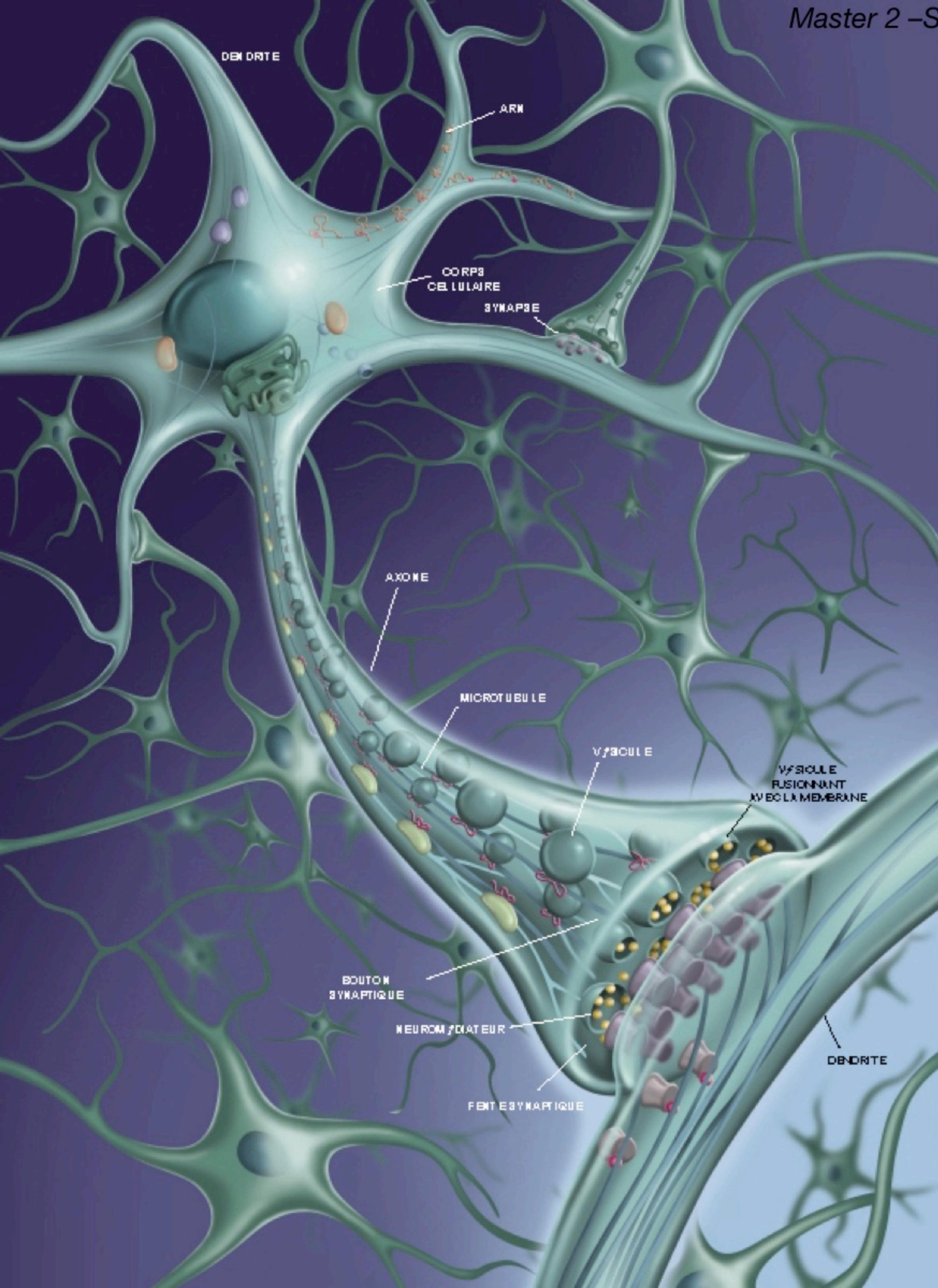


# Les neurones sont des cellules polarisées



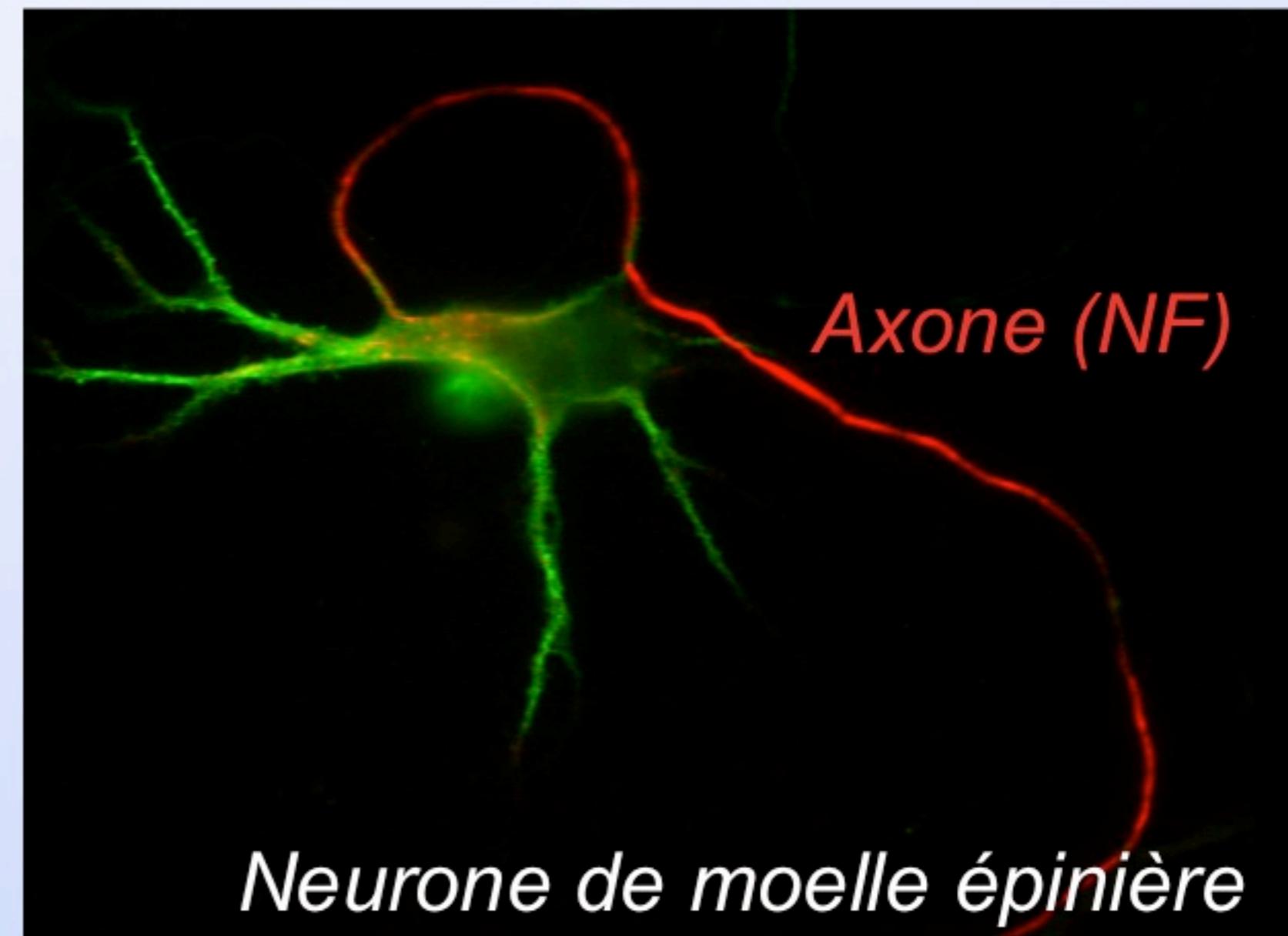
Santiago Ramón y Cajal  
(1852-1934)

Direction de  
l'influx nerveux

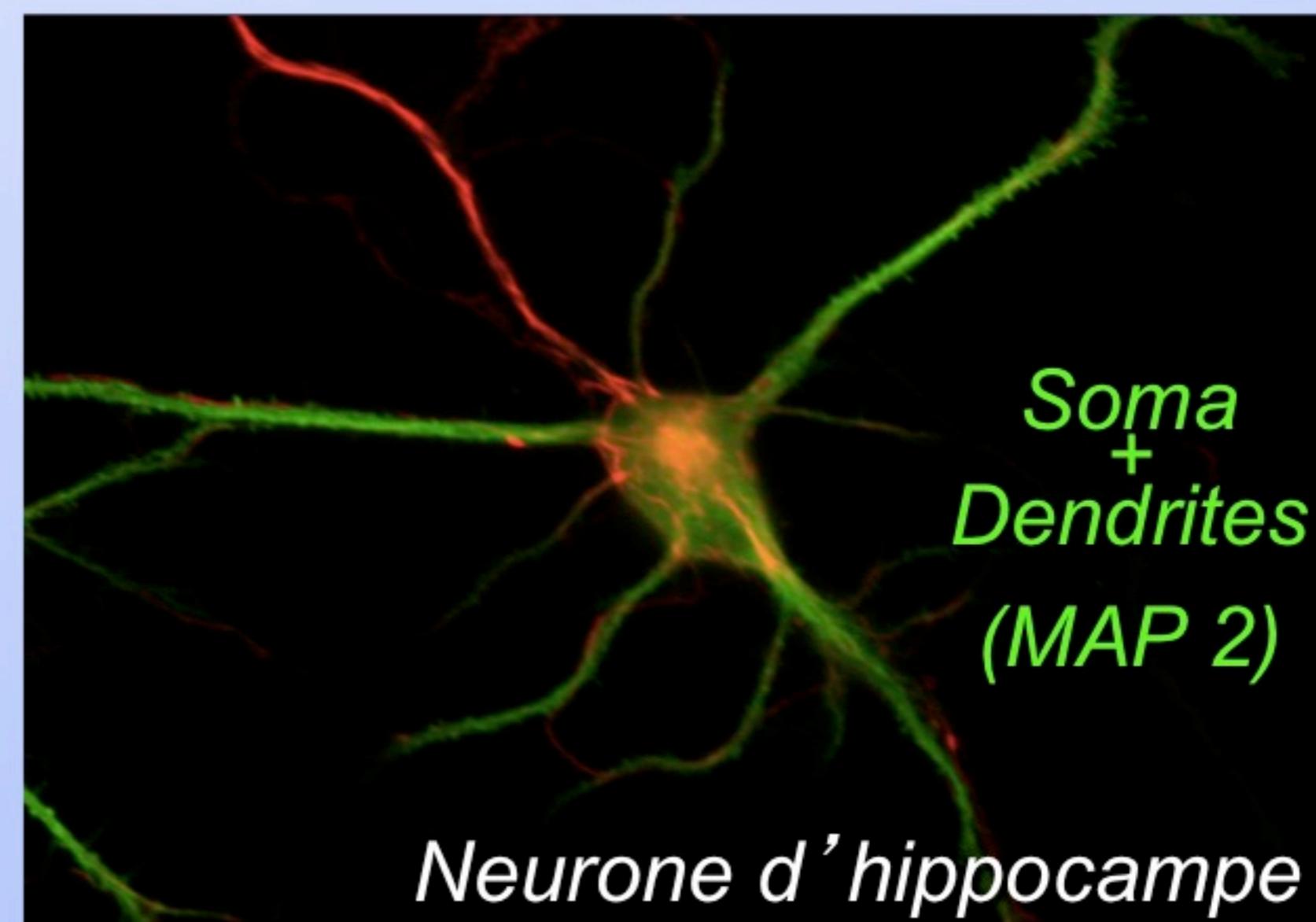


## 2 compartiments:

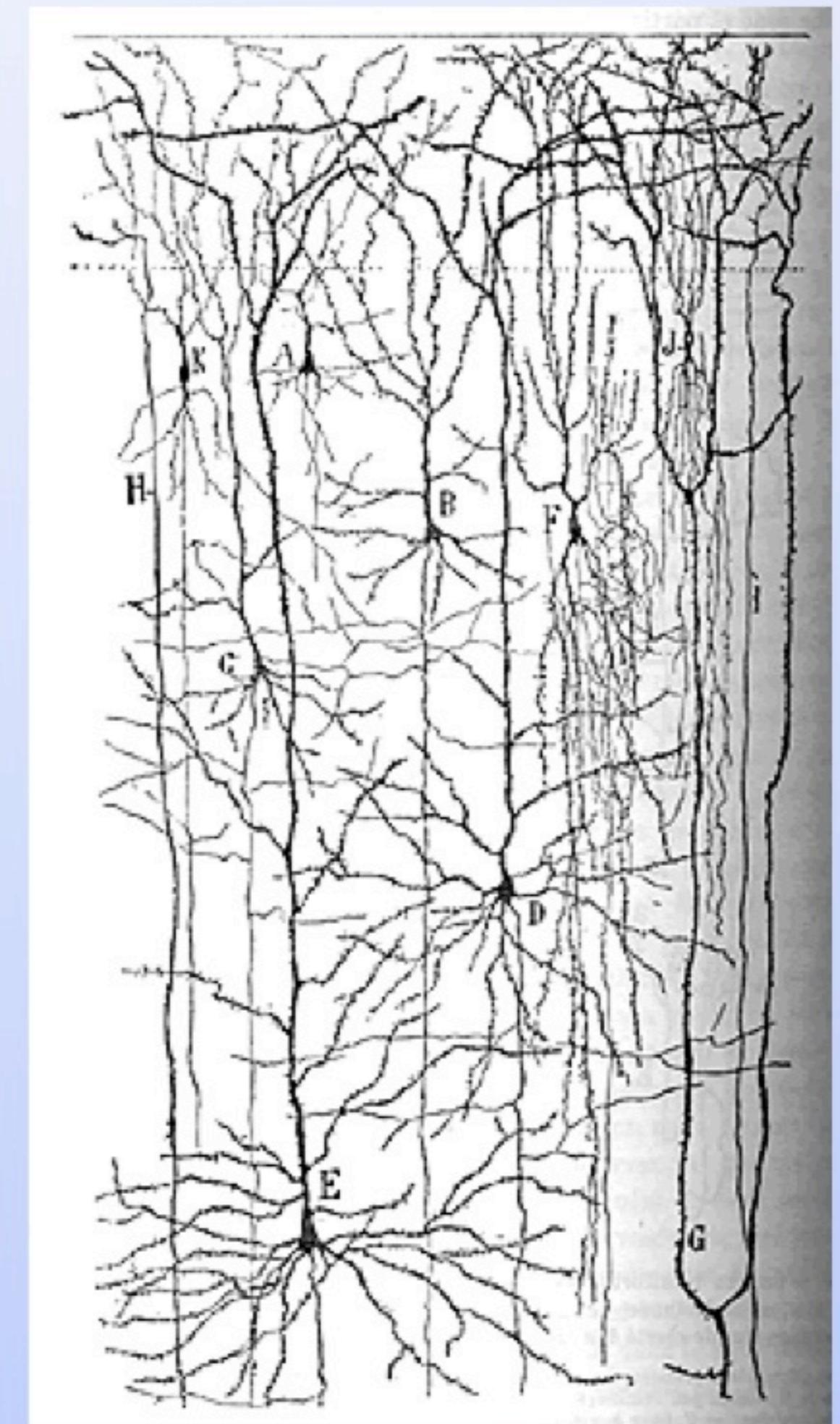
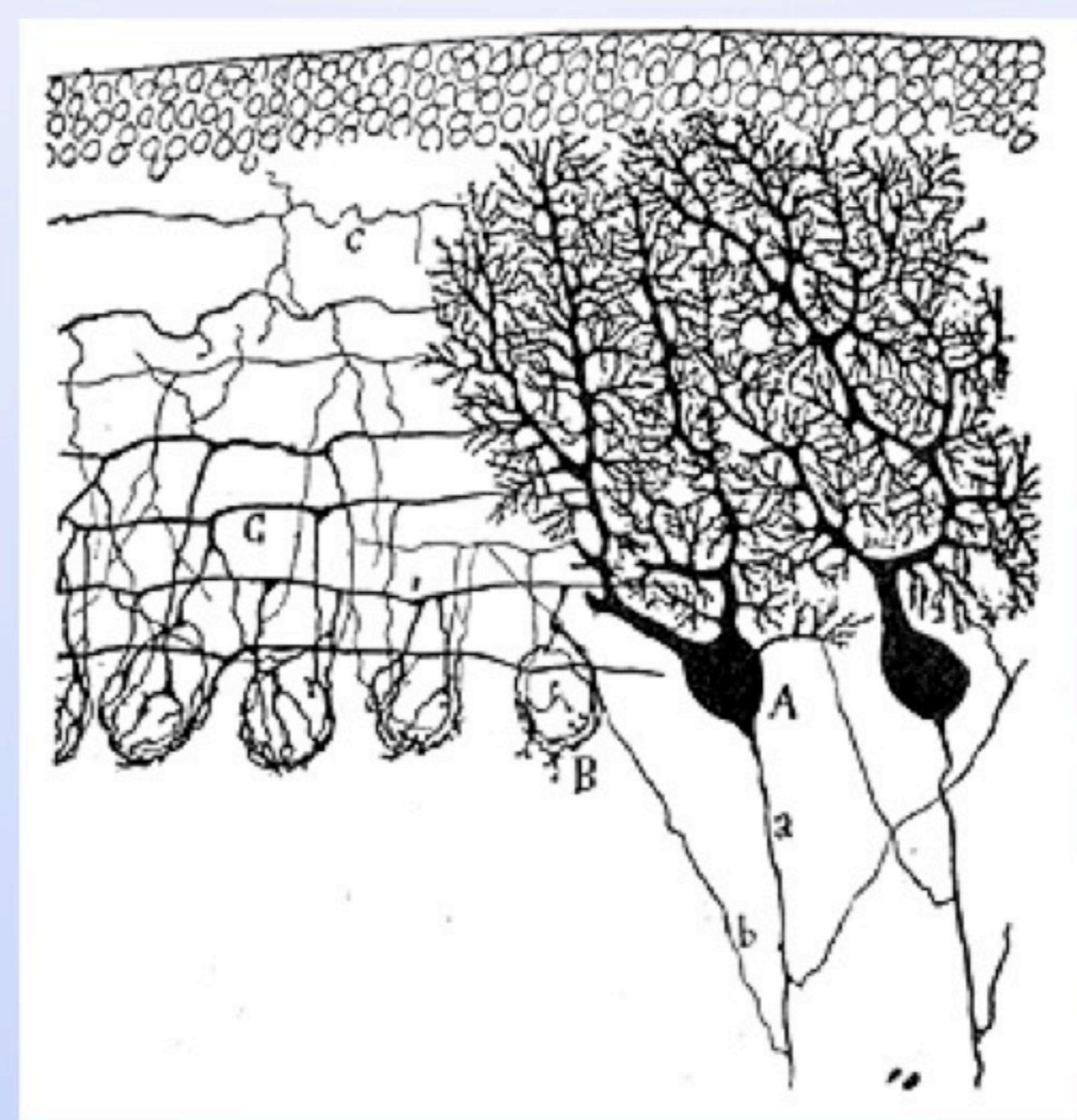
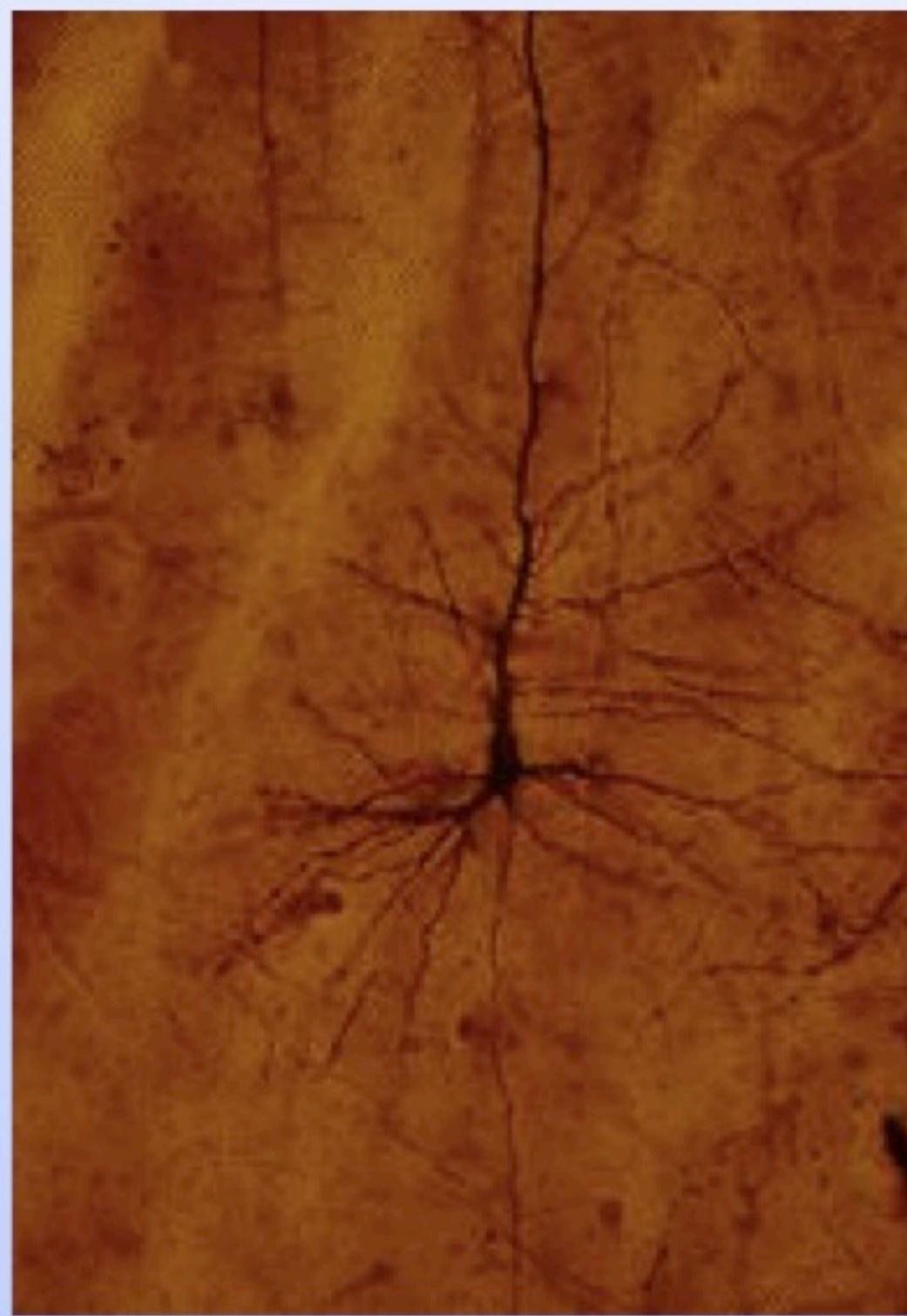
- Axonal
- Somatodendritique



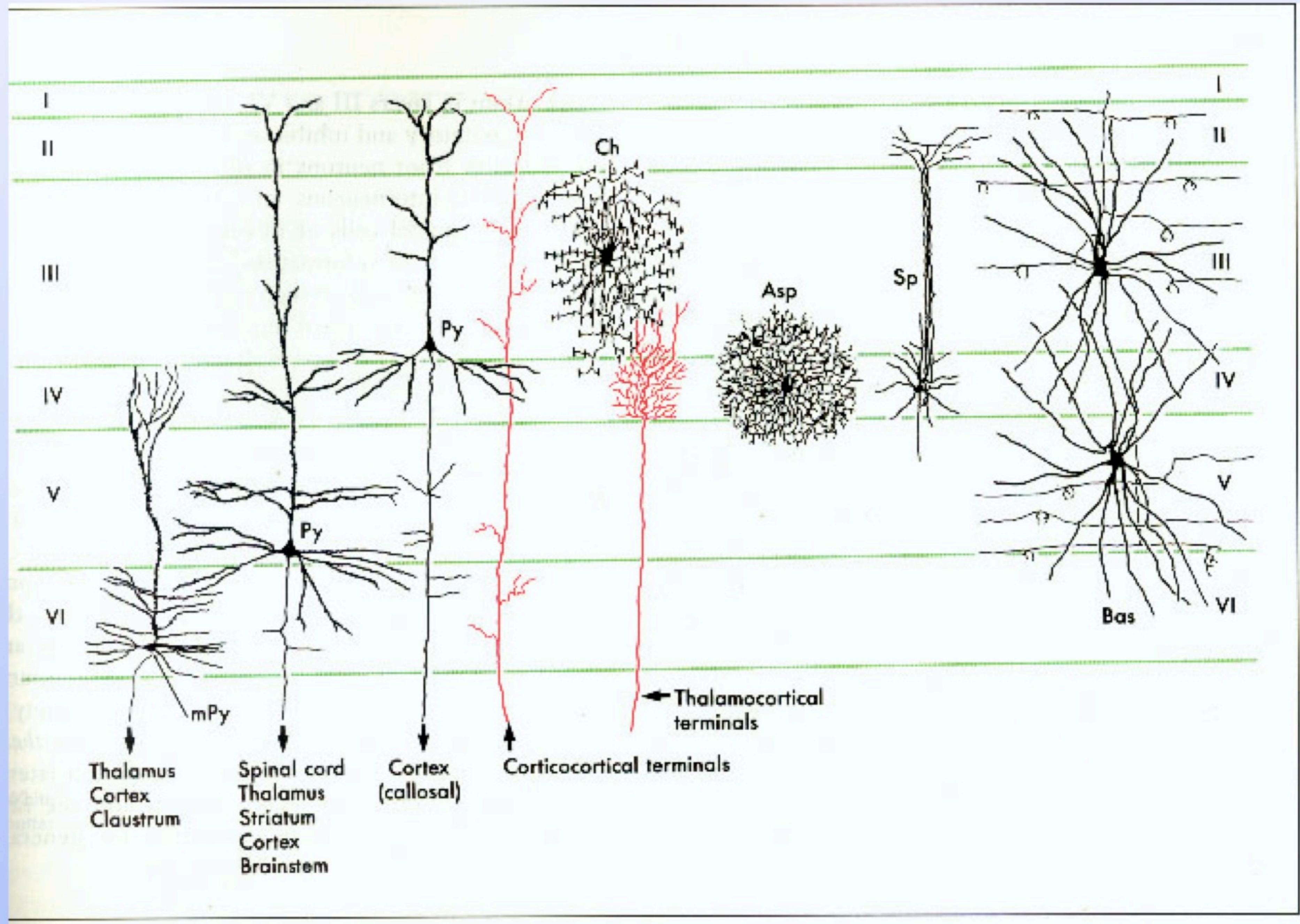
Neurone de moelle épinière



Neurone d'hippocampe

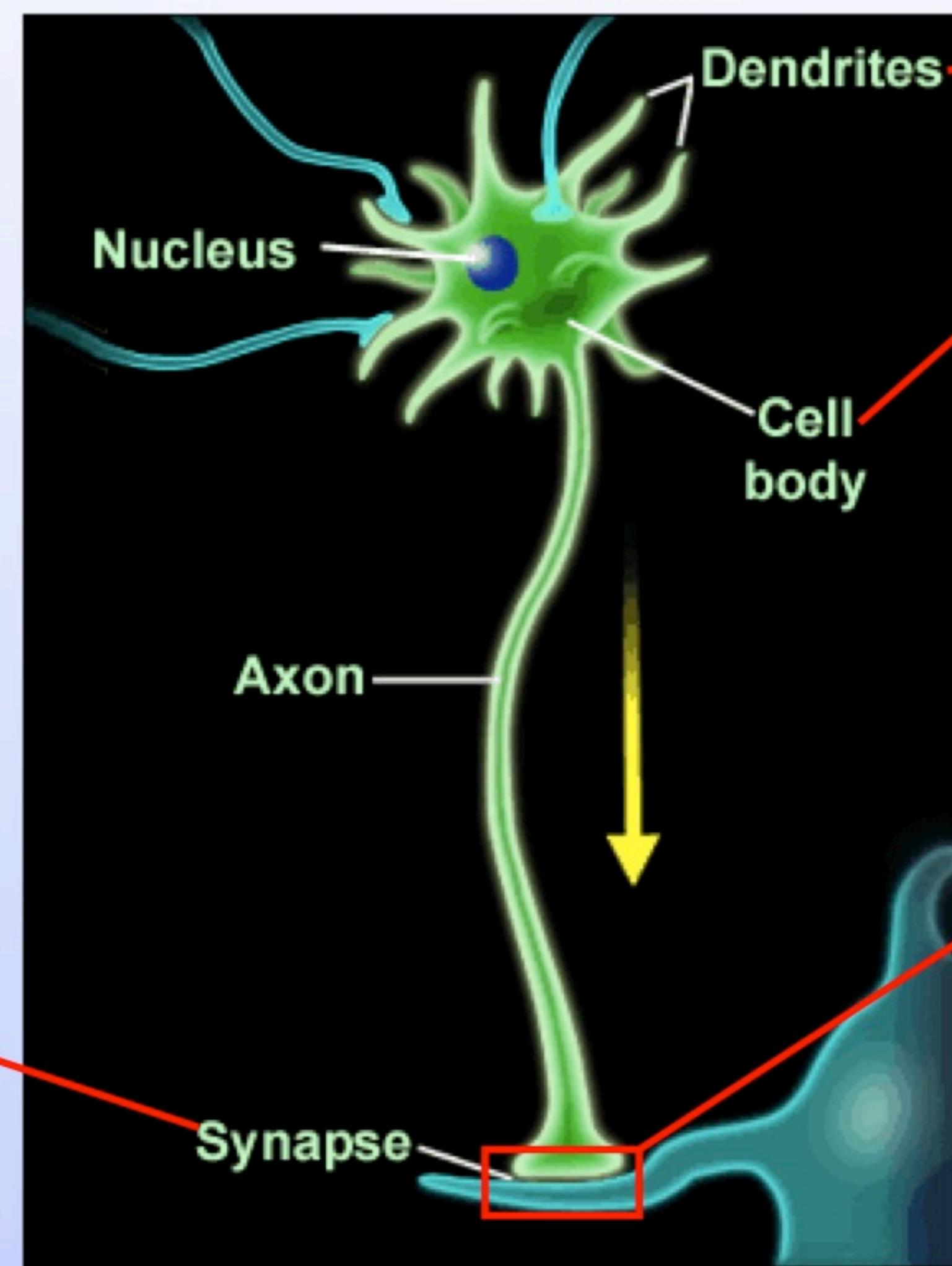


# Cerebral Cortex



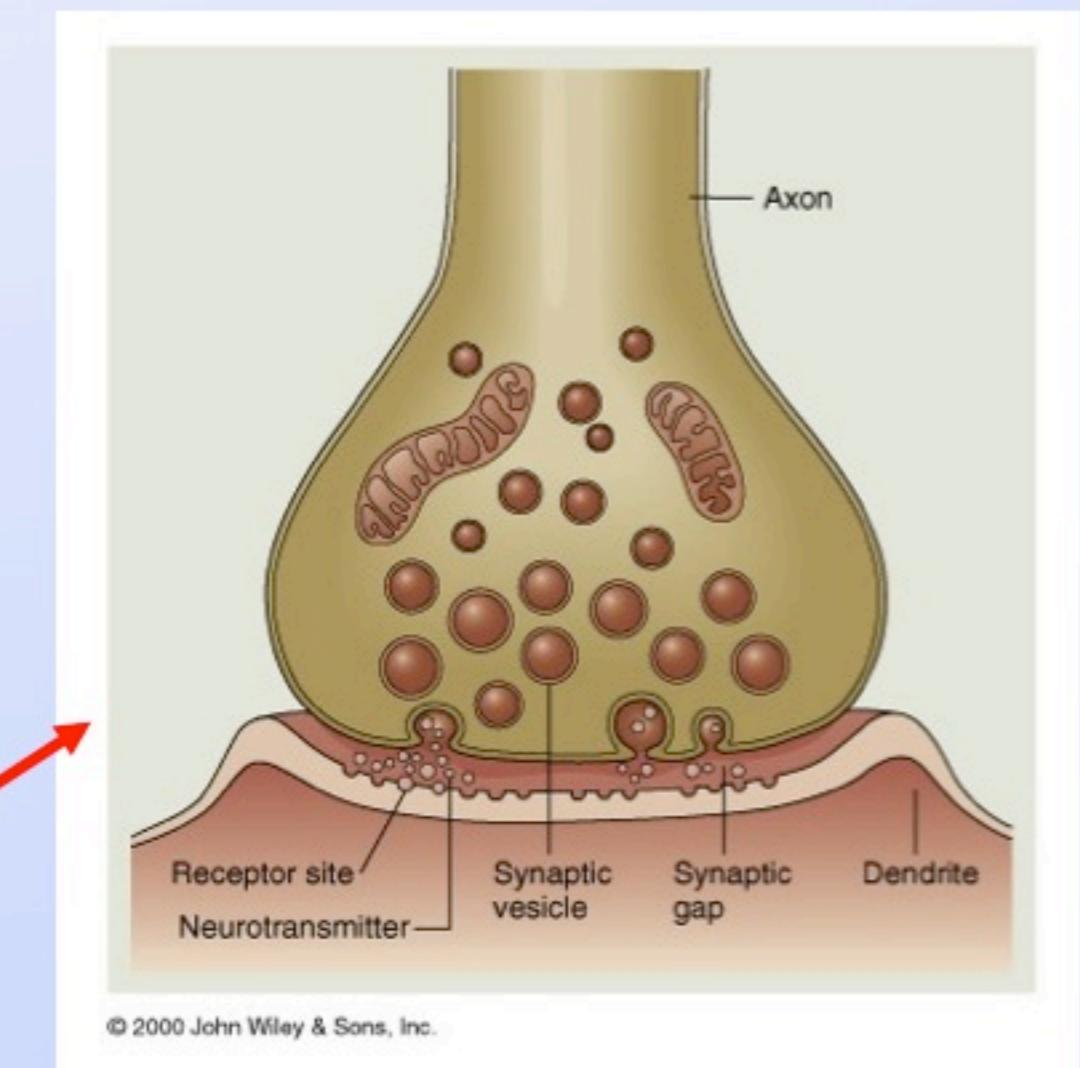
# Domaines du neurones

1 mm<sup>3</sup> de substance grise du cortex peut contenir 5 milliards de synapses.



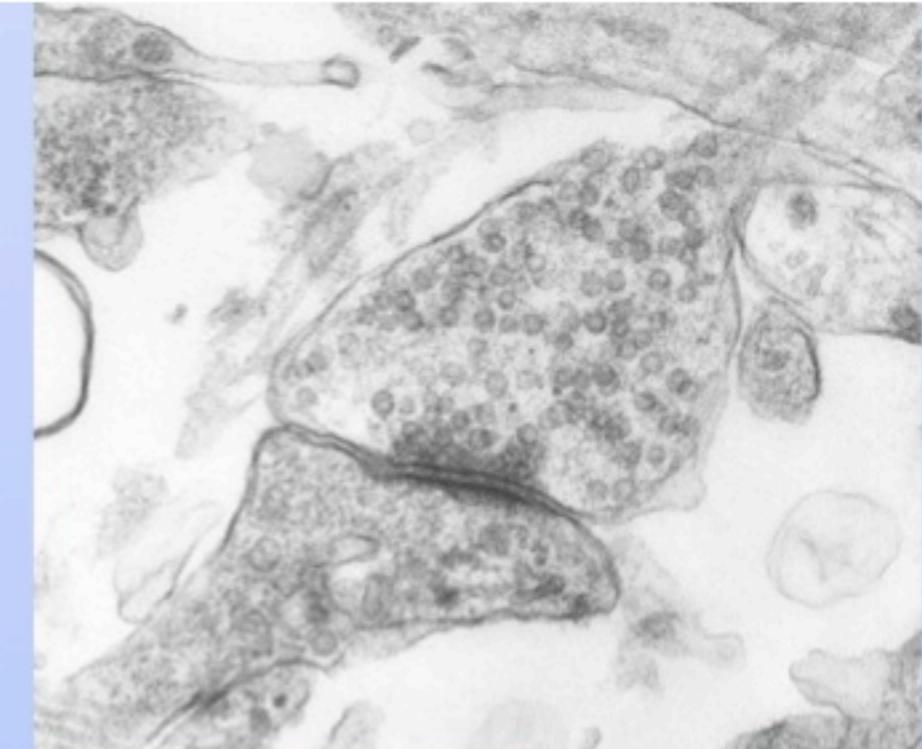
Pôle émetteur

Pôle récepteur

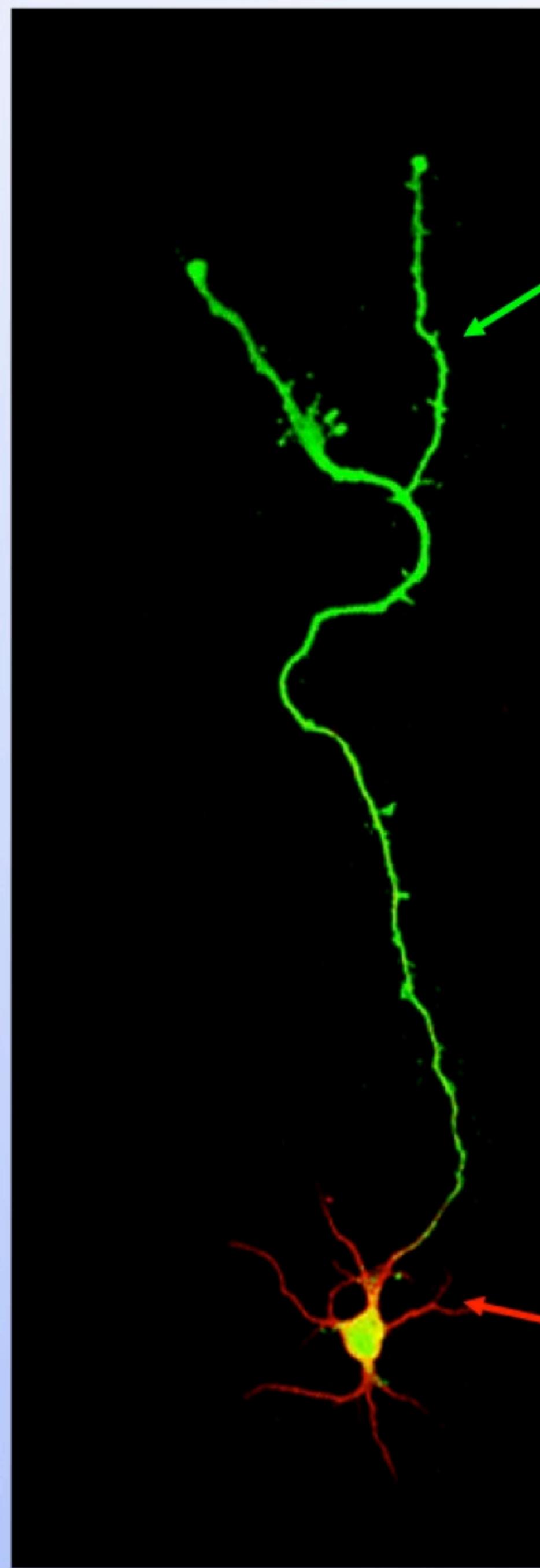


La question centrale :

Polarisation neuronale: développement de l'axone et des dendrites.

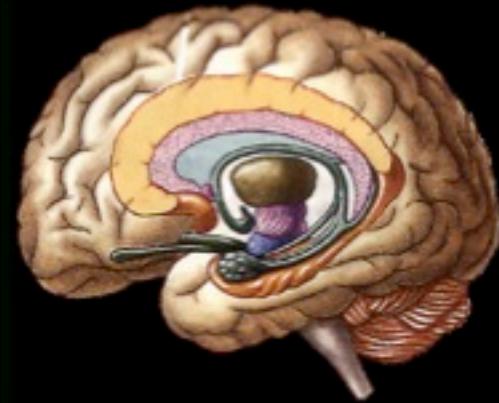


# Molecular markers for axon or dendrite

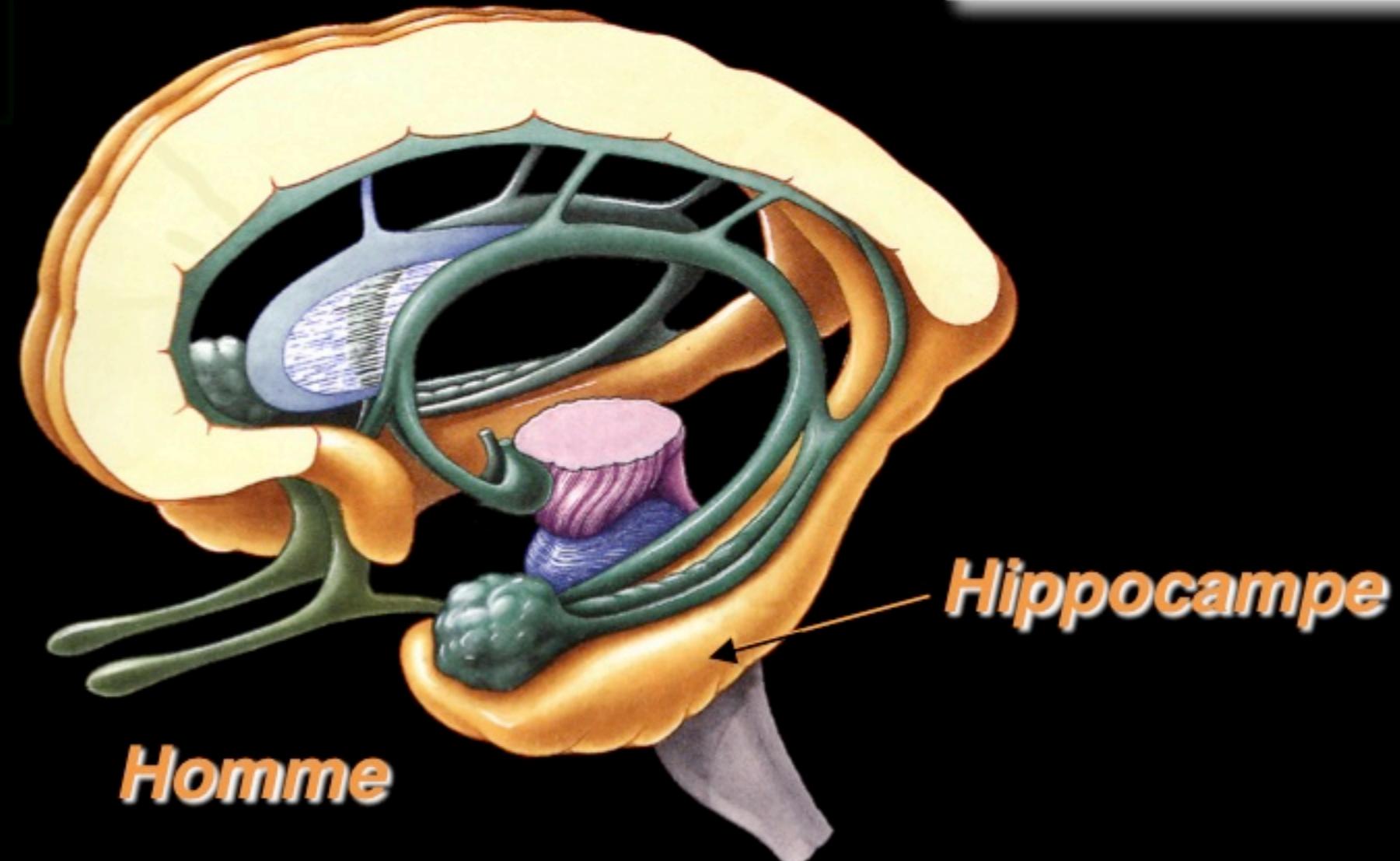


Axon: Tau1, GAP43, synapsin,  
synaptotagmin ...

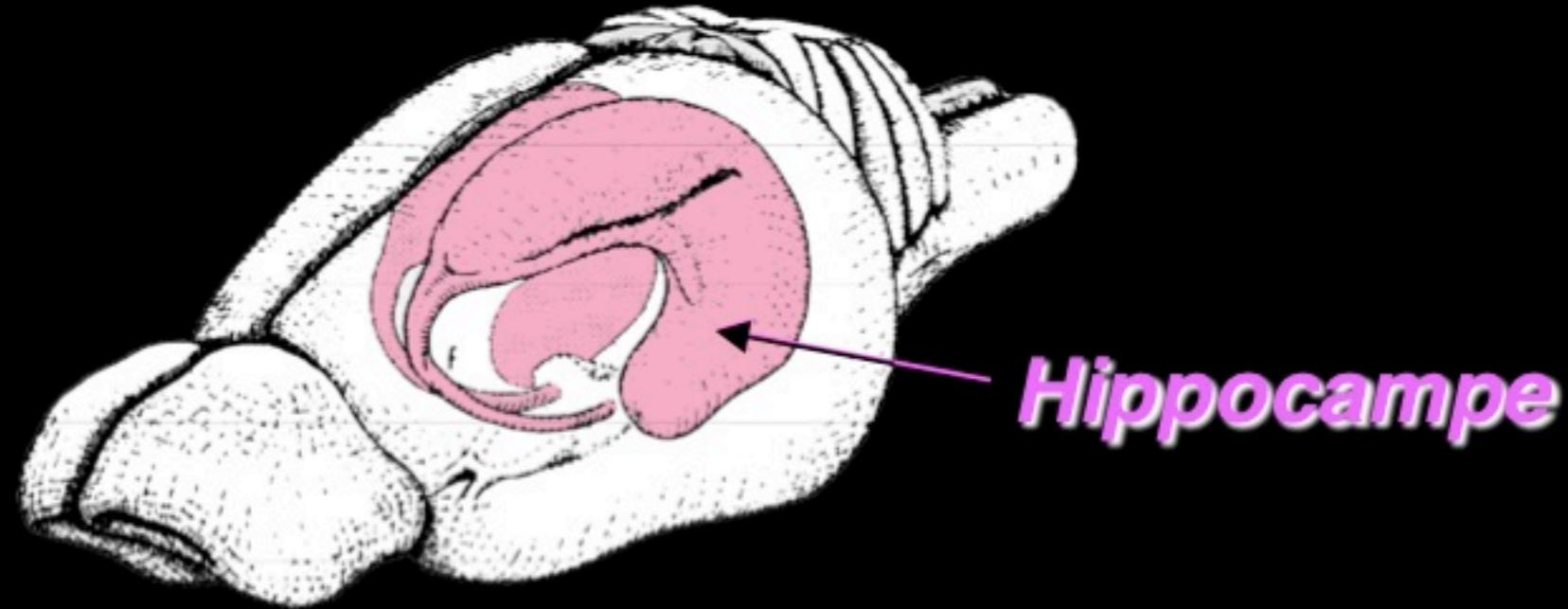
Dendrite: MAP2, Glycine receptor, GABAa  
receptor ...



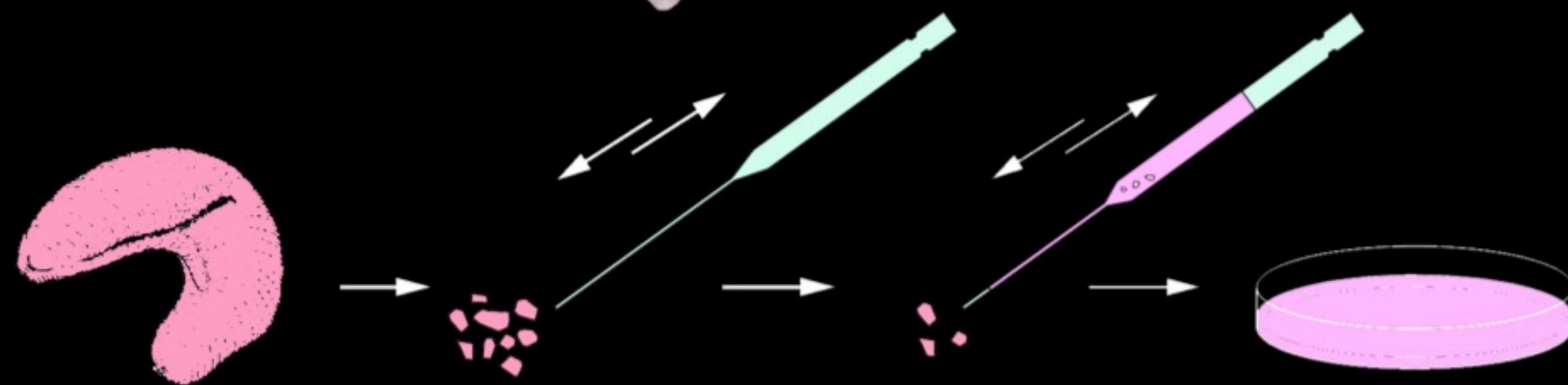
# Neurones d'hippocampe en culture



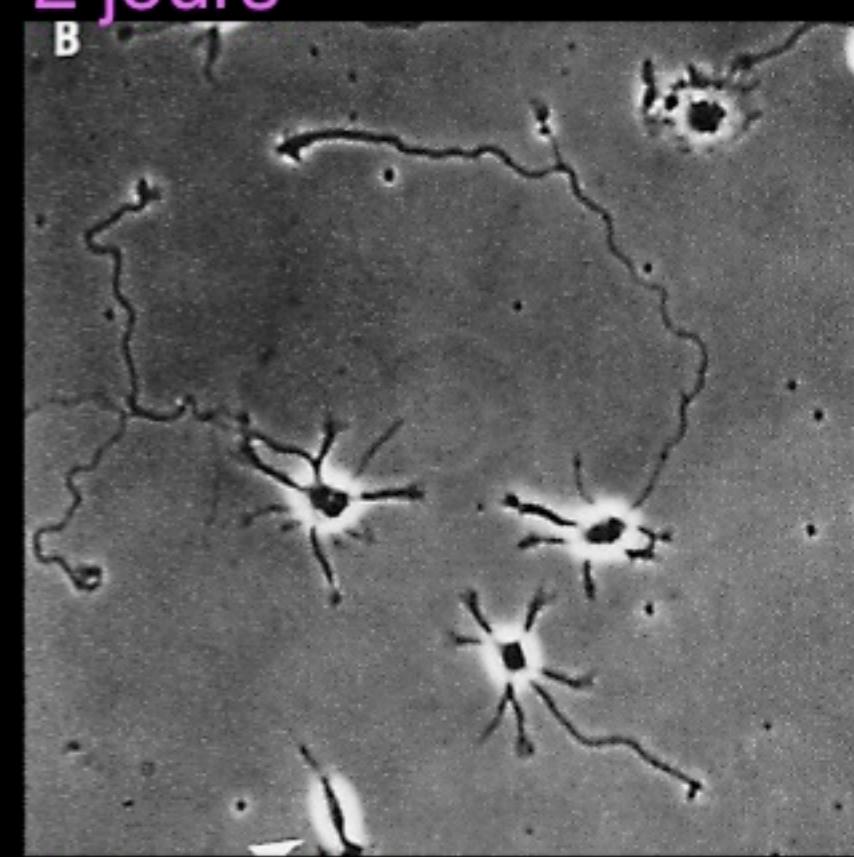
**Homme**



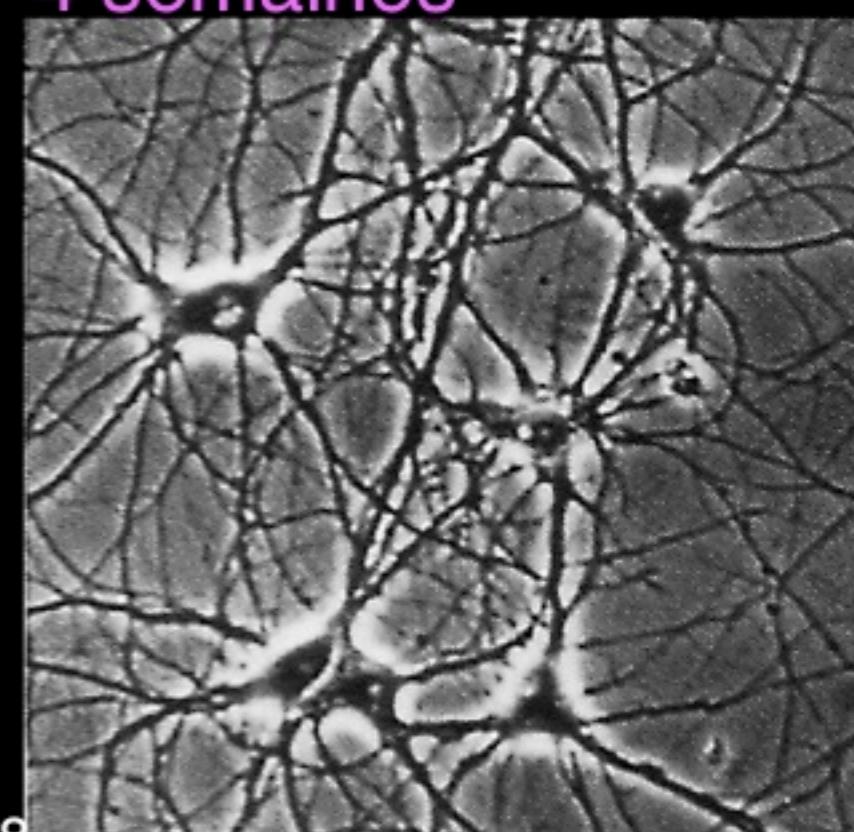
**Rat**



2 jours



4 semaines



**Temps en culture:**

6h



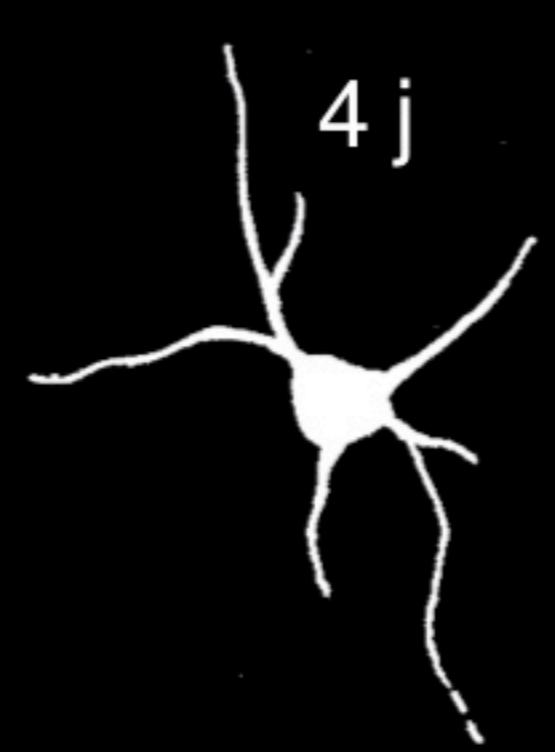
12h



1,5 j



4 j



>7j



**Stades:**

1  
Lamellipodes

2  
Neurites mineurs

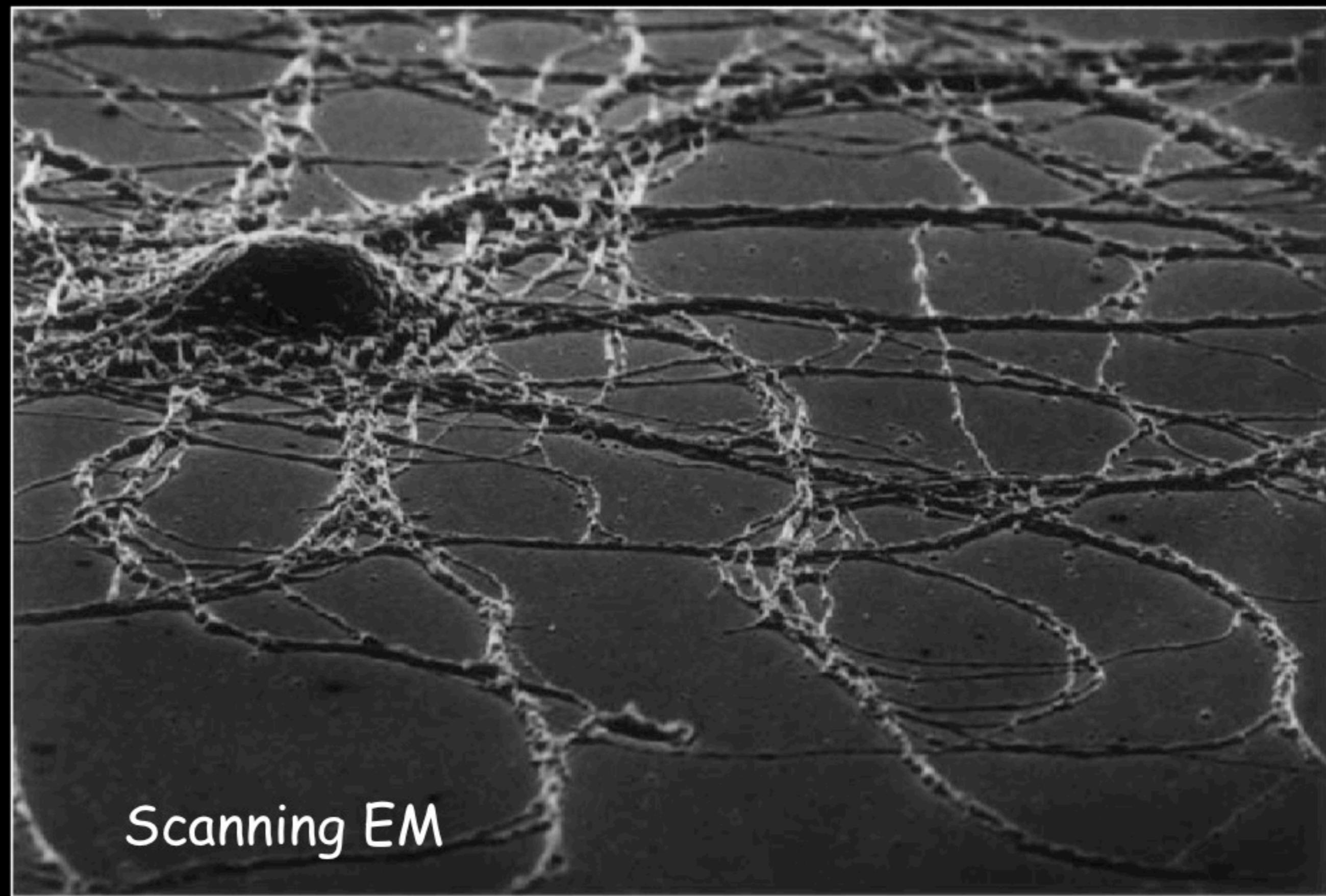
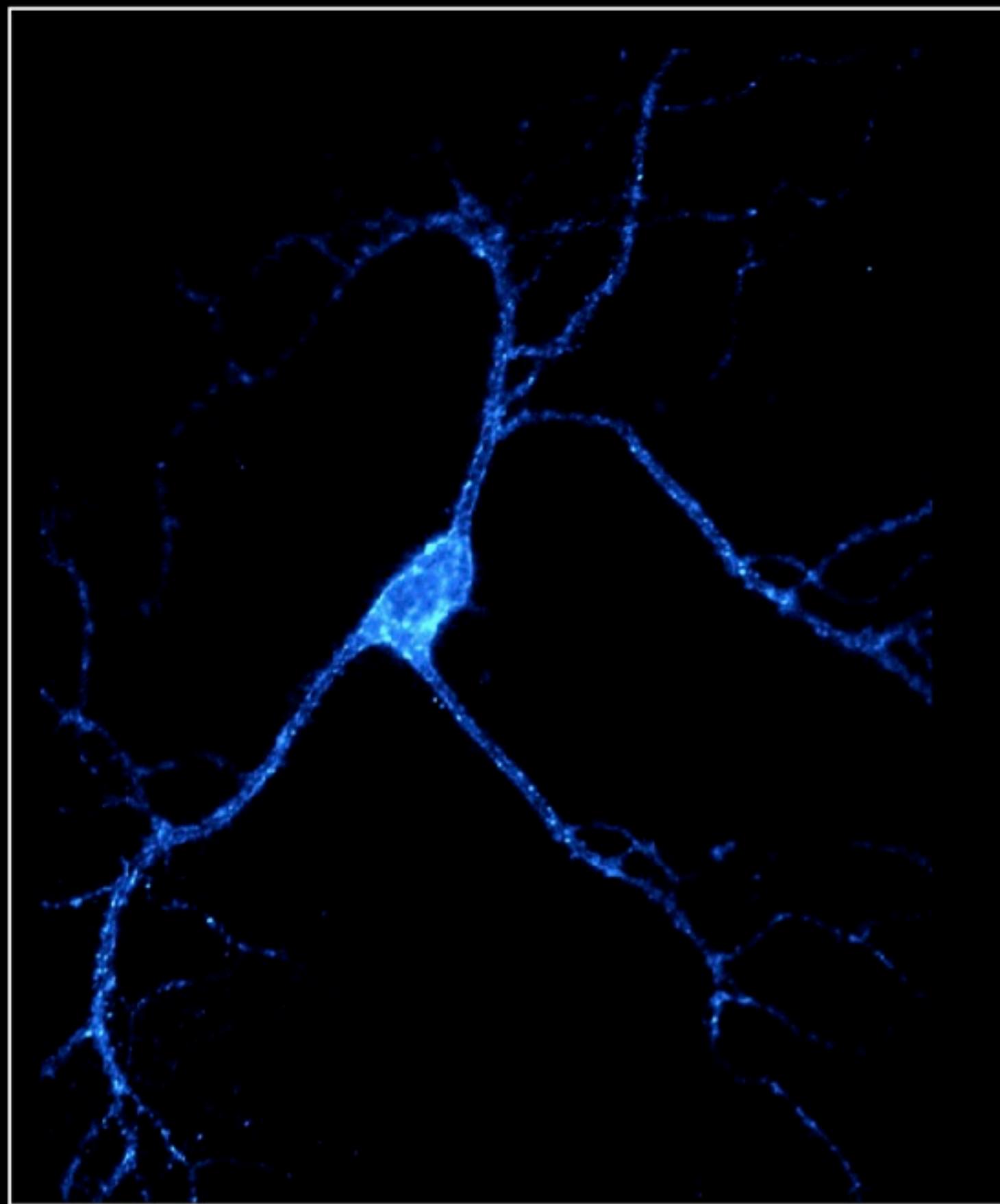
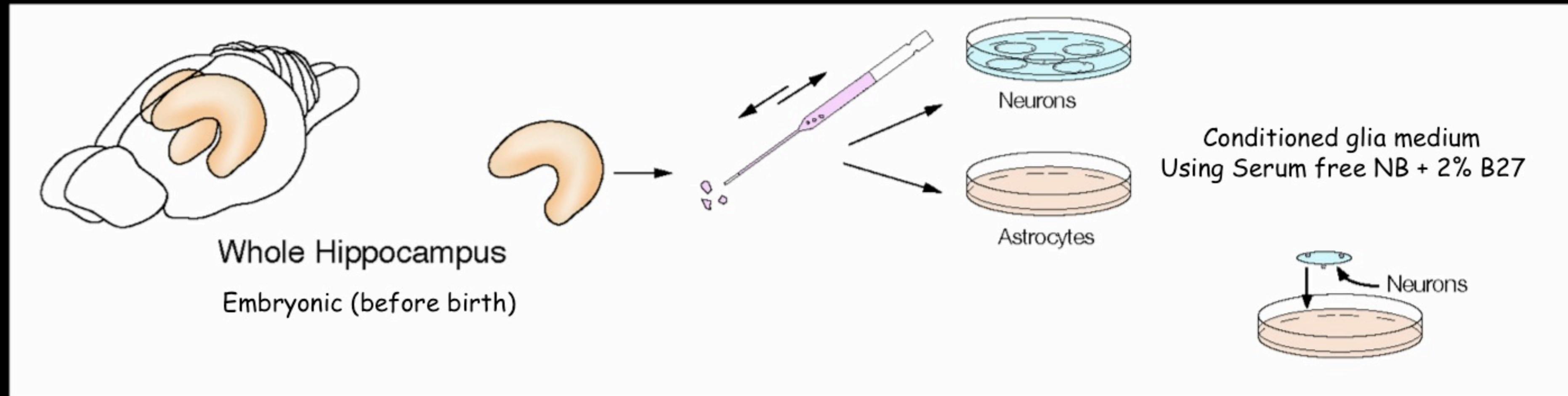
3  
Croissance axonale

4  
Croissance dendritique

5  
Maturation

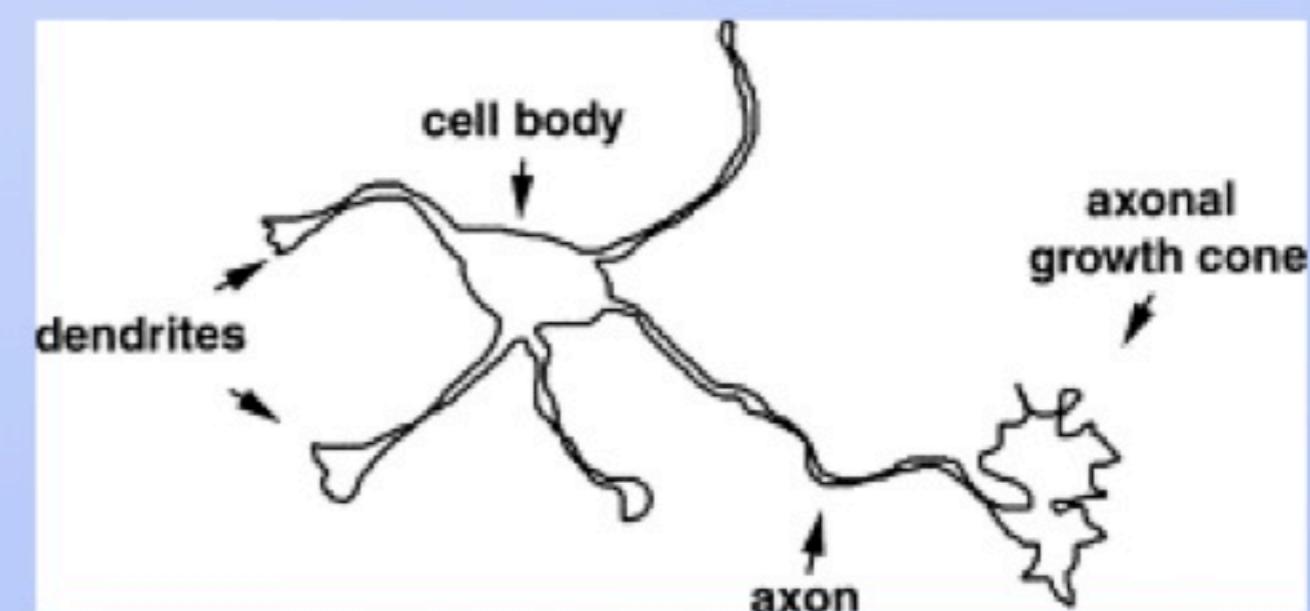
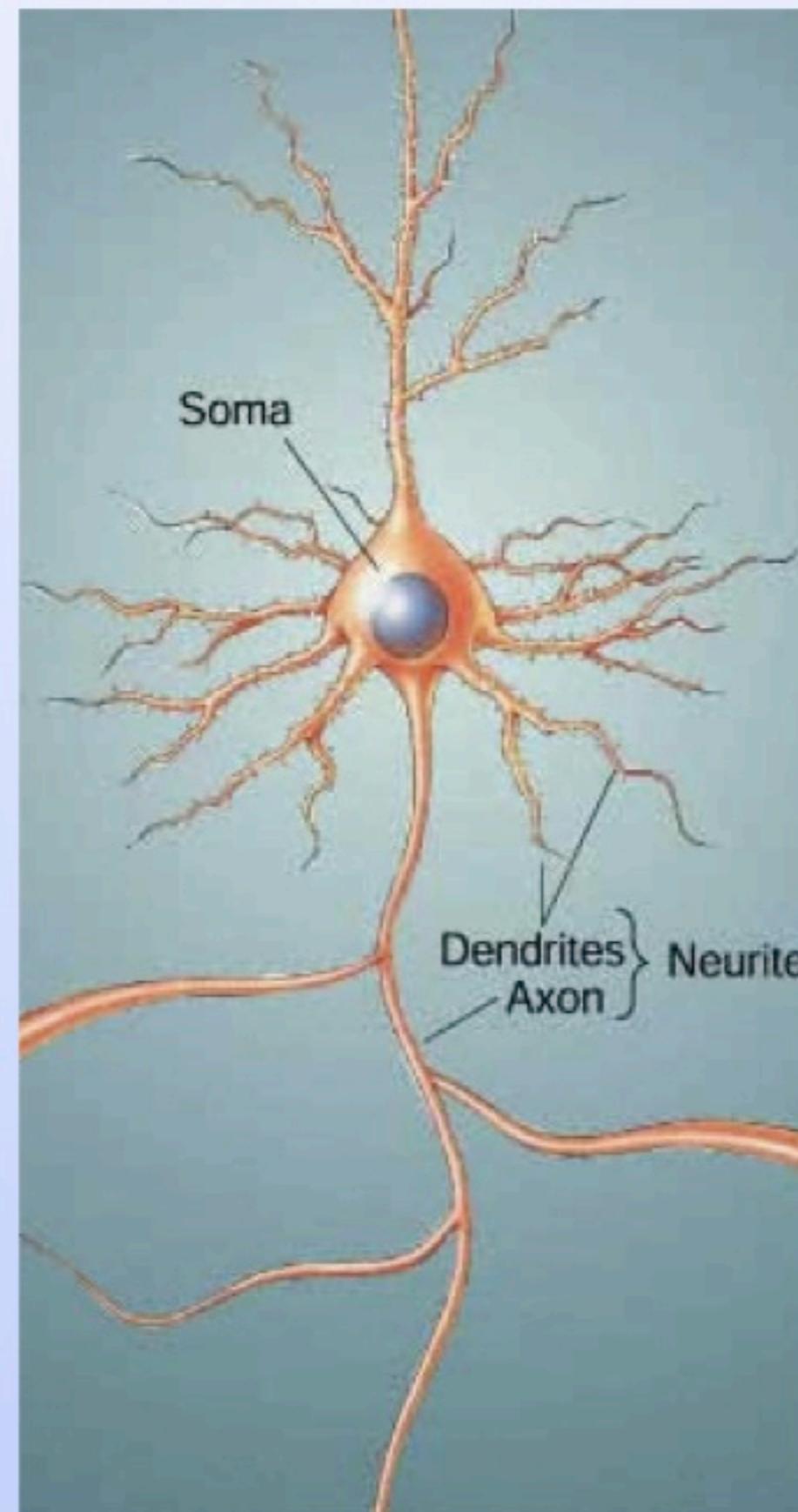
D'après Dotti et al. 1988

## Culture « à la Banker » dite « en sandwich » ou « glial feeder layer »



### 3. D'où proviennent les neurones ?

- Neurones et glie : neuroectoderm (progenitor stem cells)
- Neuroblastes -> neurones
- Astroblastes -> astrocytes



# 3. D'où proviennent les neurones ?

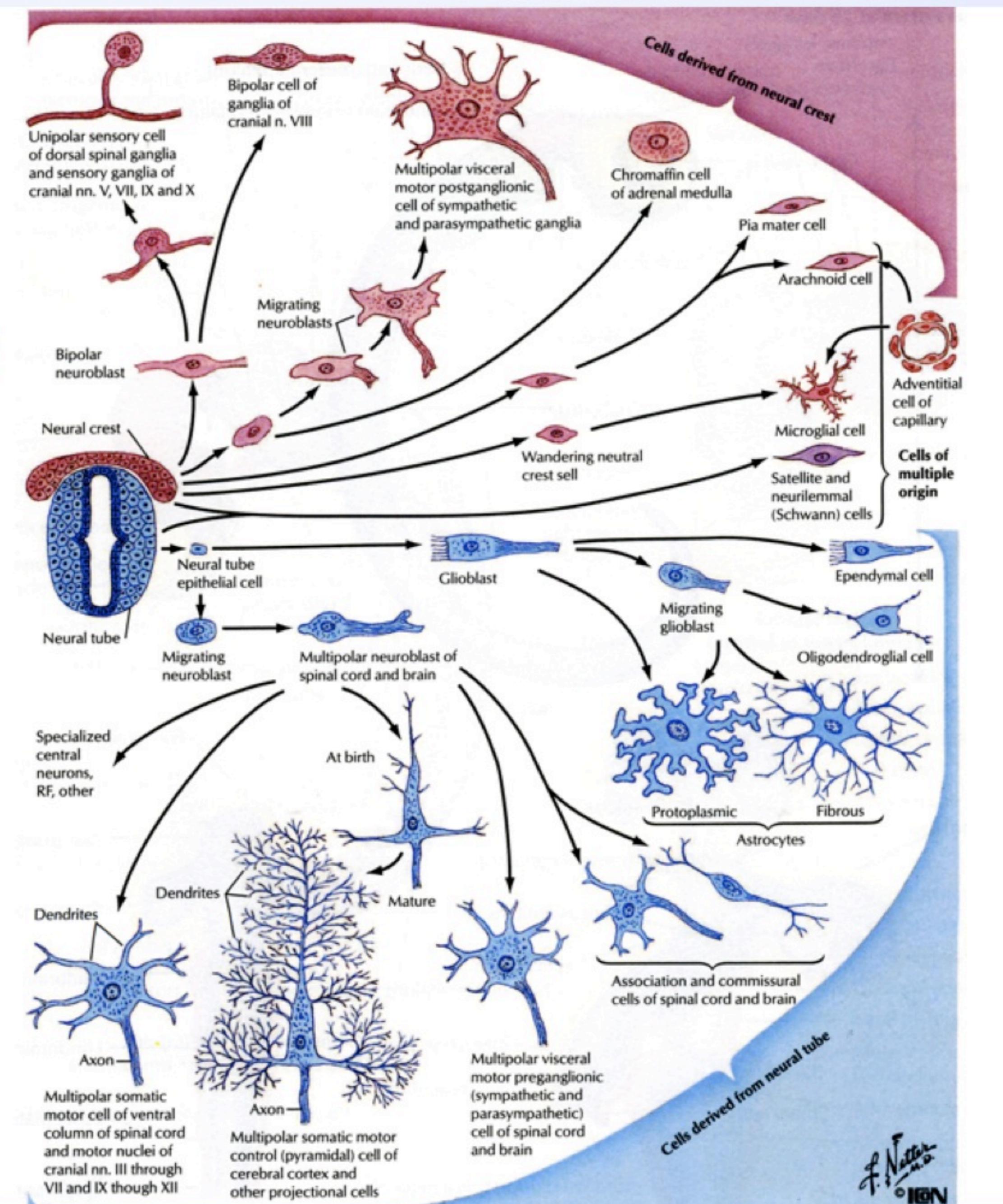
Derived cells from neural tube and neural crest

## Neural crest:

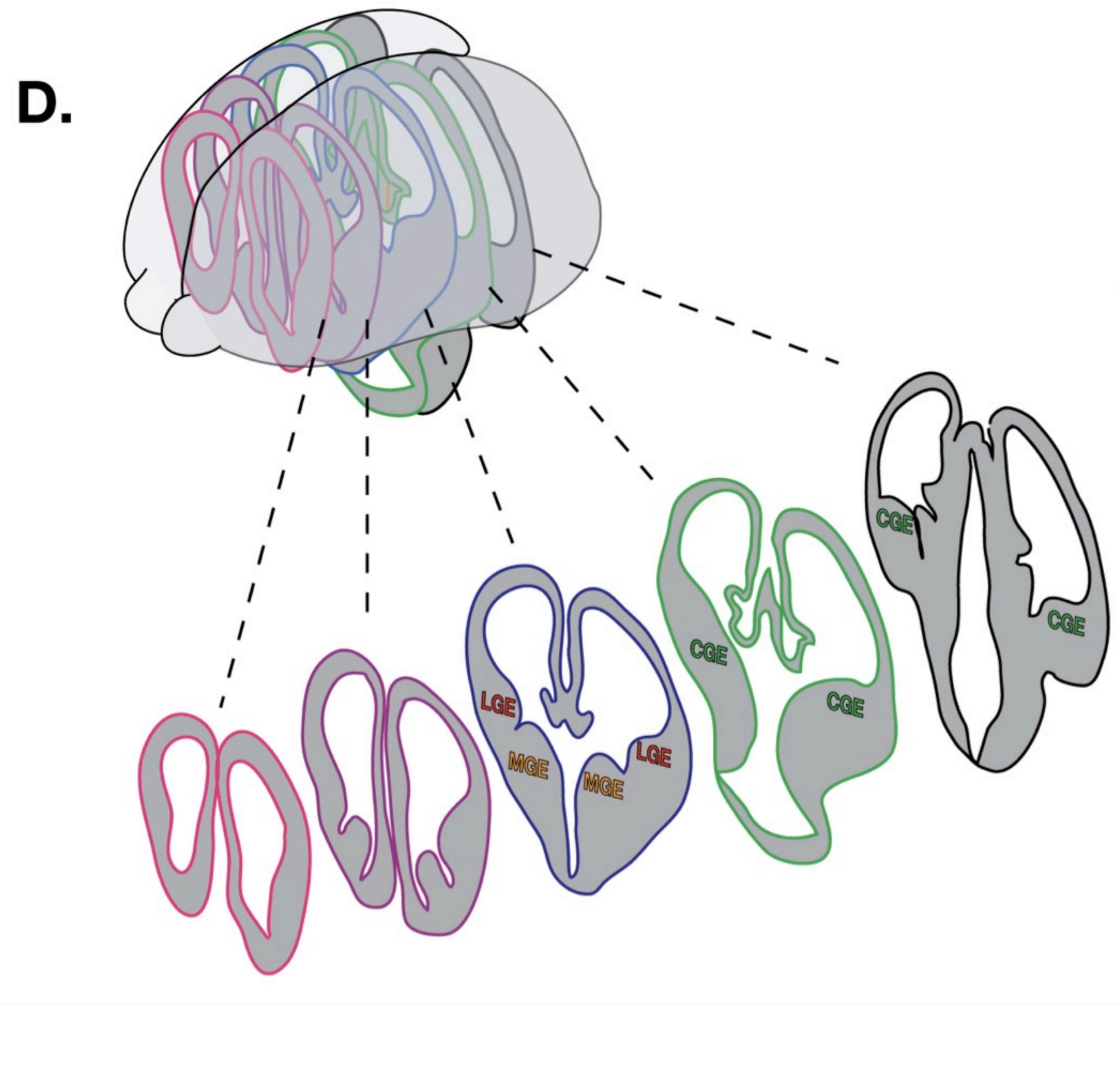
- Ganglionar cells (spinal & ANS)
- Chromaffin cells
- Glial cells:
  - Schwann cells
  - Microglia

## Neural tube:

- Neurons (brain & spinal cord)
- Ependymal cells
- Glial cells:
  - Oligodendrocytes
  - astrocytes



## Migration of neurons during development



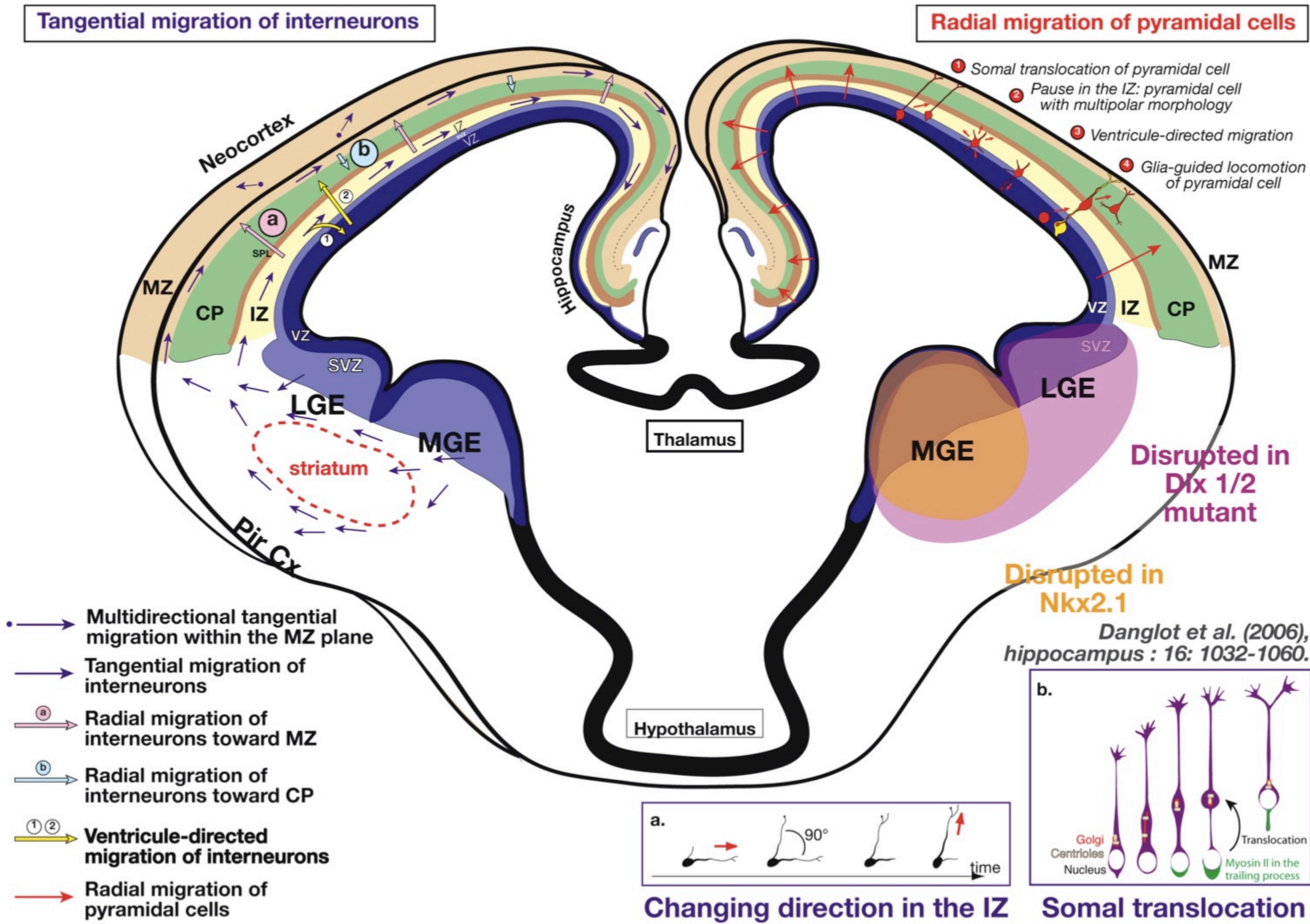
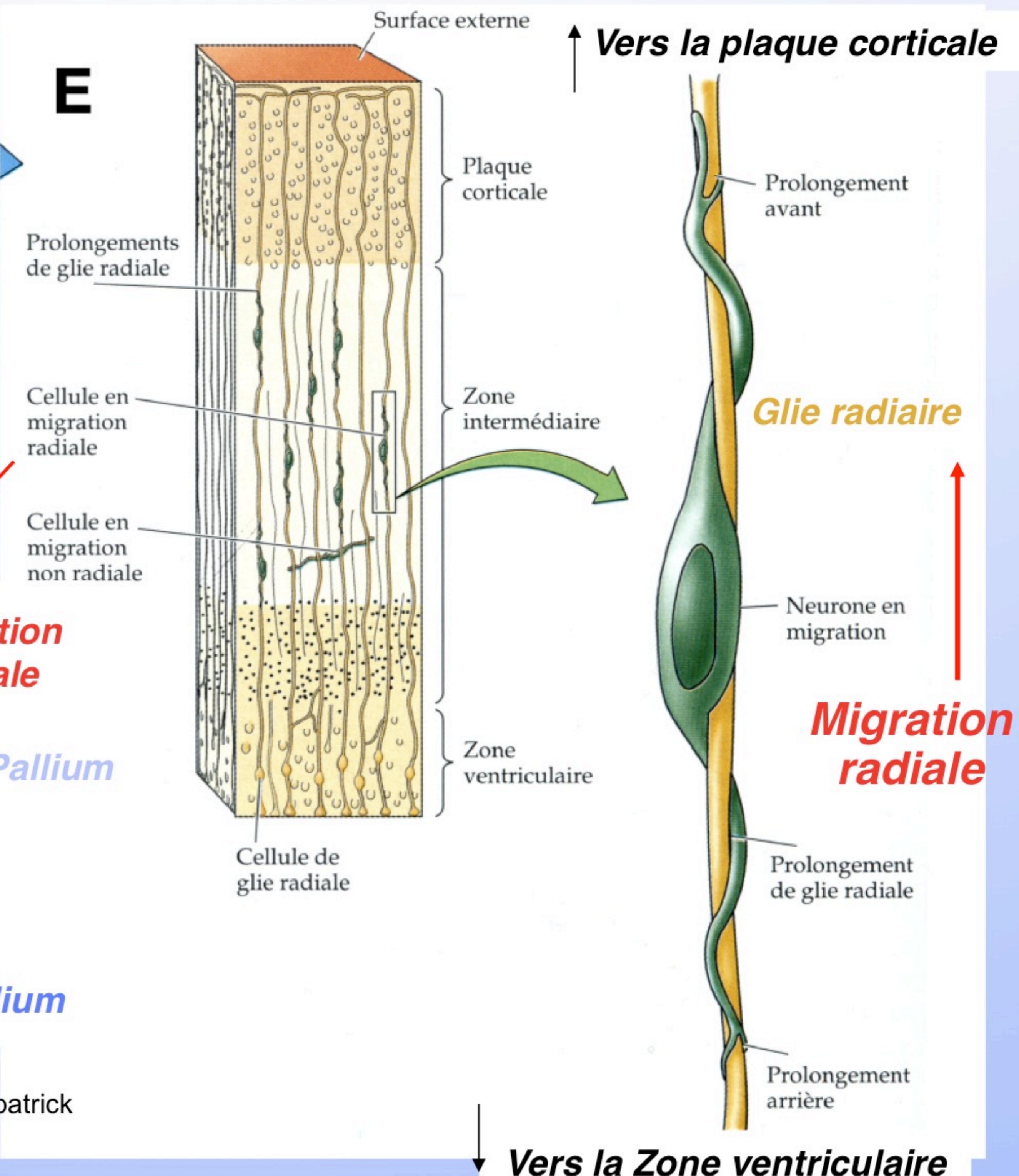
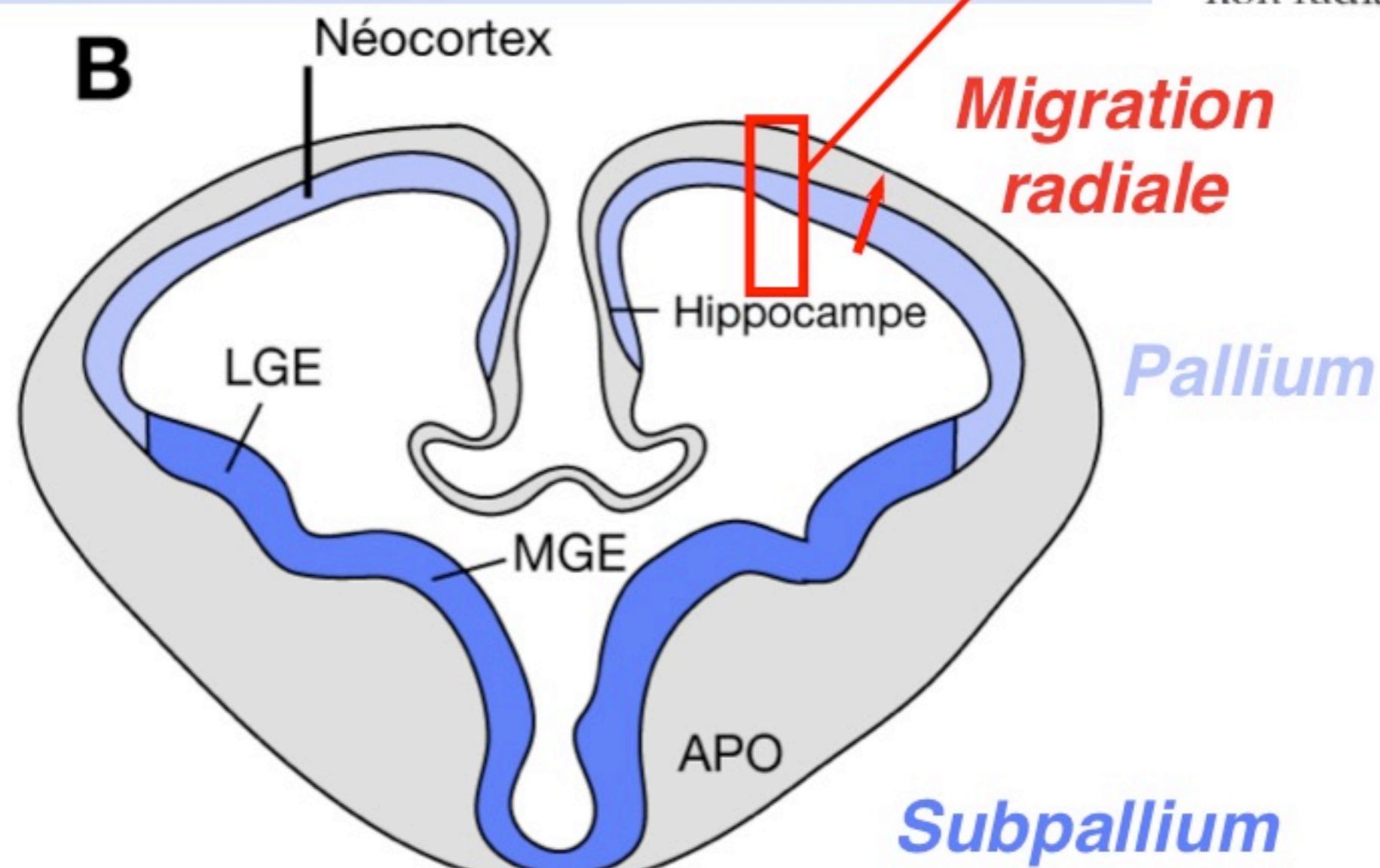
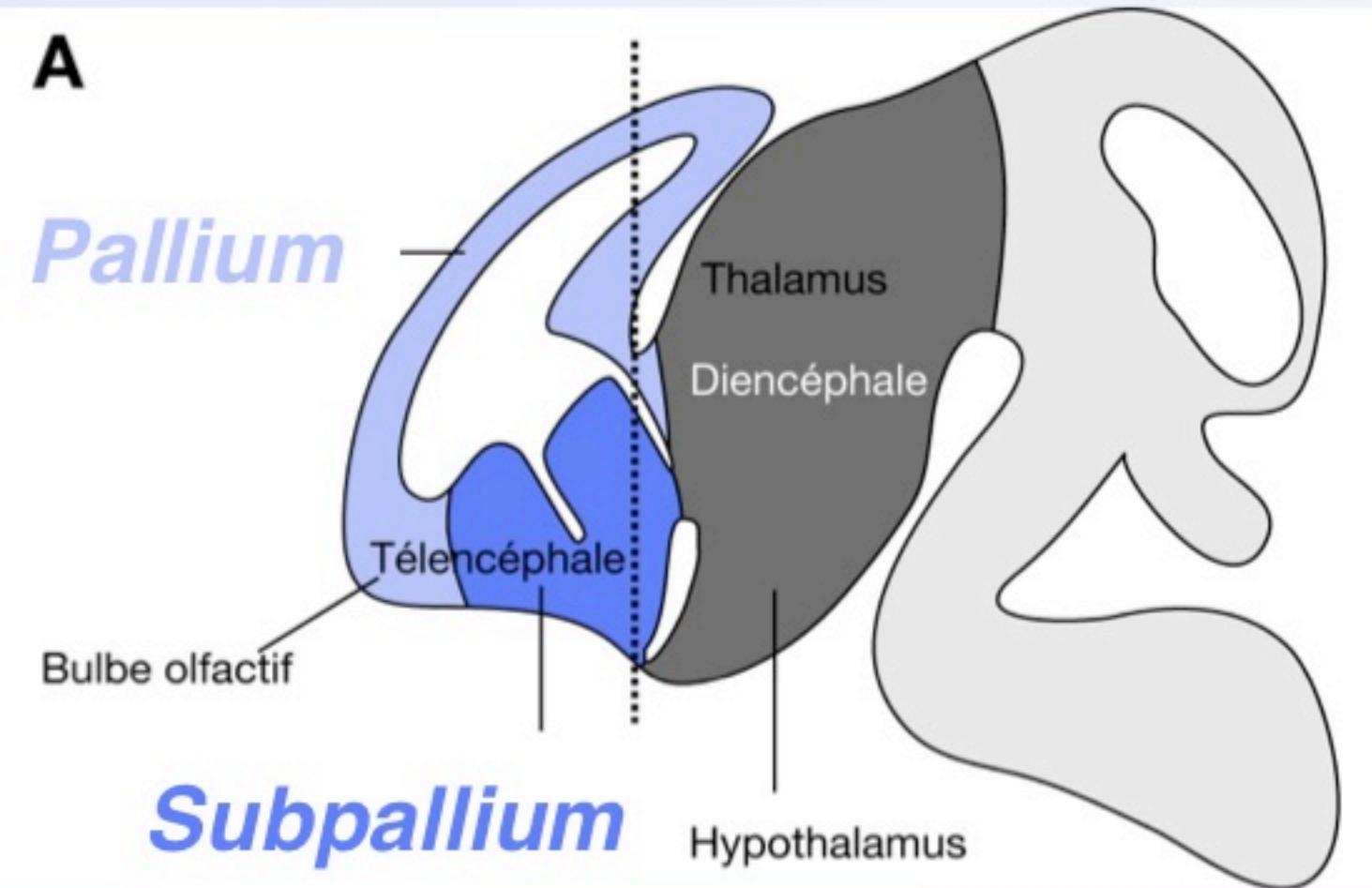


Figure 5 : Modes of migration of interneurons from the subpallial telencephalon toward the cortical and hippocampal anlagen.

## Télencéphale



Adapted from « Neurosciences »:

De Dale Purves, George J. Augustine, David Fitzpatrick

Traduit par Jean-Marie Coquery

De Boeck Université 3ème édition, 2005

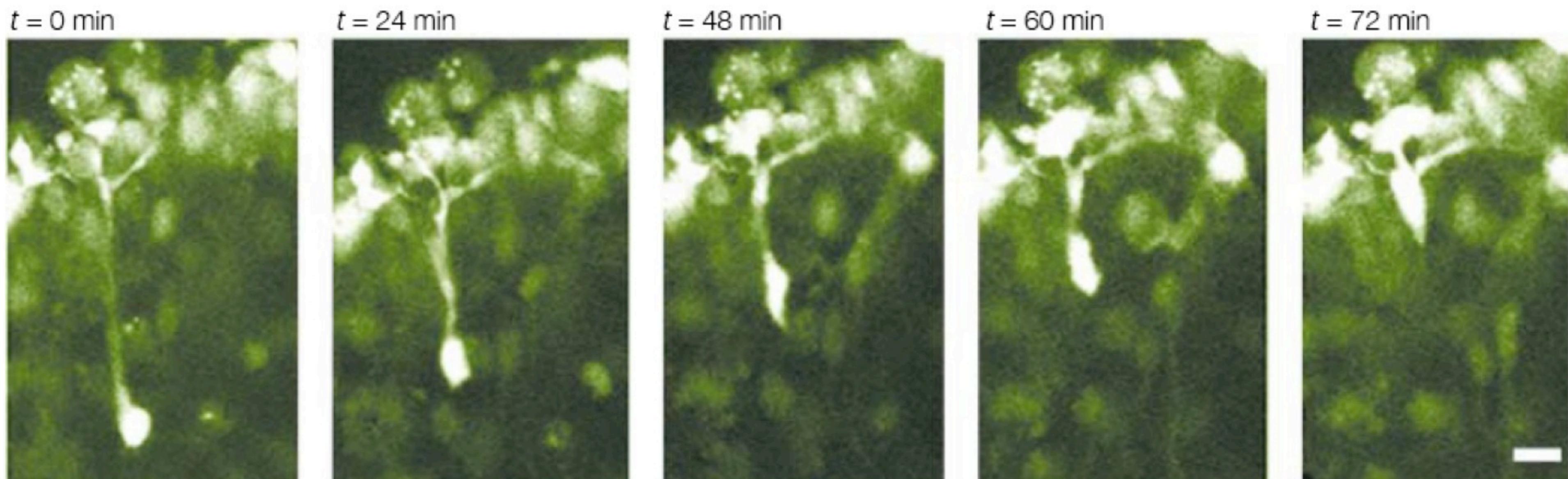
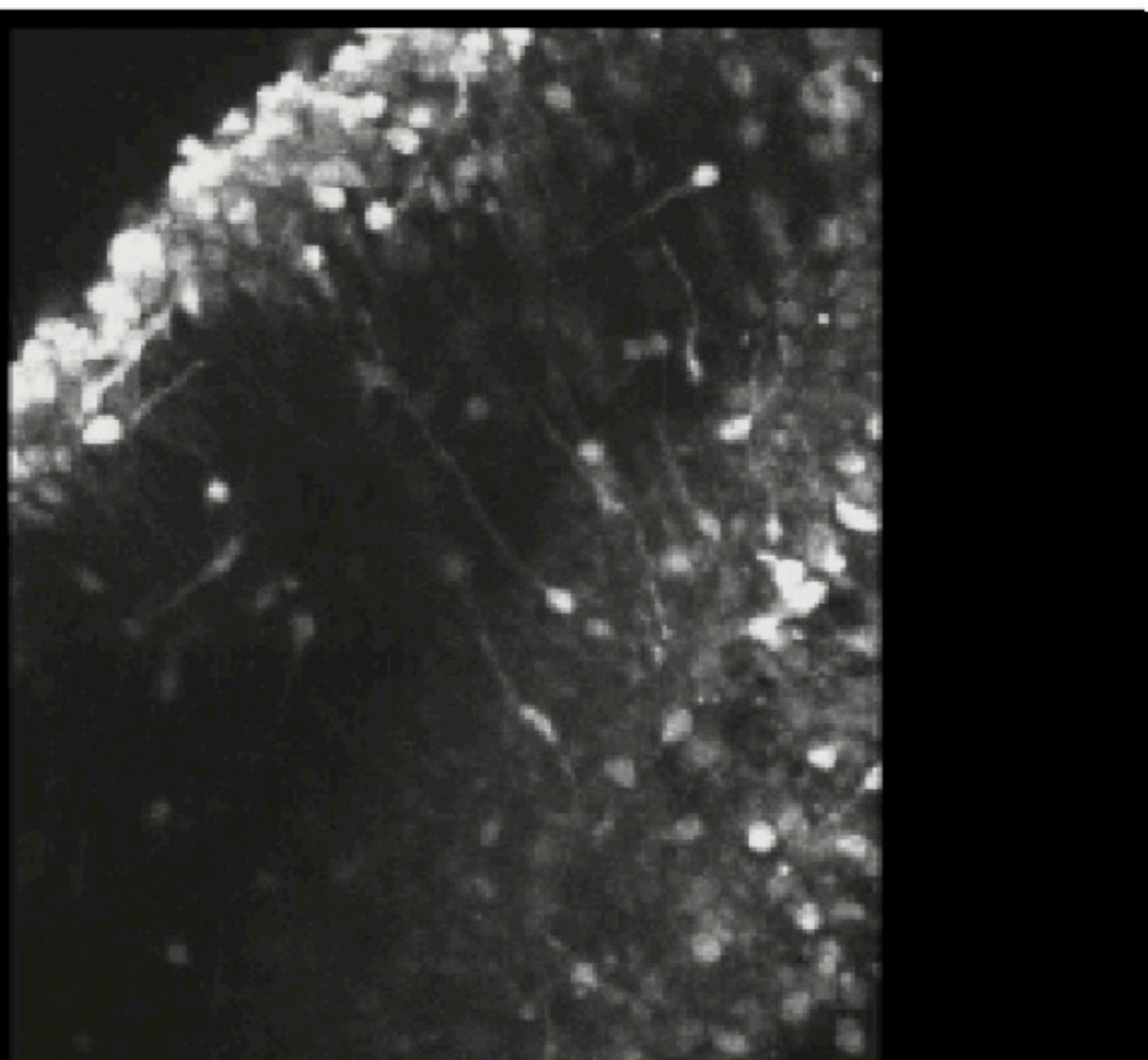


Figure 3 | **Somal translocation.** Time-lapse images of a cell showing somal translocation in a mouse cortical slice that was labelled with Oregon Green BAPTA-1 488 AM. Images were acquired every minute and each frame shows a single optical section. Scale bar,  $10 \mu\text{m}$ . See [Supplementary Movie](#) from REF.31 © 2001 Macmillan Magazines Ltd.



Nadarajah & Parnavelas  
Nat Rev Neur (2002) vol.3:423.

Somal translocation  
E16 - 300min

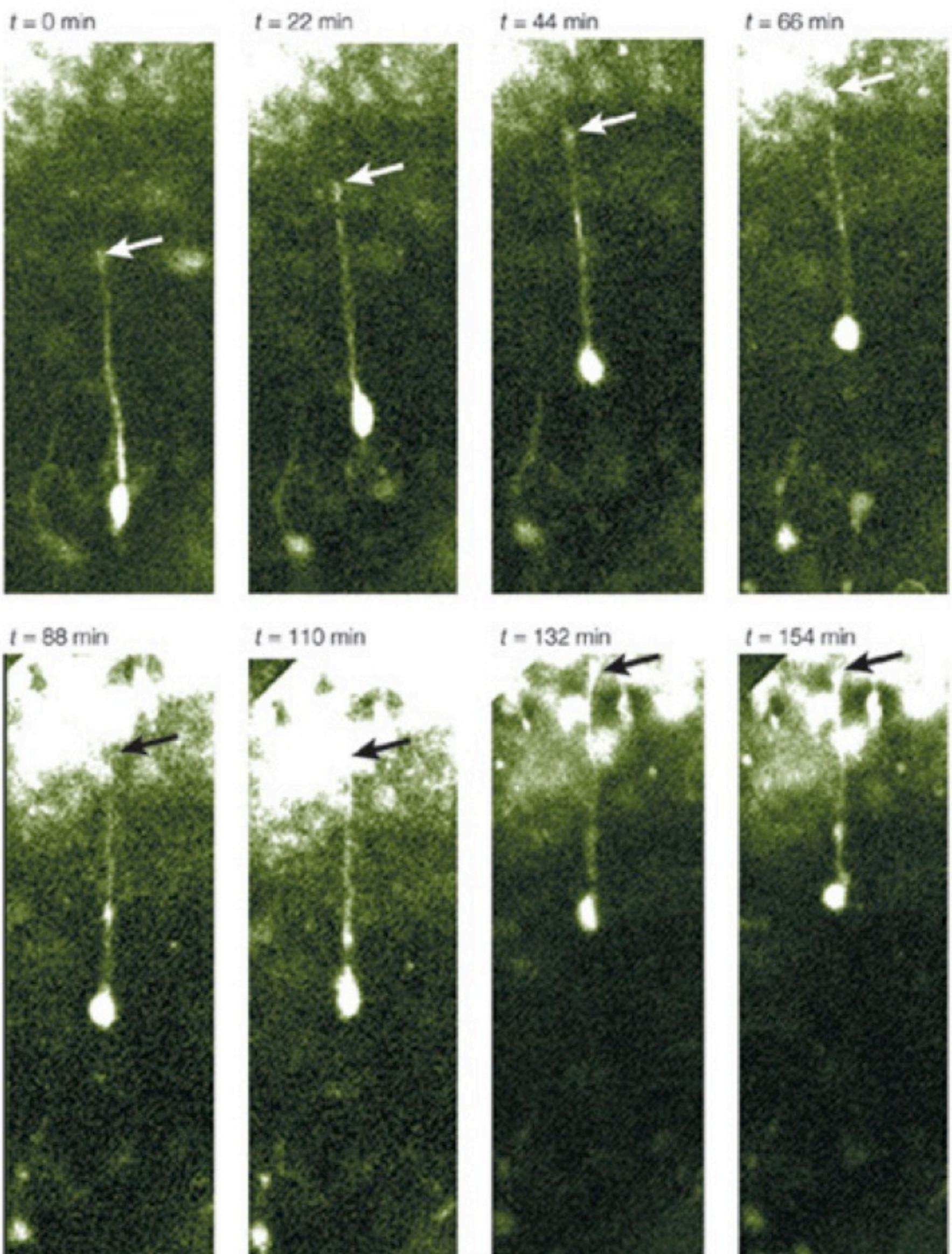
## Glia-guided locomotion

**Glial-Guidance**

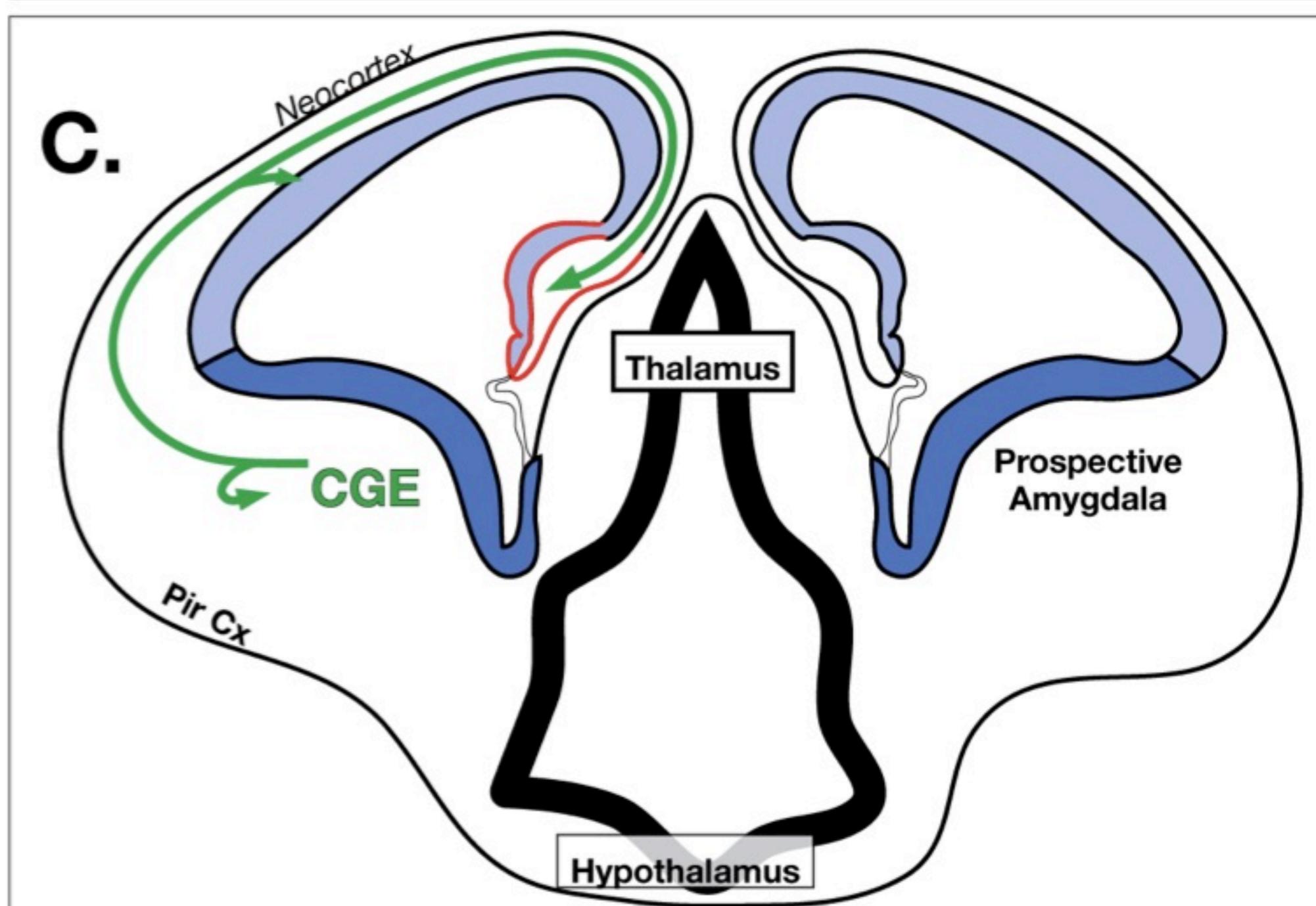
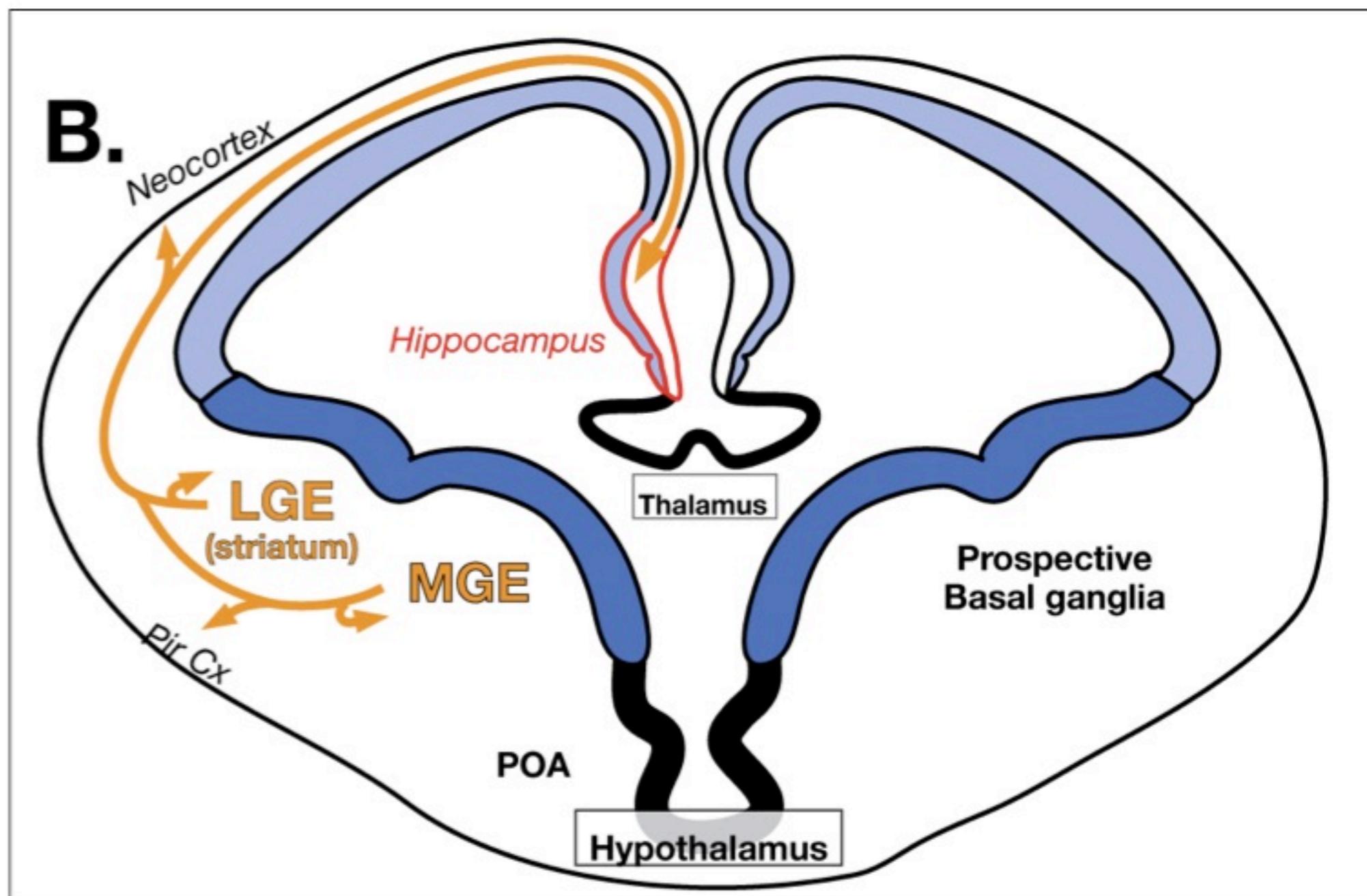
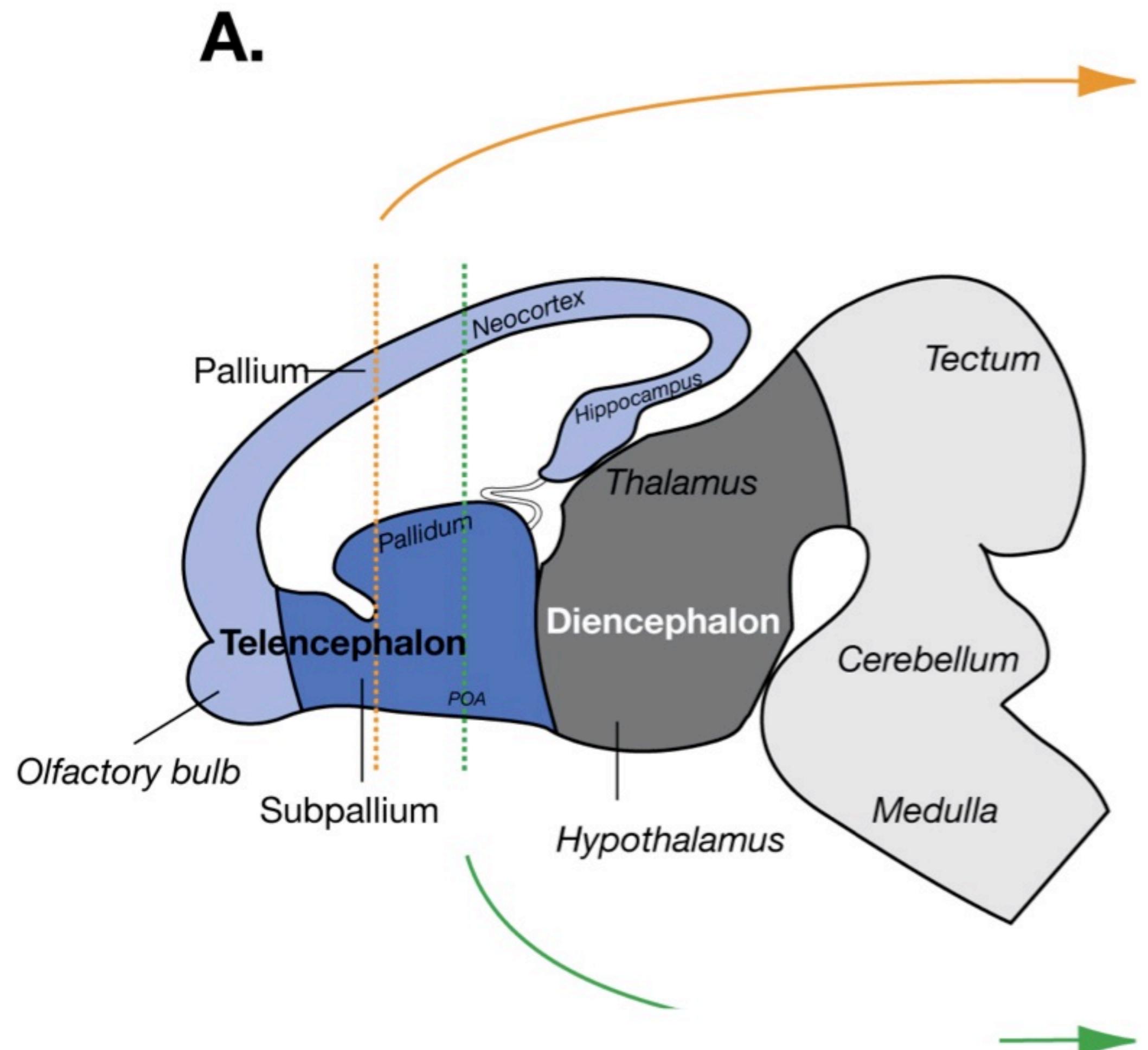
**recording time:160 min**

Nadarajah, Nature Neurosci. 4, 143–150 (2001).

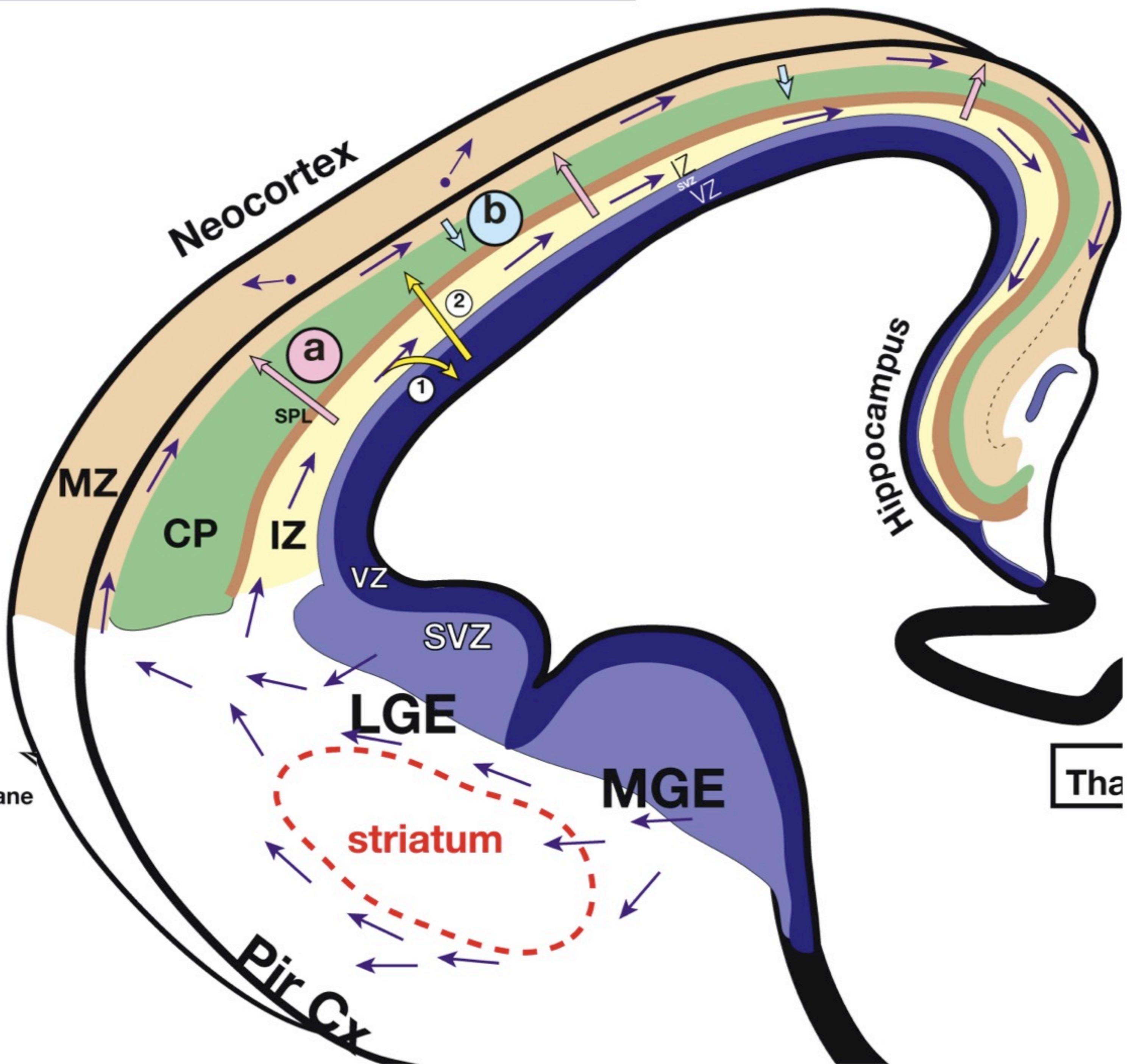
Nadarajah & Parnavelas  
Nat Rev Neur (2002) vol.3:423.



## Migration of inhibitory interneurons



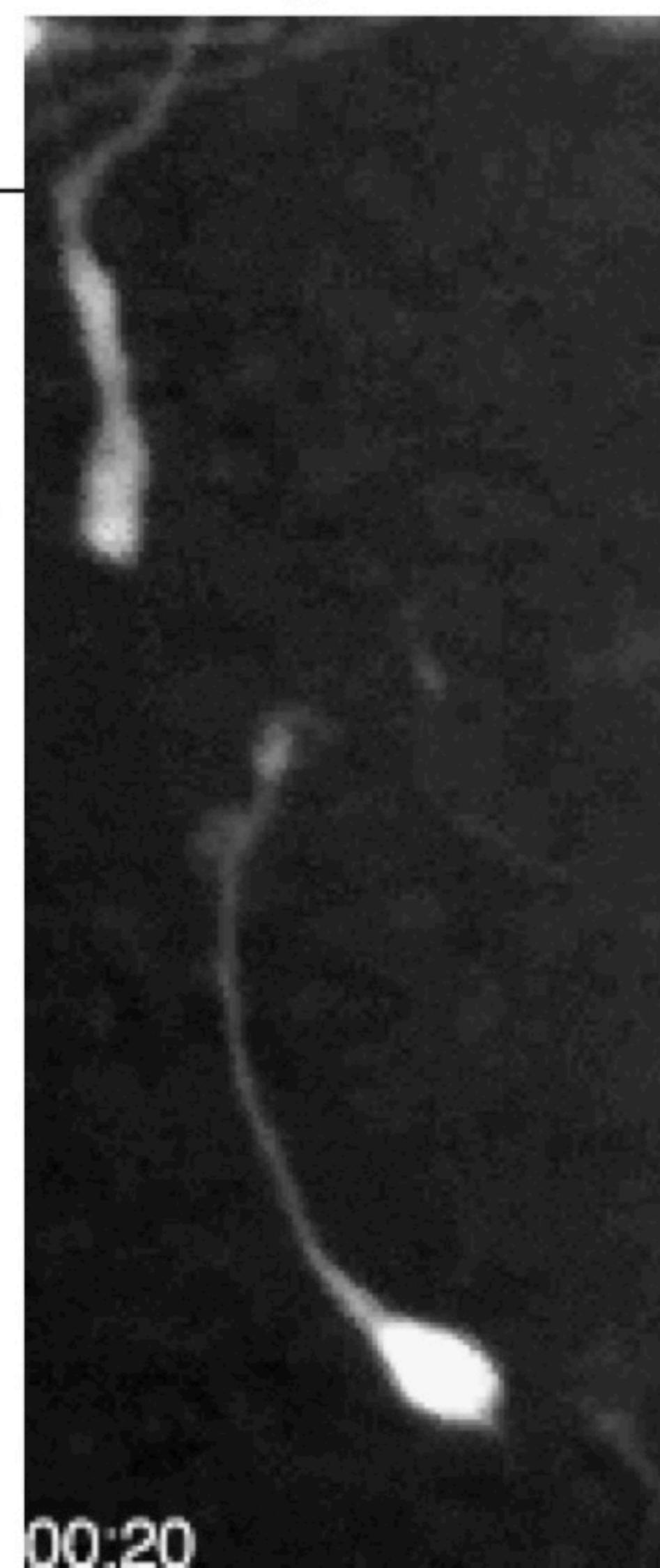
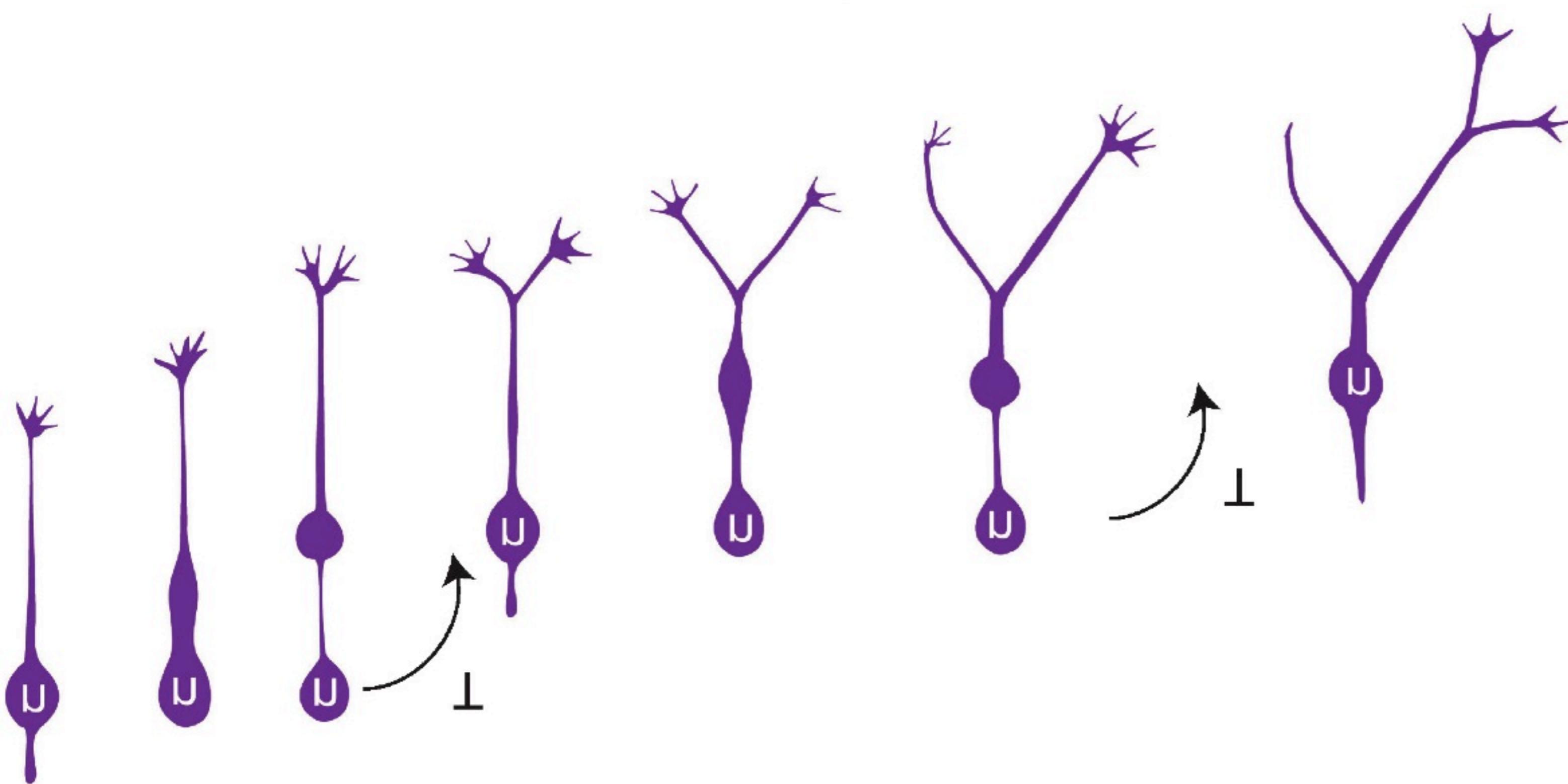
## Tangential migration of interneurons



## REVIEW ARTICLE

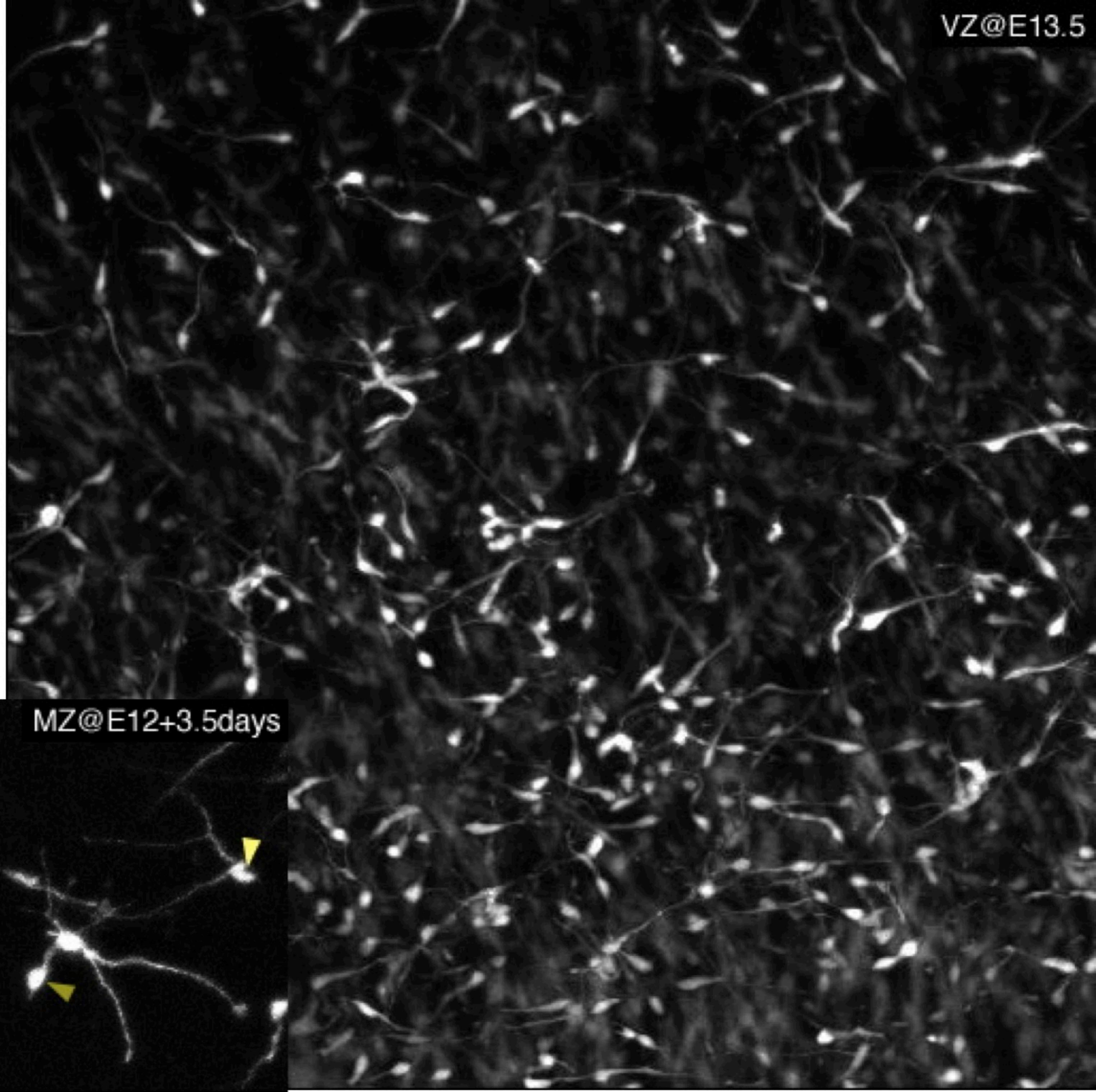
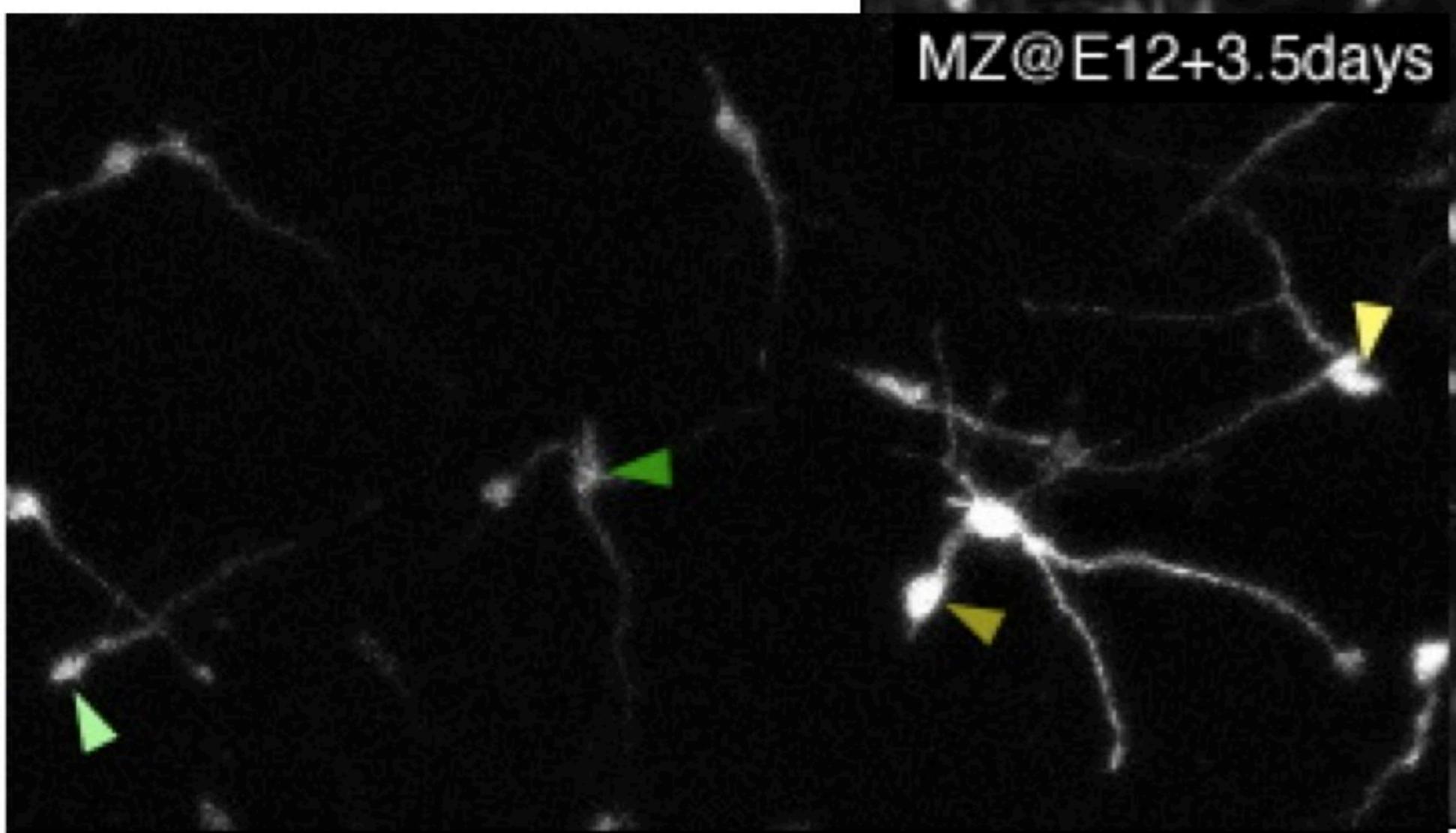
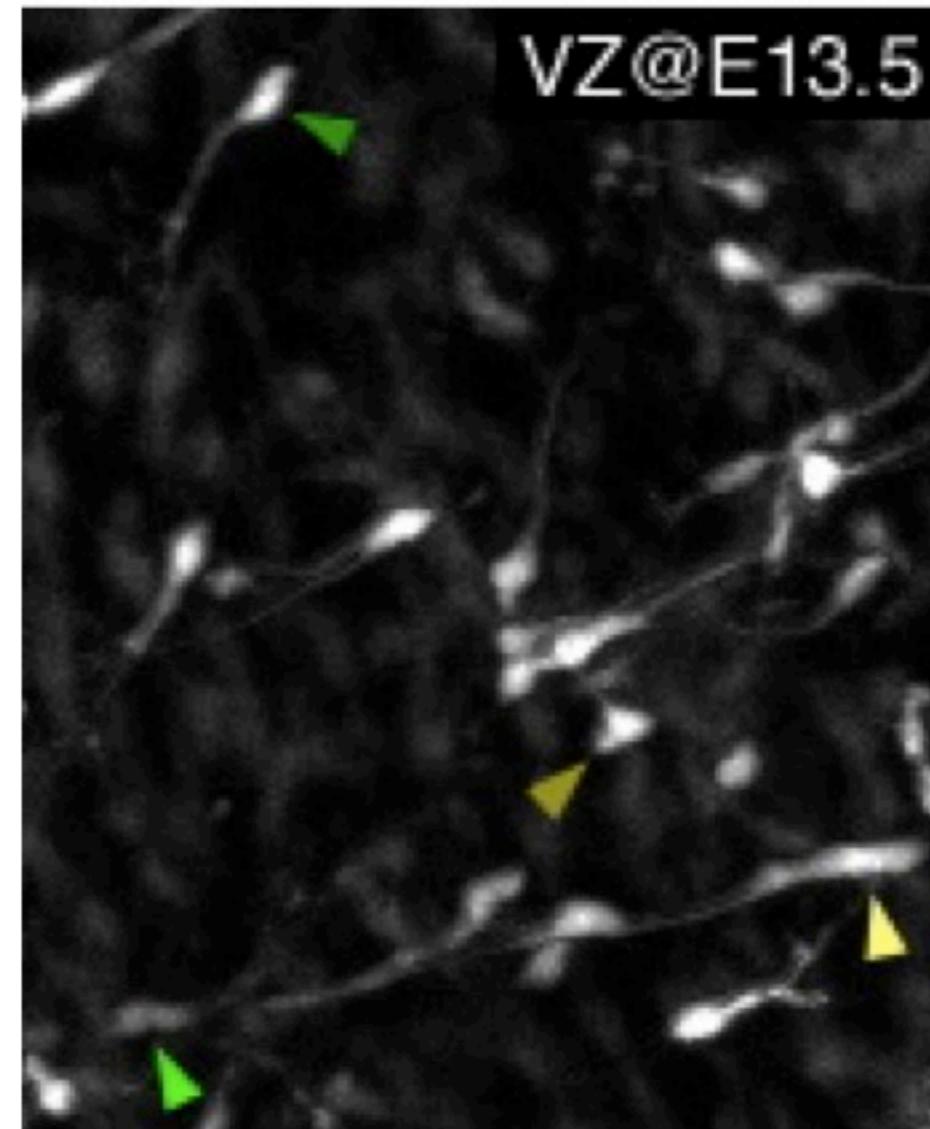
# Cell and molecular mechanisms involved in the migration of cortical interneurons

Christine Métin,<sup>1,2</sup> Jean-Pierre Baudoin,<sup>1,2</sup> Sonja Rakic<sup>3</sup> and John G. Parnavelas<sup>3</sup>

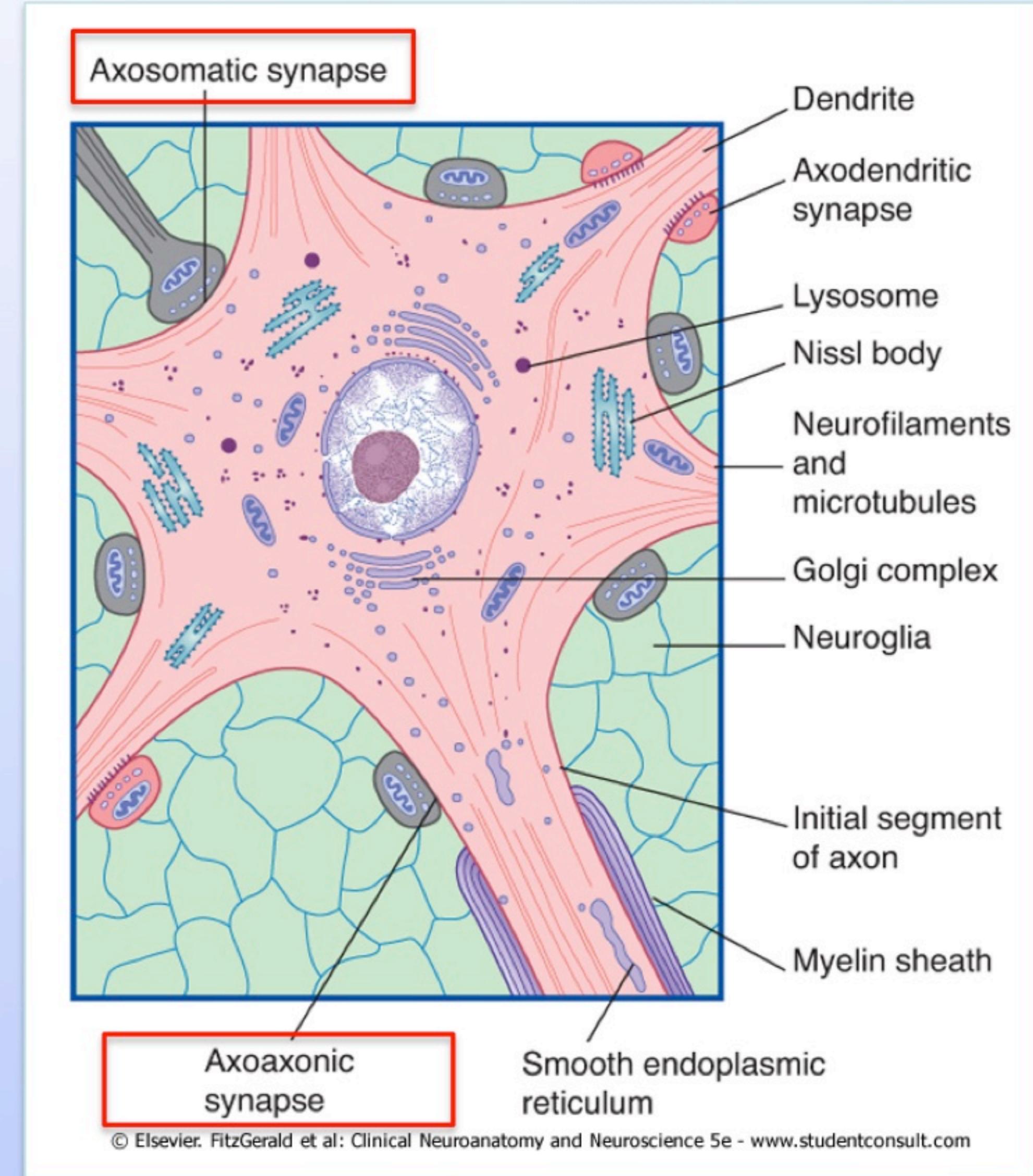
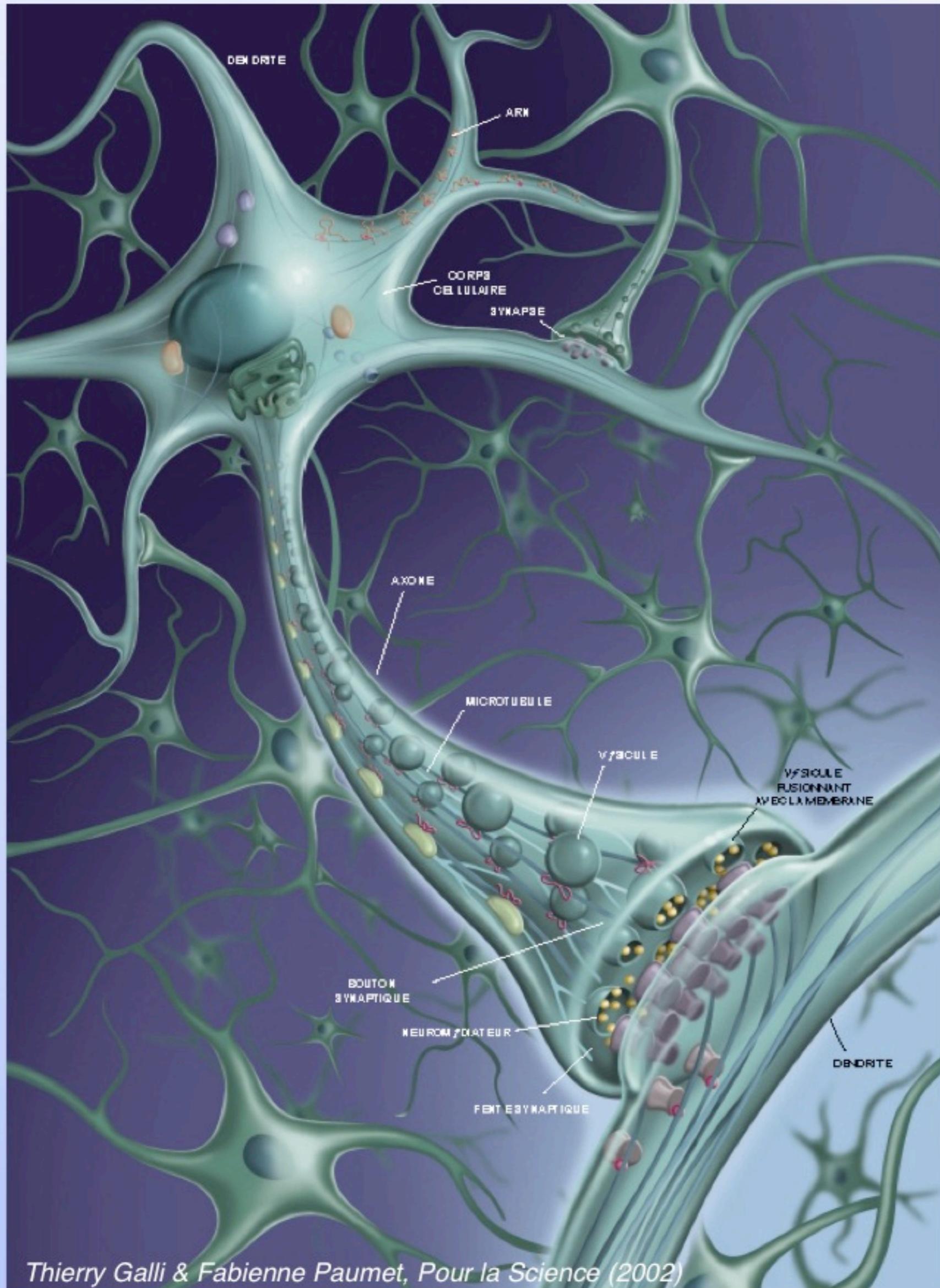


Multidirectional and  
multizonal tangential  
migration of GABAergic  
interneurons in the  
developing cerebral cortex  
Development 133, 2167-2176 (2006)

VZ@E13.5

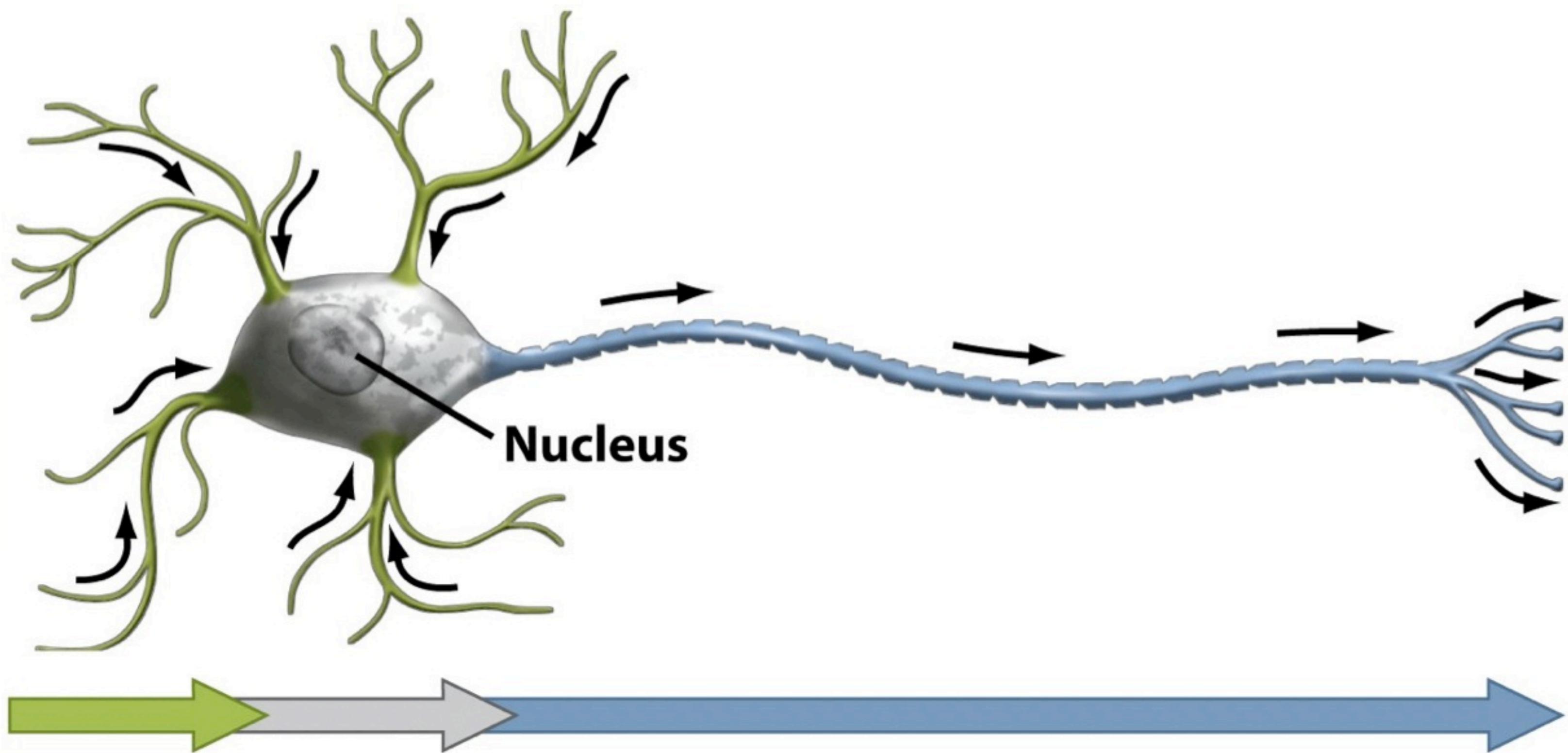


# 4. Polarité neuronale : les domaines



# La signalisation neuronale nécessite la polarité

## Information flow through neurons

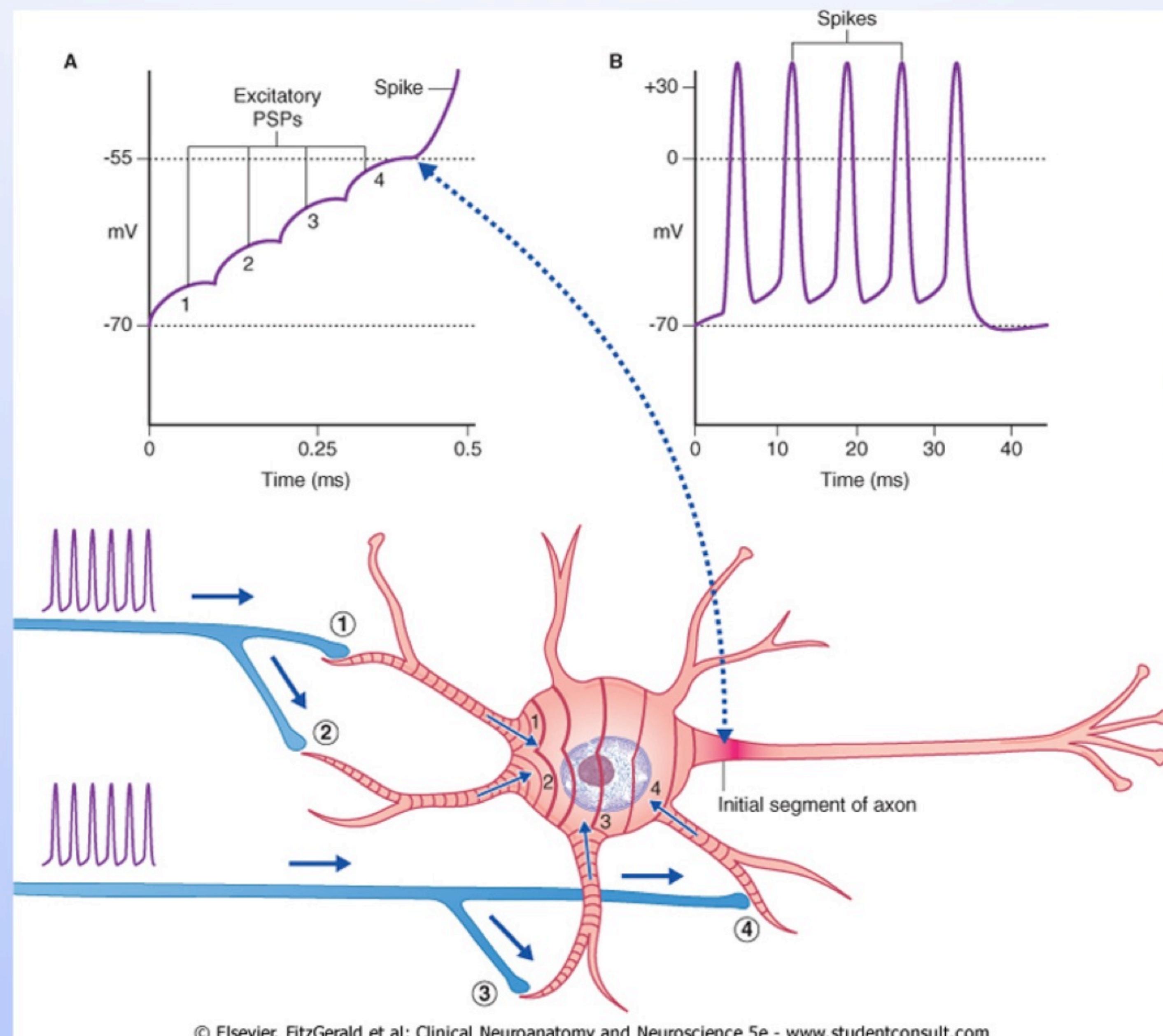


**Dendrites**  
Collect  
electrical  
signals

**Cell body**  
Integrates incoming  
signals and generates  
outgoing signal to  
axon

**Axon**  
Passes electrical signals  
to dendrites of another  
cell or to an effector cell

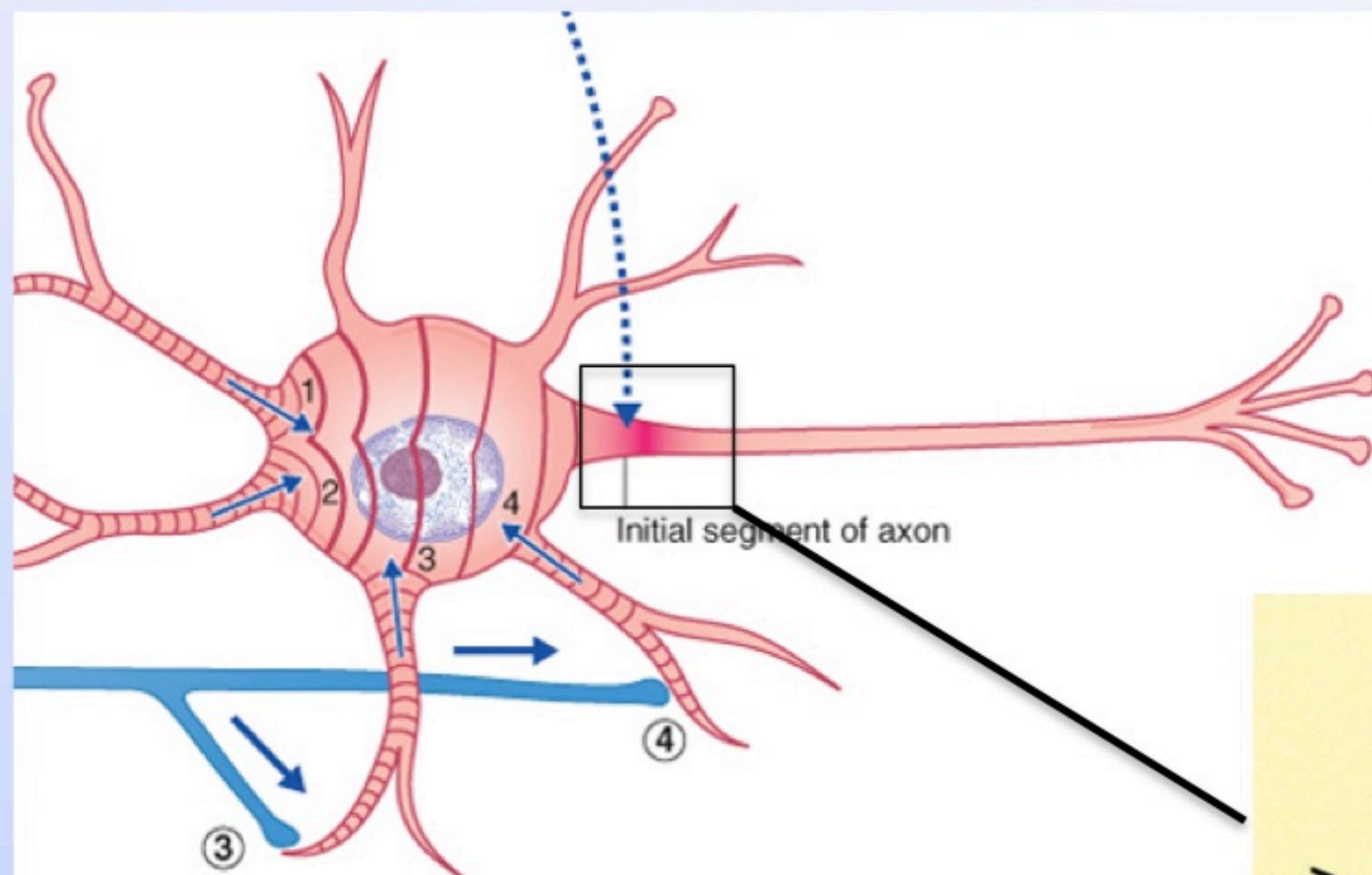
# La signalisation neuronale nécessite la polarité



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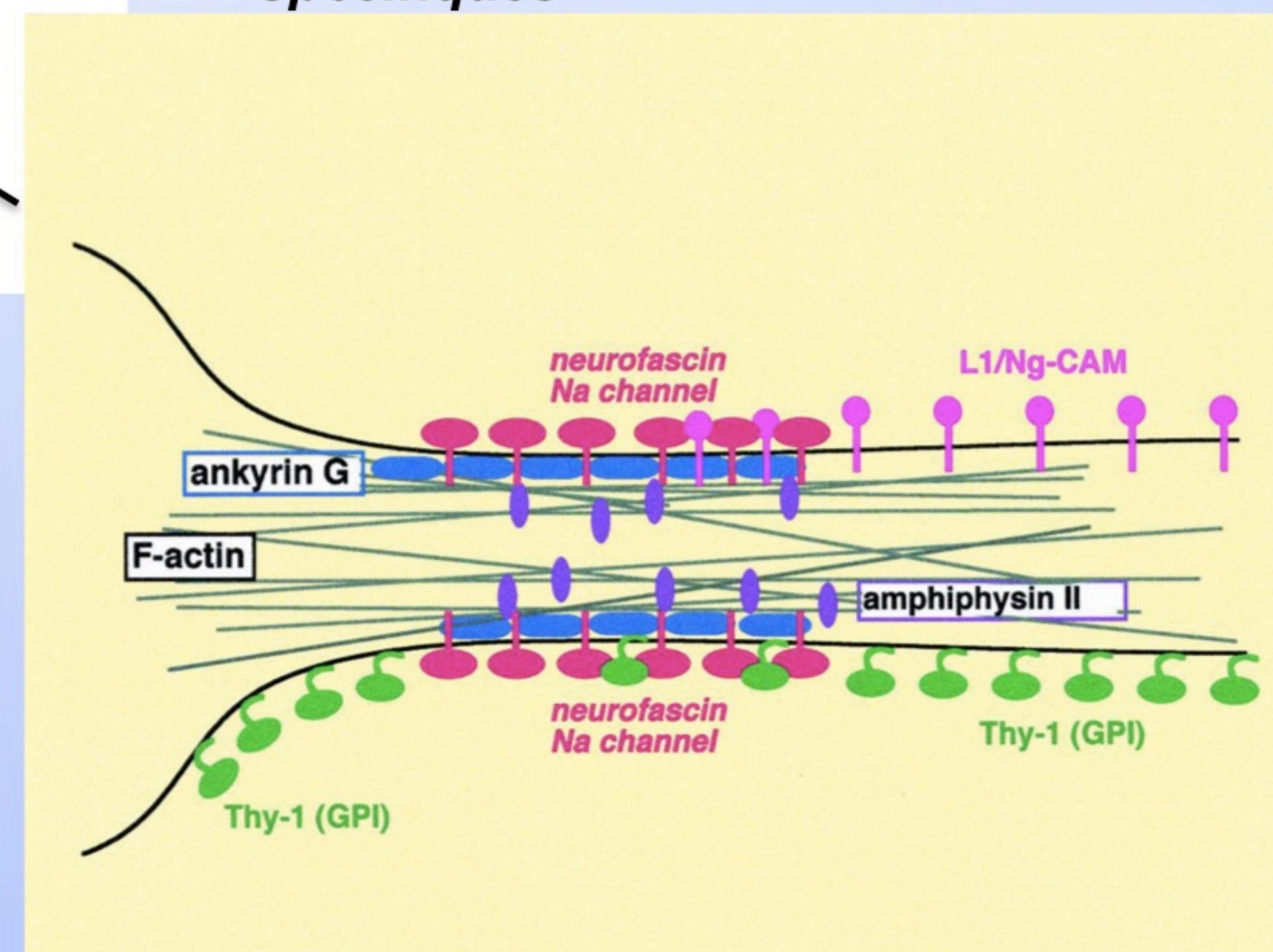
©Elsevier.Fitz Gerald et al., Clinical Neuronantomy and Neuroscience  
www.studentconsult.com

# 4. Les domaines: segment initial de l'axone

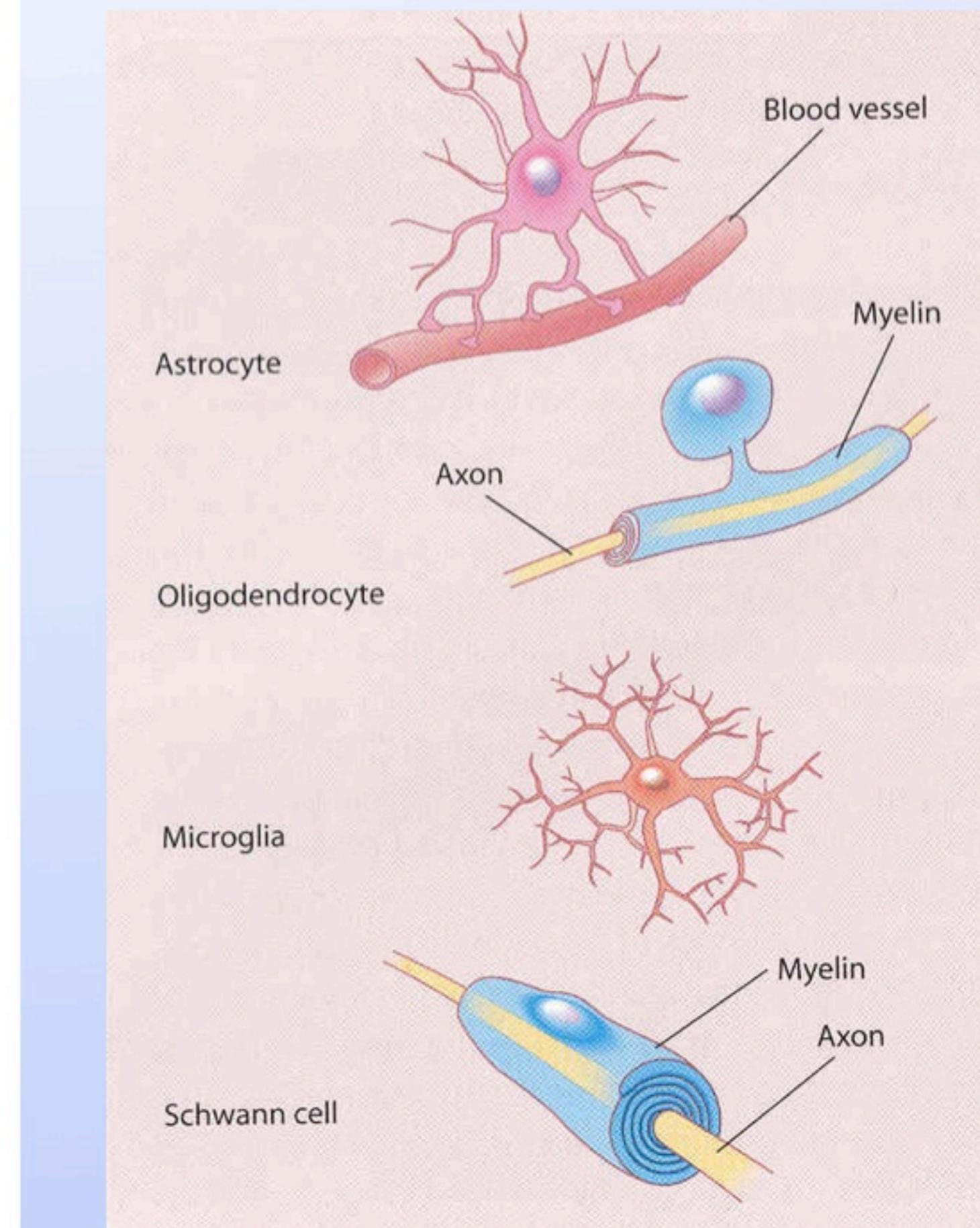
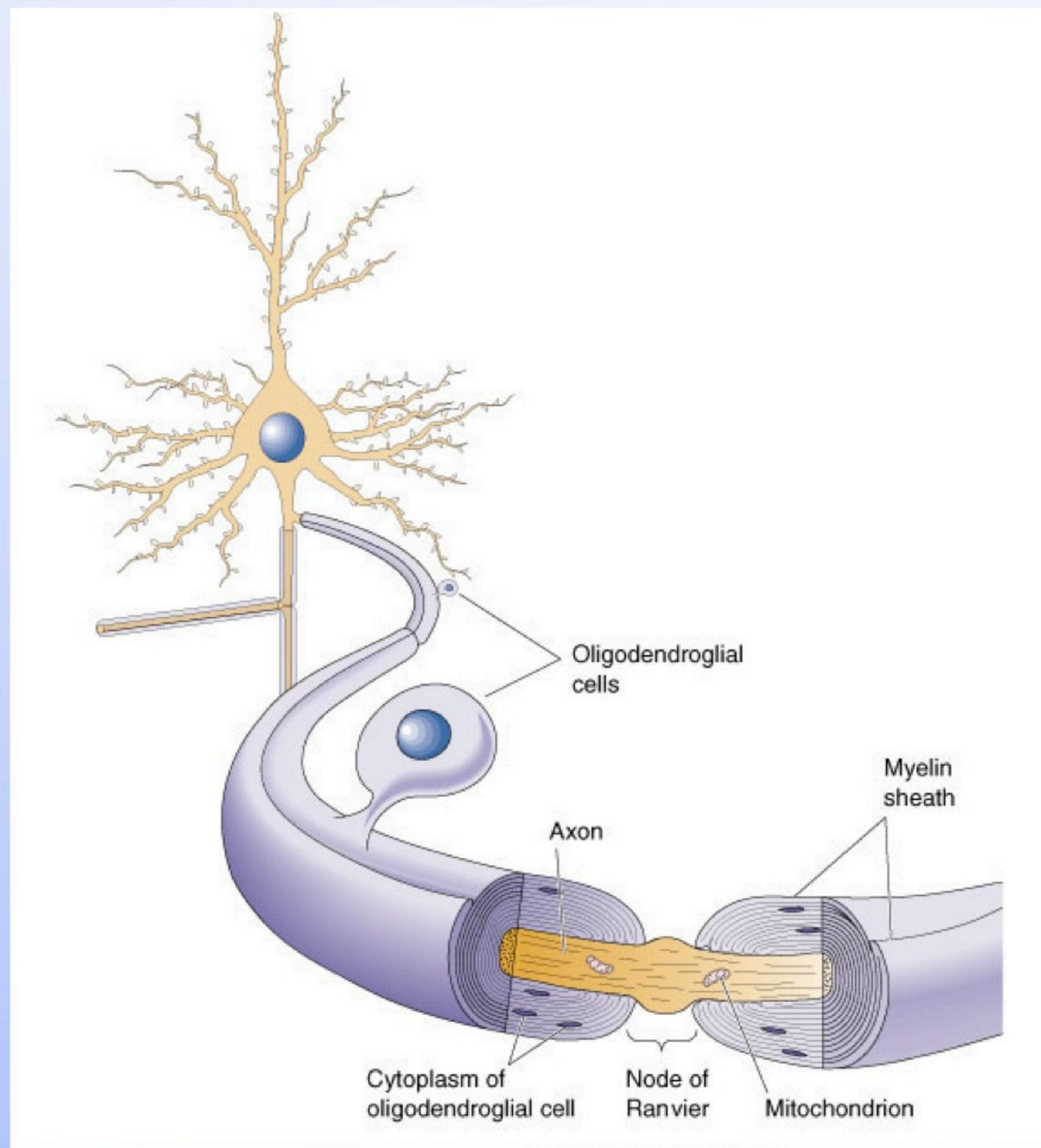


I et al: Clinical Neuroanatomy and Neuroscience 5e - www.studentconsult.com

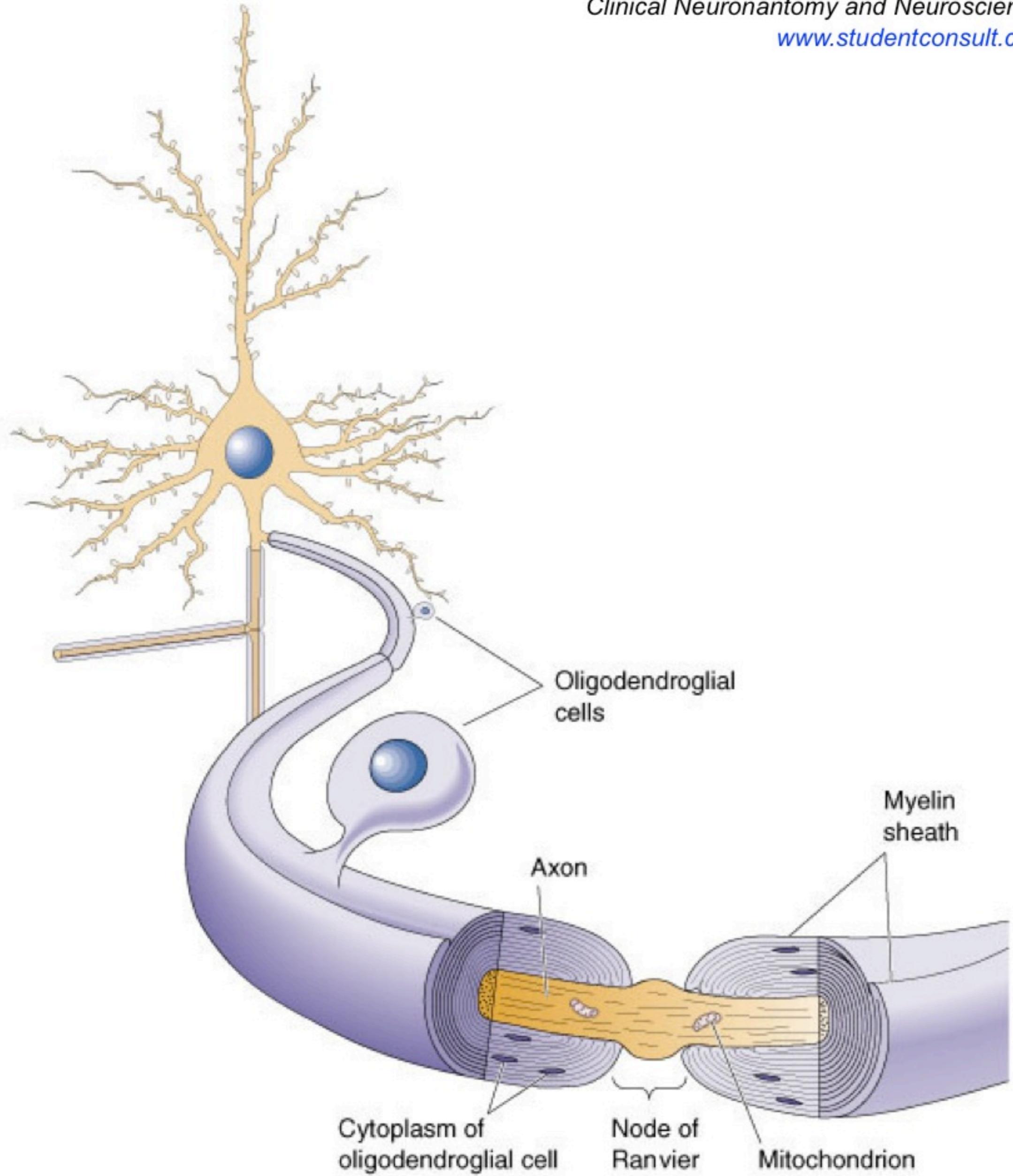
## Expression et ciblage de protéines spécifiques



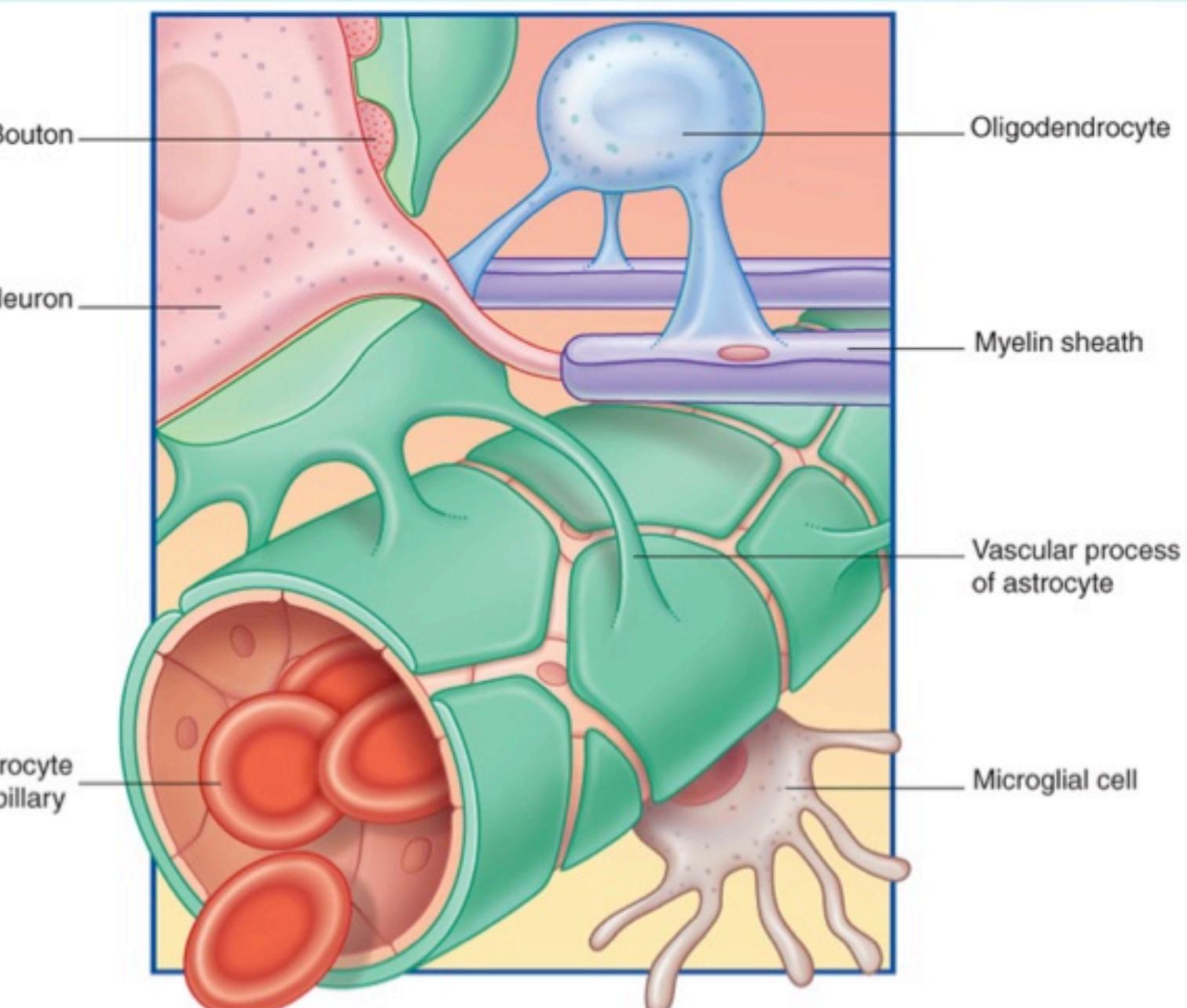
# Oligodendrocytes and the formation of myelin



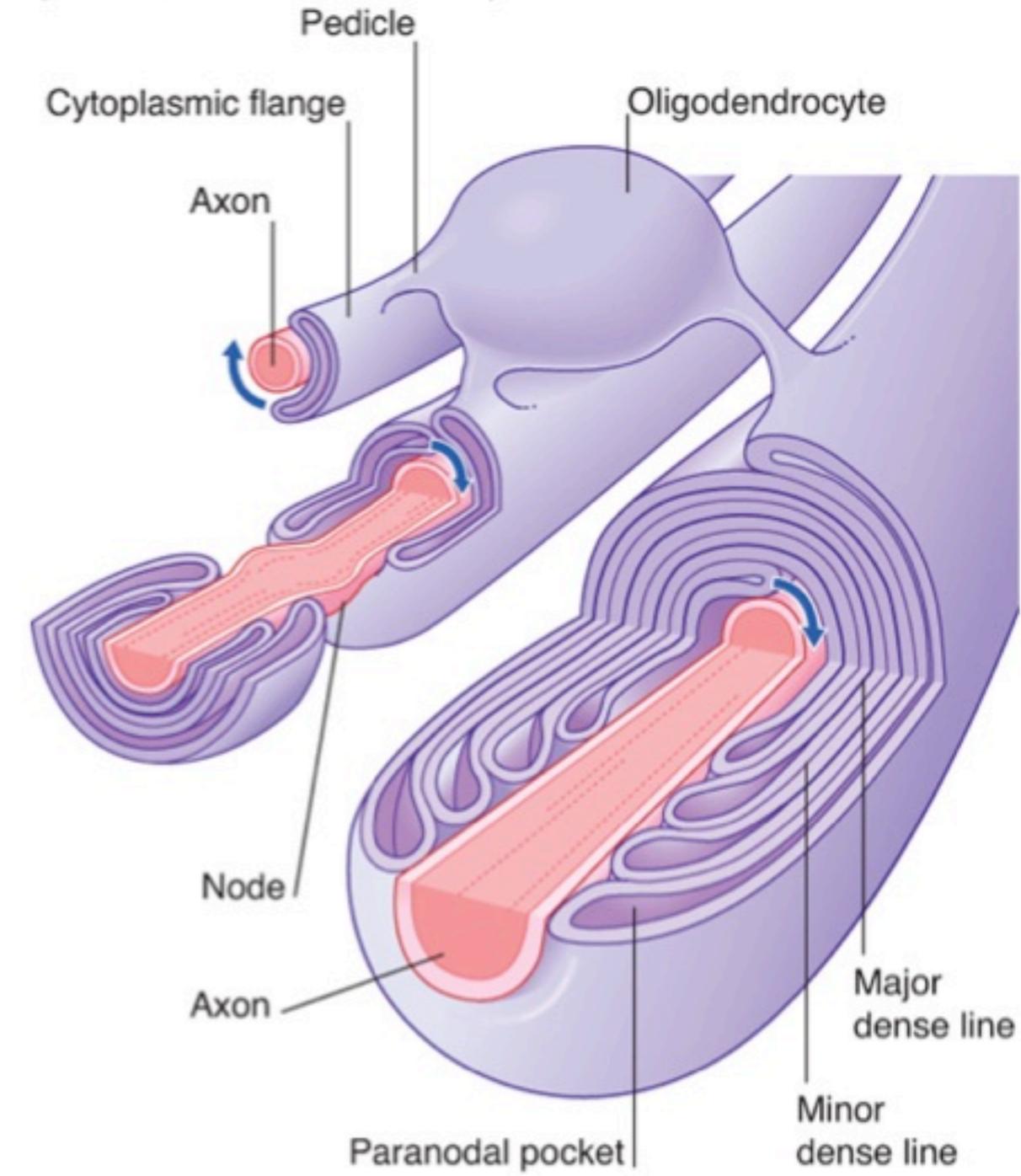
# Oligodendrocytes and the formation of myelin



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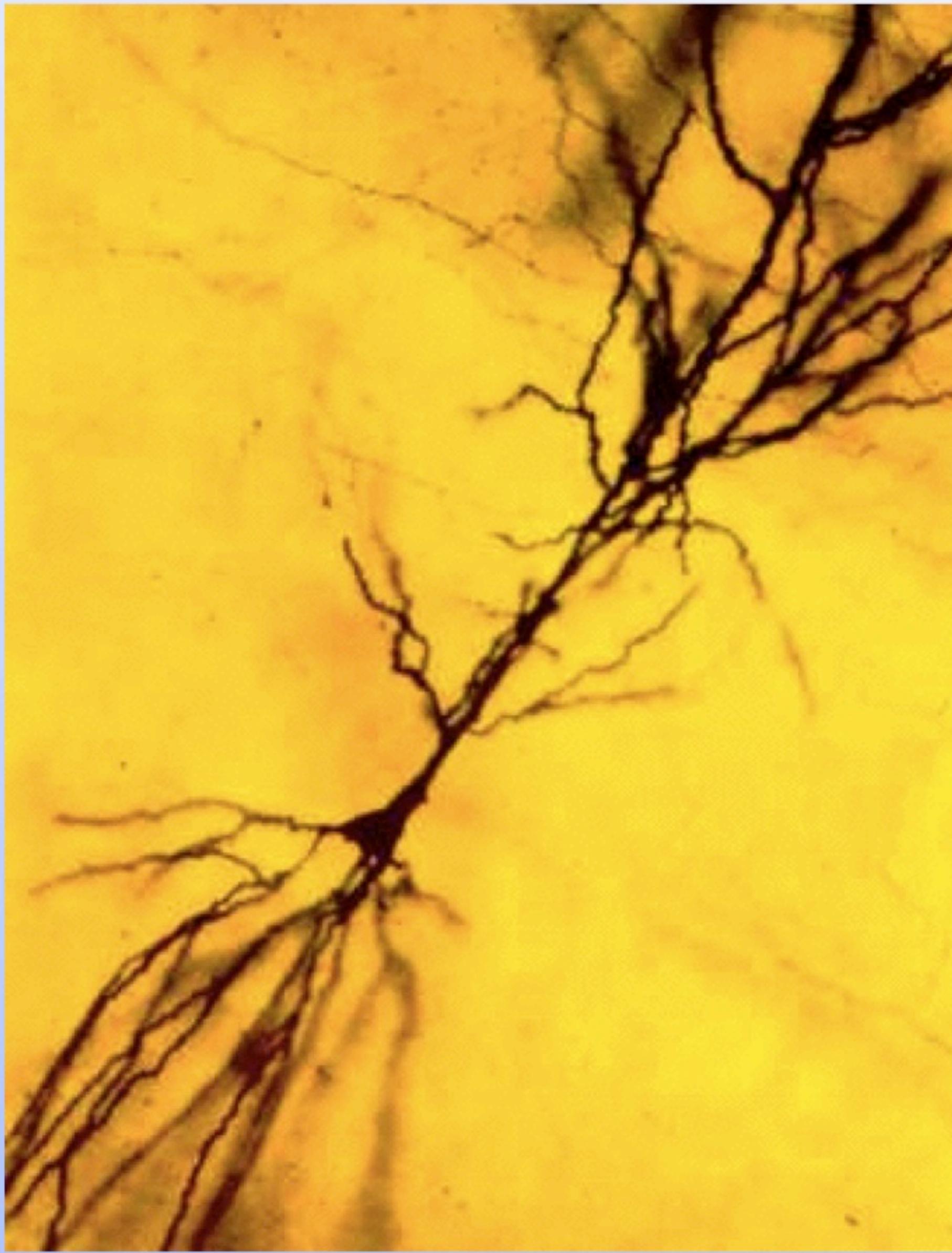


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# PYRAMIDAL NEURON (HIPPOCAMPAL)

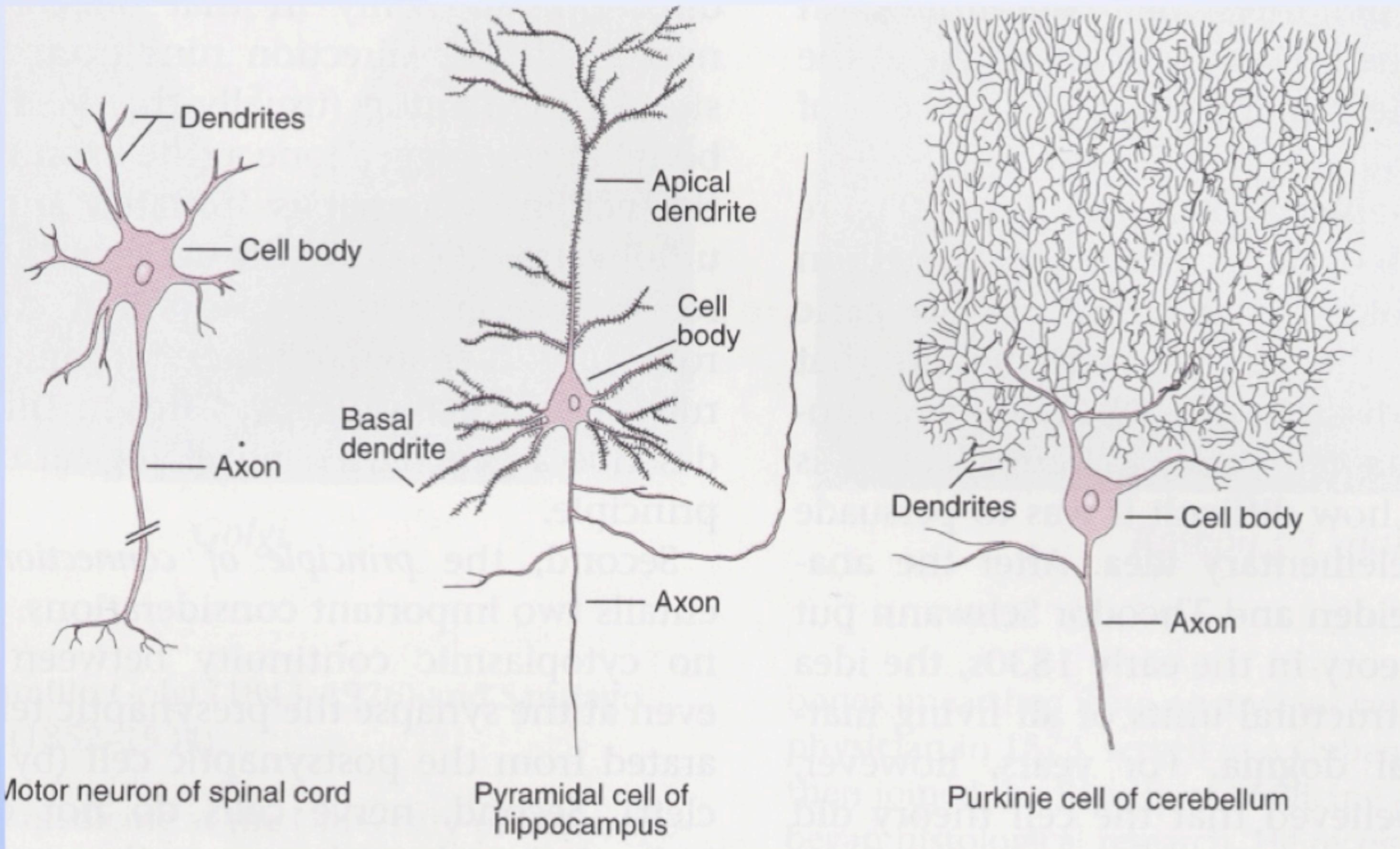


GOLGI STAIN: *potassium dichromate + silver nitrate=silver chromate*  
precipitates within the matrix of the cytoplasm

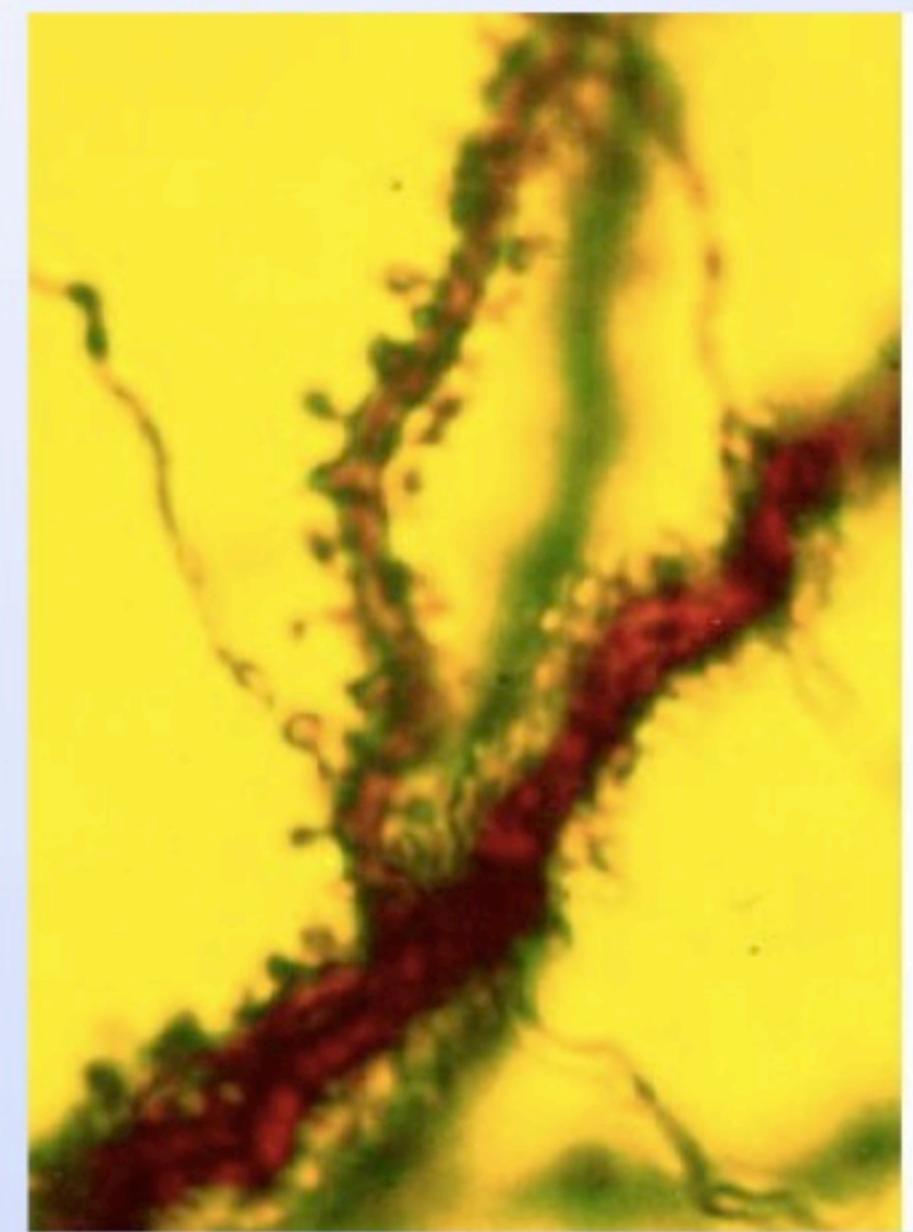
SERENDIPITOUSLY RANDOM!!

USED BY RAMON Y CAJAL TO FORMULATE THE NEURON DOCTRINE

# Different forms of dendrites



dendrite



spine



3D reconstruction from Synapse Web, Kristen M. Harris, PI  
<http://synapses.clm.utexas.edu/>

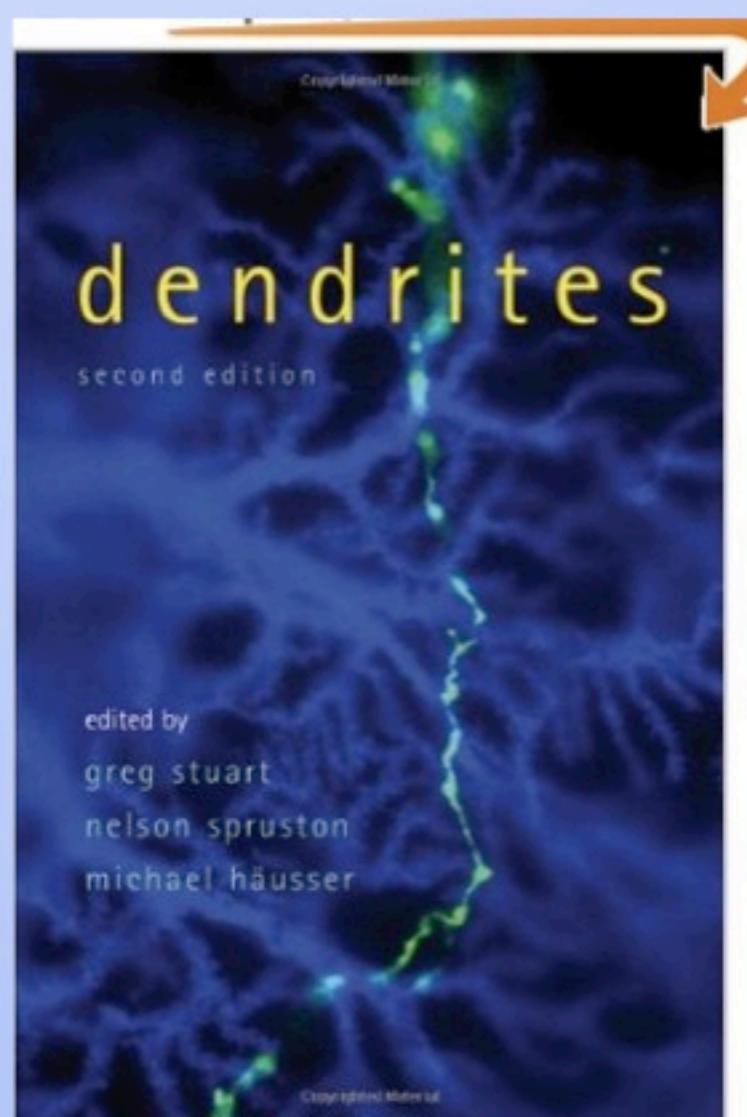
# Different forms of spines

Pattern	Characteristics	Examples	
Varicosity	An enlargement in a thinner dendrite associated with synaptic contacts	Retinal amacrine cells	
Filopodium	A long, thin protrusion with a dense actin matrix and few internal organelles	Normally only seen during development	
Simple Spine Sessile	Synaptic protrusions without a neck constriction	Sessile spine Stubby spine Crook thorn	Pyramidal cells of cortex Cerebellar dentate nucleus
Pedunculated	Bulbous enlargement at tip	Thin spine Mushroom spine Gemmule	Pyramidal cells of cortex Pyramidal cells of cortex Olfactory bulb granule cell
Branched Spine	Each branch has a unique presynaptic partner and each branch has the shape characteristics of a simple spine	CA1 pyramidal cells Granule cells of dentate gyrus Cerebellar Purkinje cells	

Adapted from Fiala JC, Harris KM (1999) Dendrite Structure. In: G Stuart, N Spruston, M Häusser, (eds.) *Dendrites*. Oxford University Press: Oxford UK.

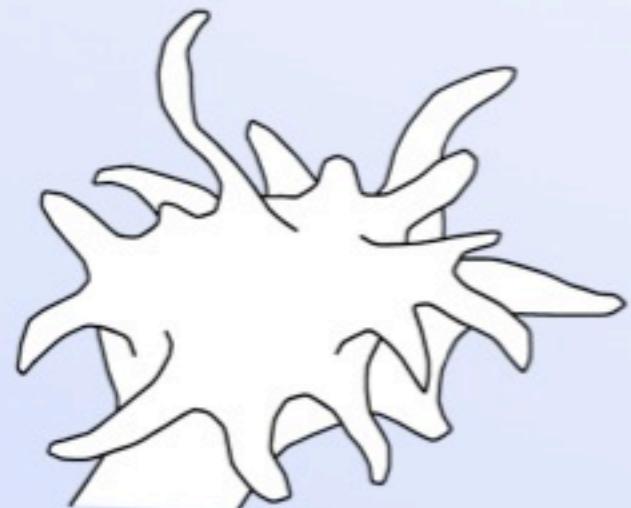


3D reconstruction from Synapse Web, Kristen M. Harris, PI  
<http://synapses.clm.utexas.edu/>

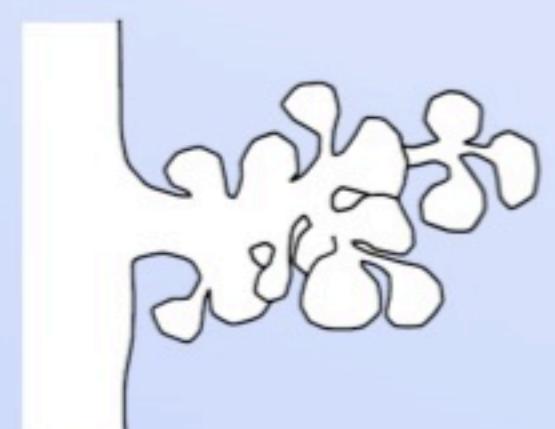


# Different forms of spines

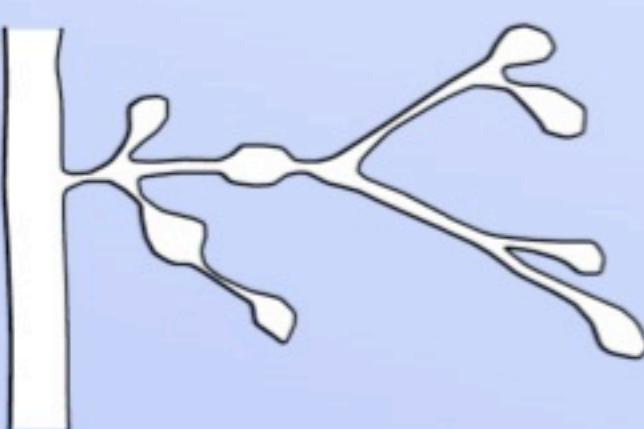
Brush Ending



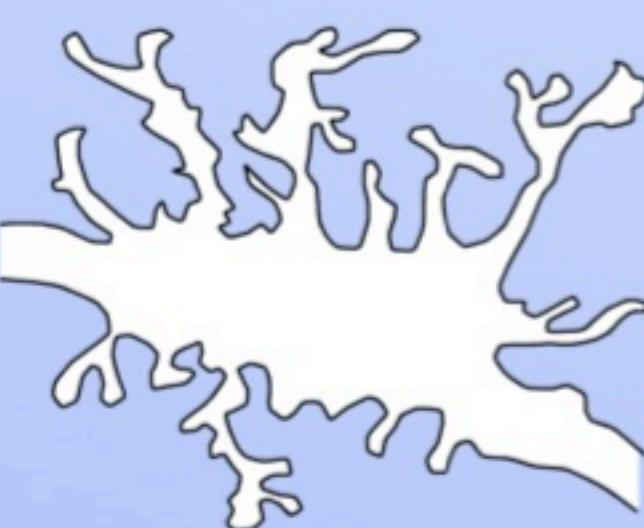
Thorny Excrescence



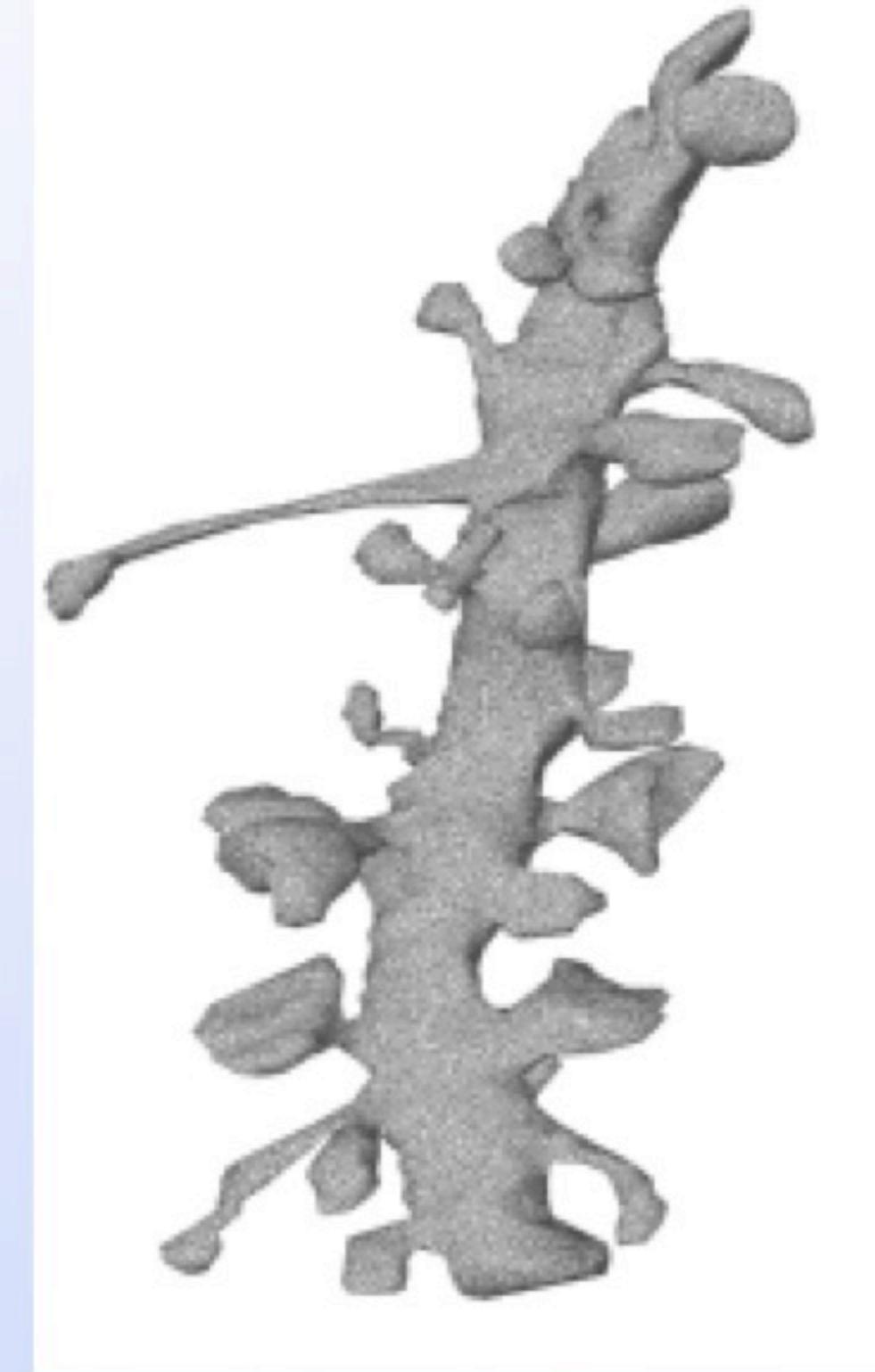
Racemose Appendage



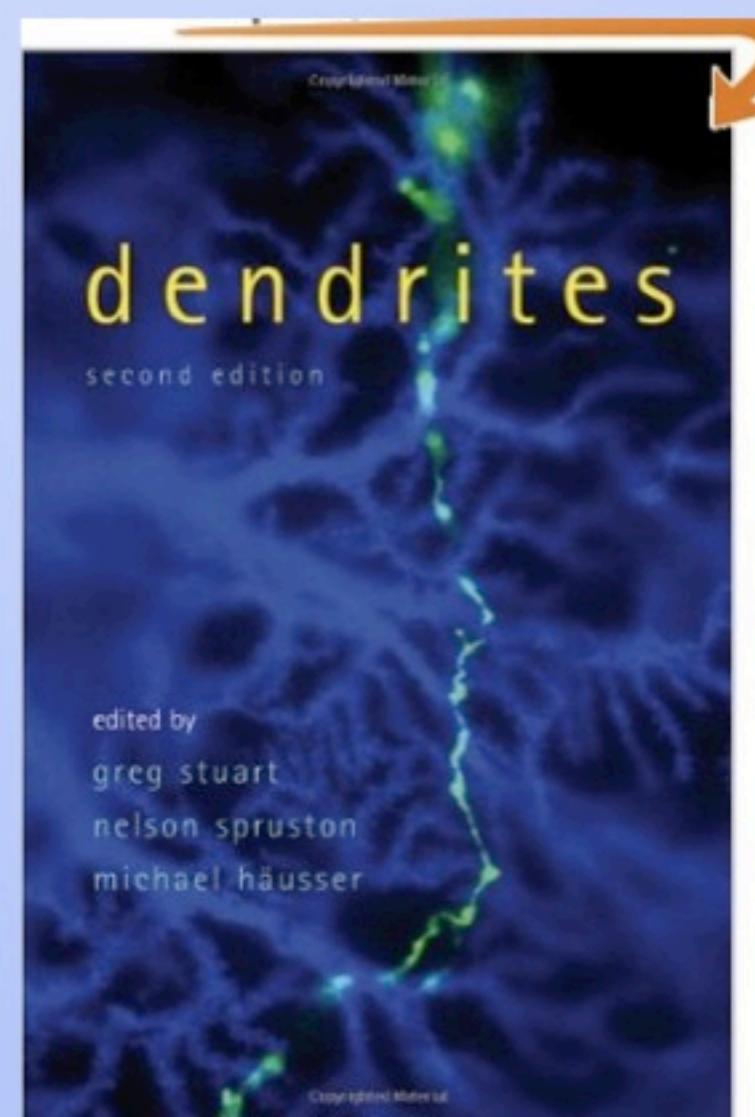
Coralline Excrescence



	Characteristics	Examples
Brush Ending	Spray of complex dendritic protrusions at the end of dendrite that extends into glomerulus and contains presynaptic elements	Unipolar brush cells of cerebellar cortex and dorsal cochlear nucleus
Thorny Excrescence	Densely lobed dendritic protrusion into a glomerulus	Proximal dendrites of CA3 pyramidal cells and dentate gyrus mossy cells
Racemose Appendage	Twig-like branched dendritic appendages that contain synaptic varicosities and bulbous tips	Inferior olive Relay cells of lateral geniculate nucleus
Coralline Excrescence	Dendritic varicosity extending numerous thin protrusions, velamentous expansions and tendrils	Cerebellar dentate nucleus Lateral vestibular nucleus



3D reconstruction from Synapse Web, Kristen M. Harris, PI  
<http://synapses.clm.utexas.edu/>



Fiala JC, Harris KM (1999) Dendrite Structure. In: G Stuart, N Spruston, M Häusser, (eds.) Dendrites. Oxford University Press: Oxford UK.

Dendrite from a normal infant

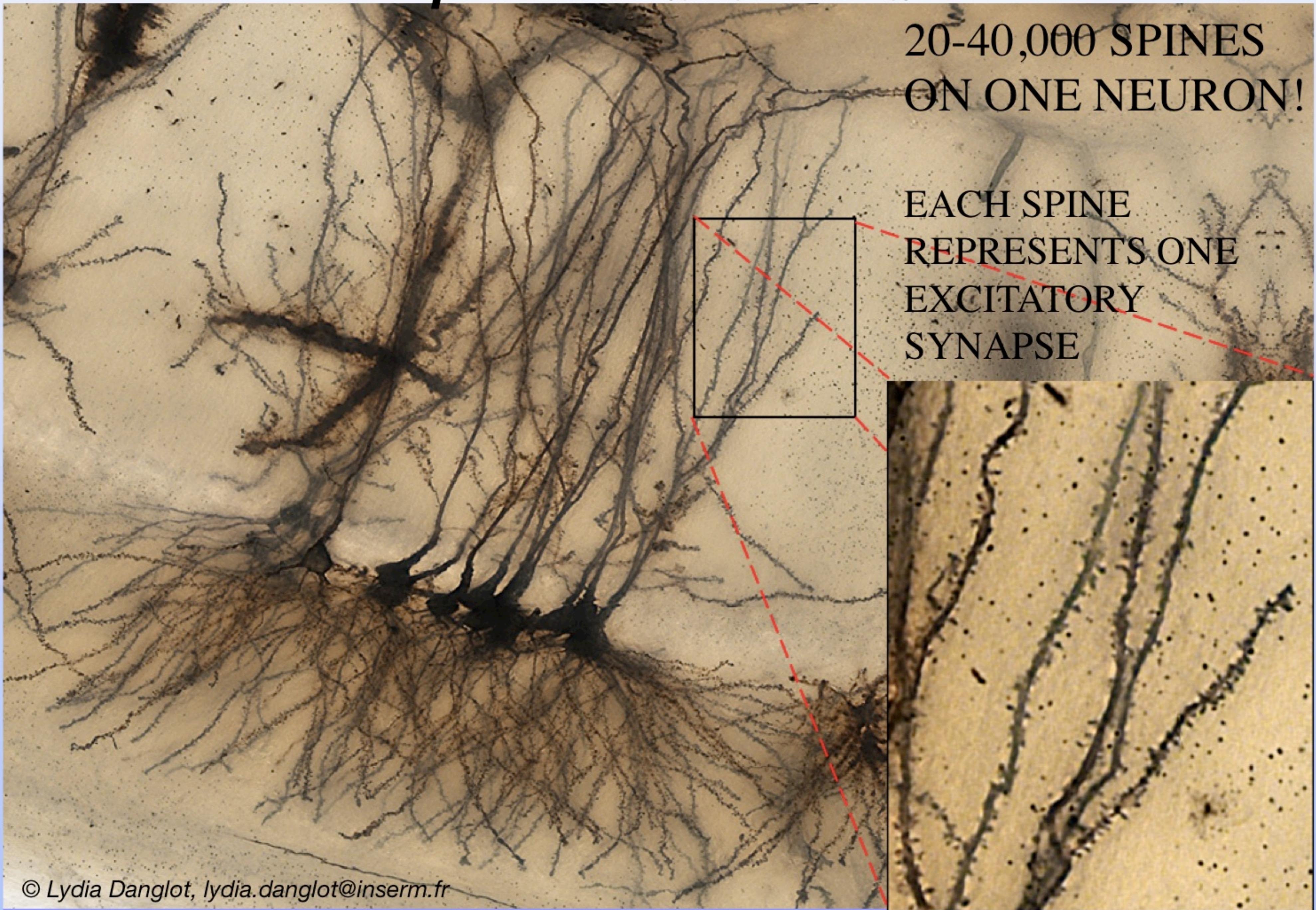


Dendrite from a mentally retarded infant

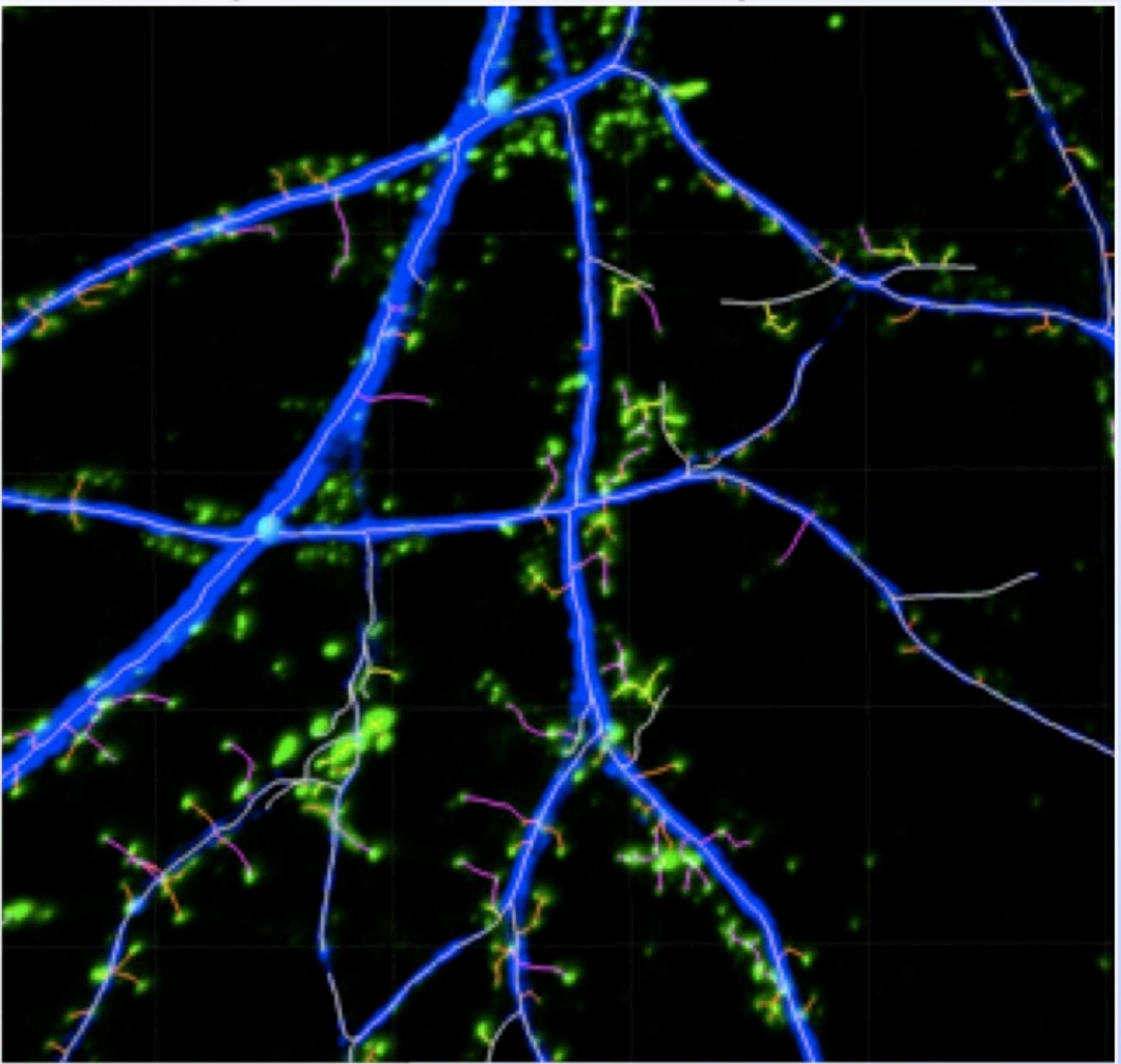
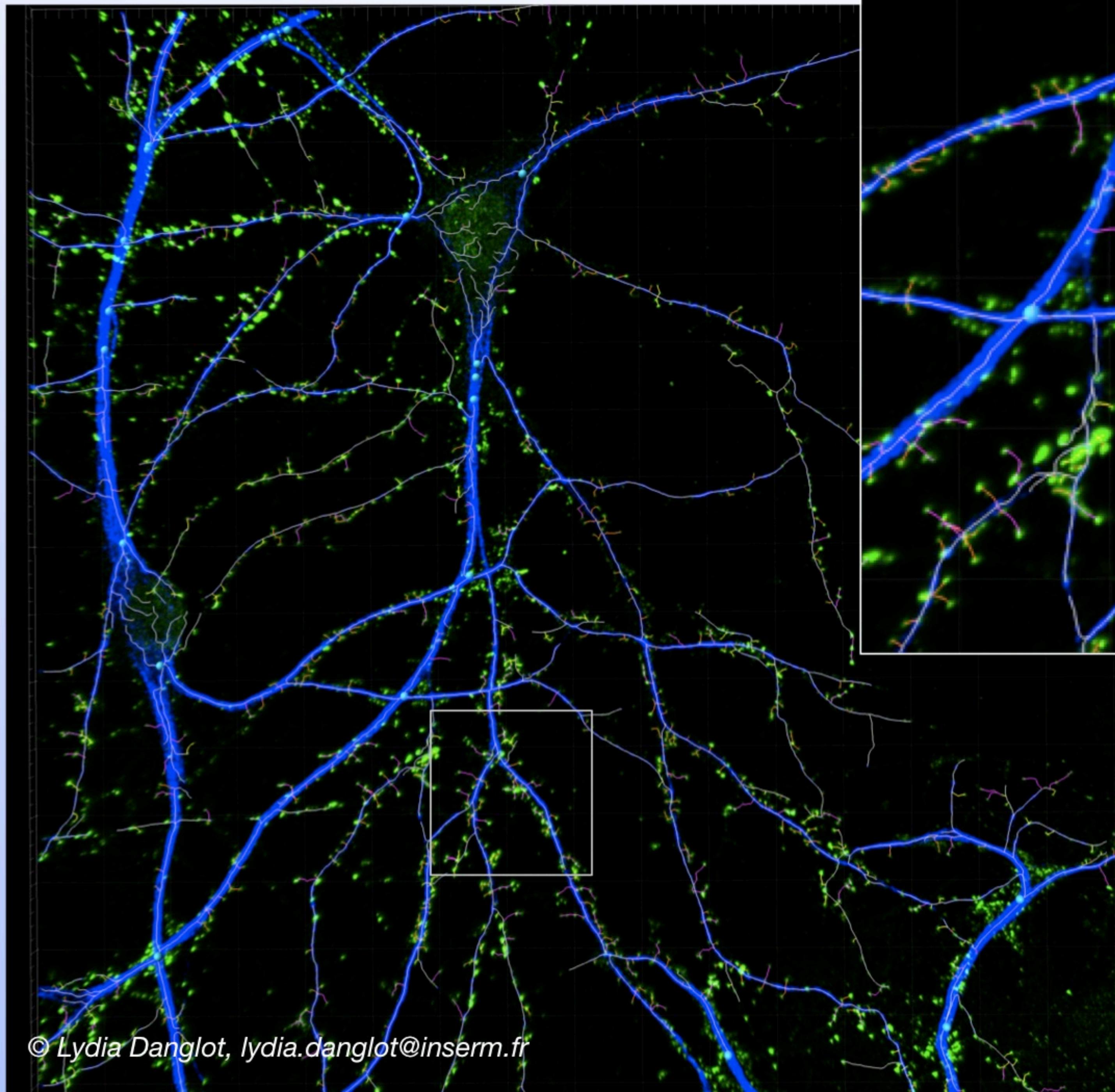


10  $\mu\text{m}$

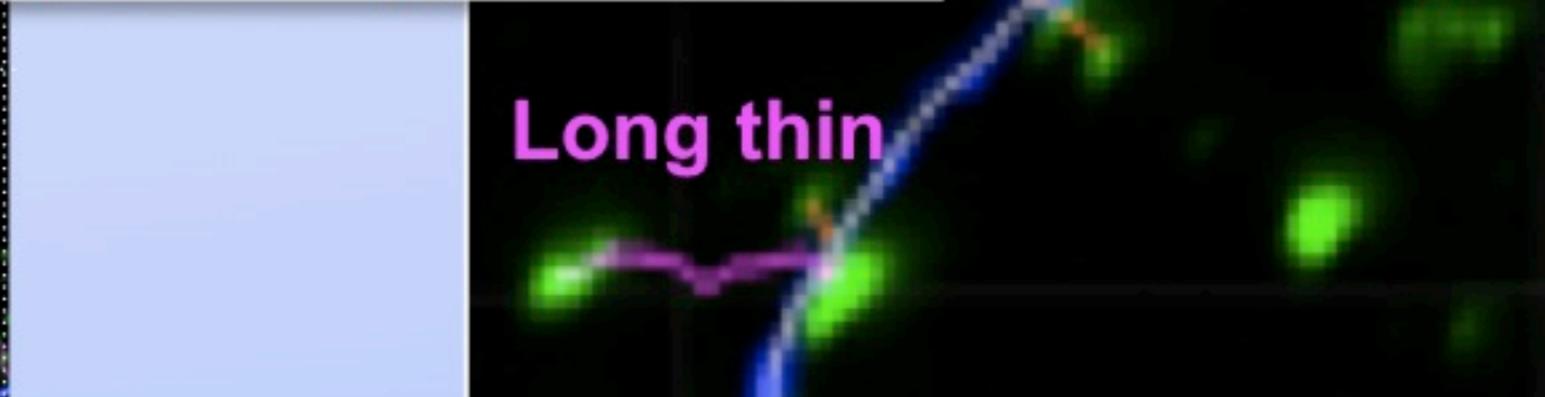
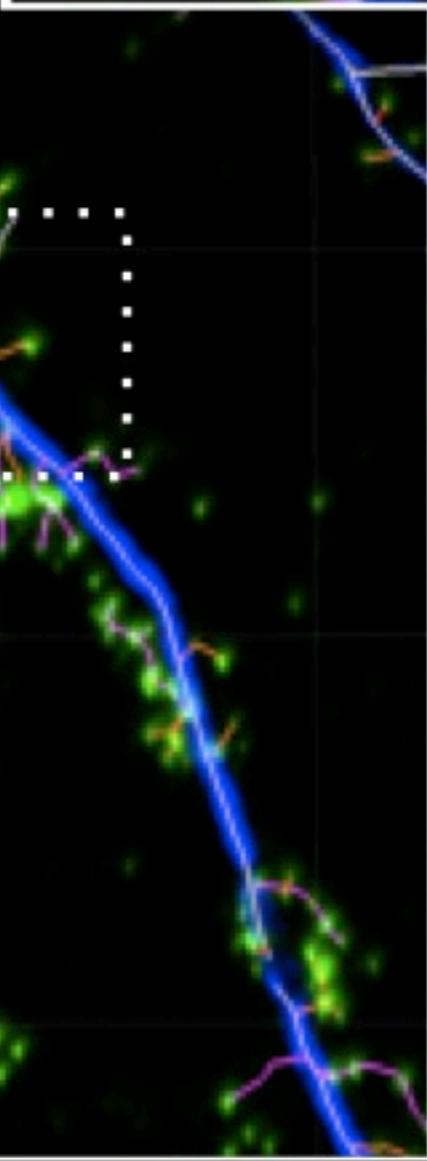
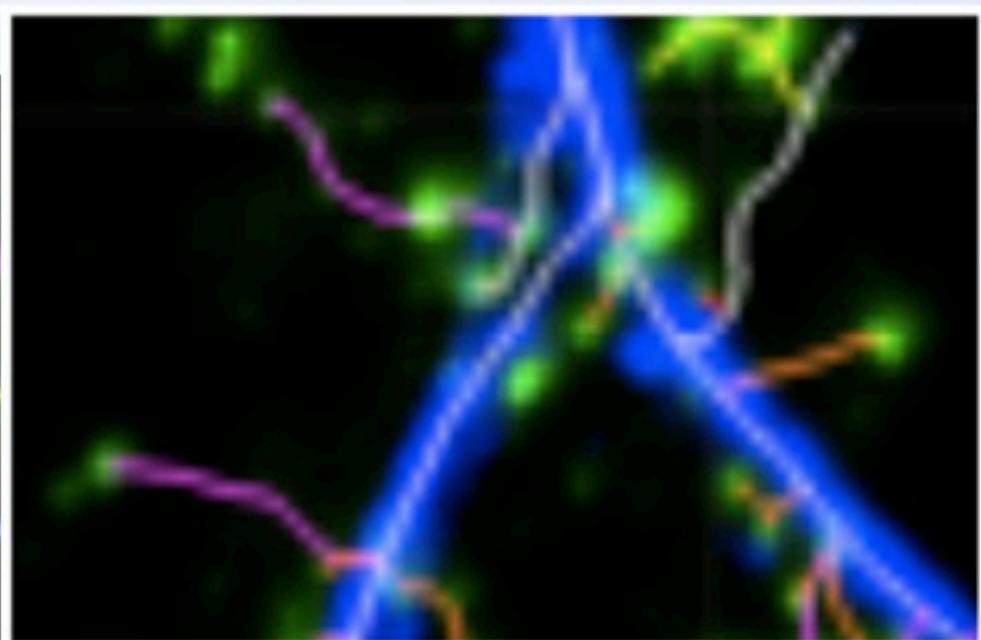
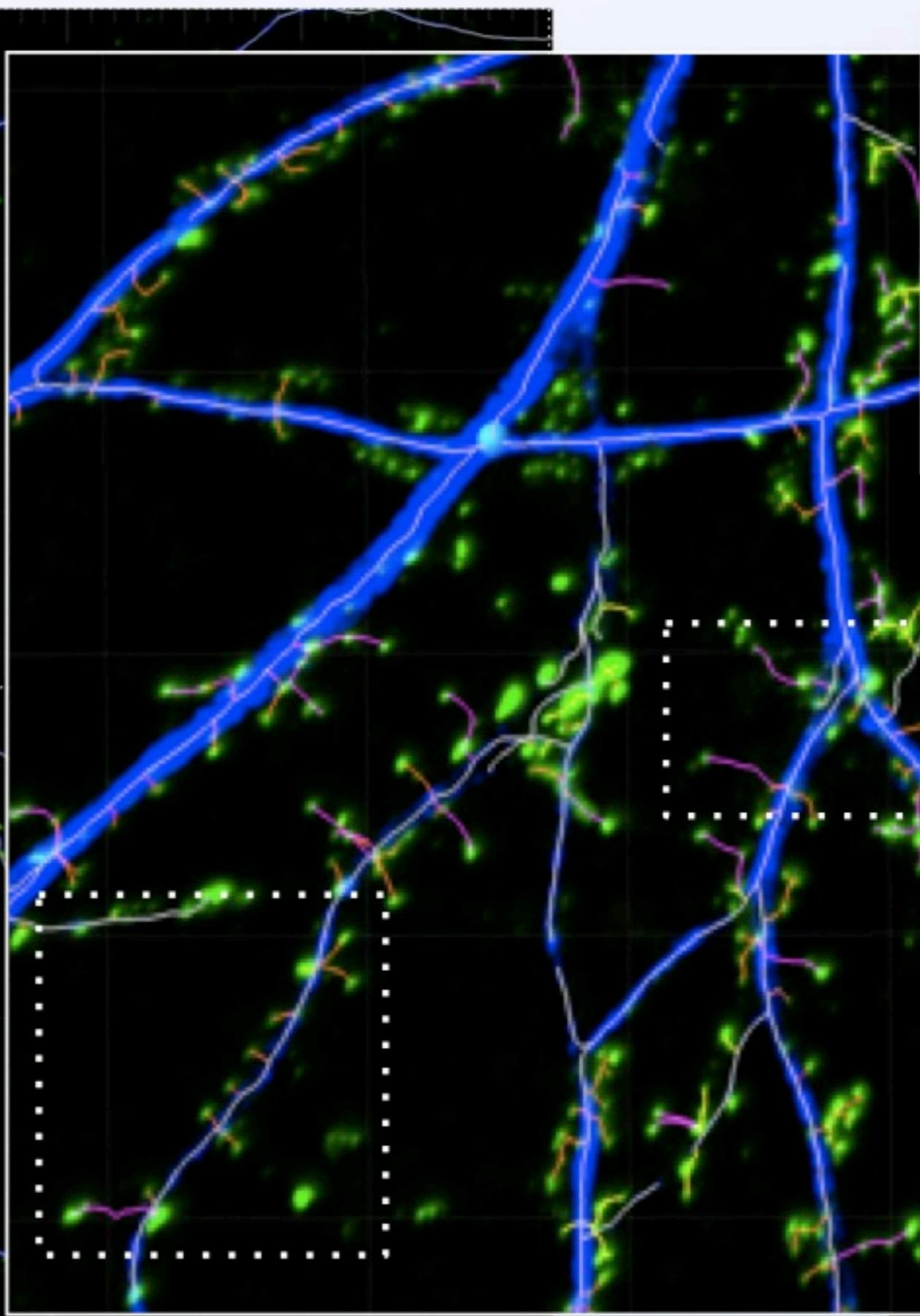
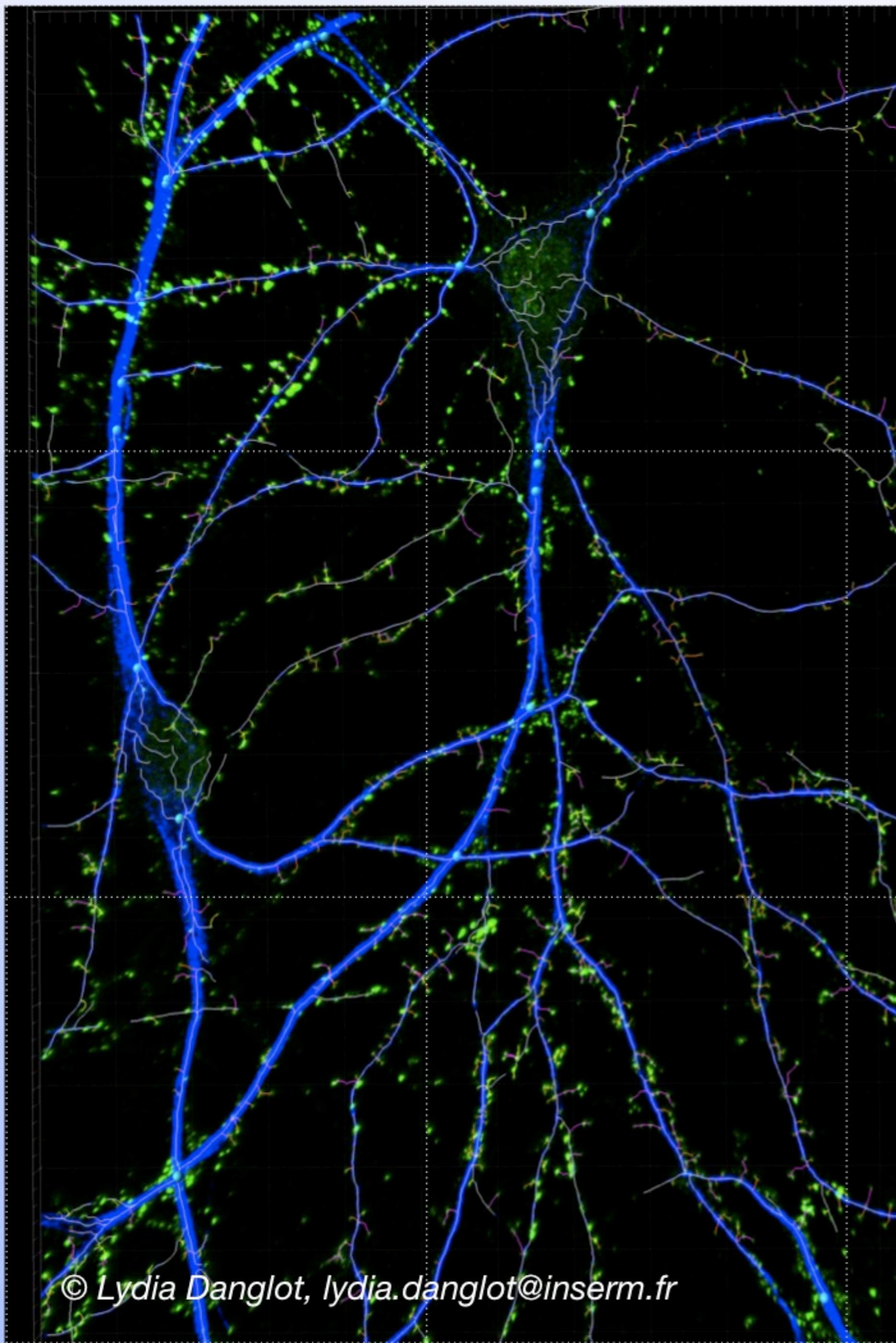
# Dendritic Spines *(golgi staining)*



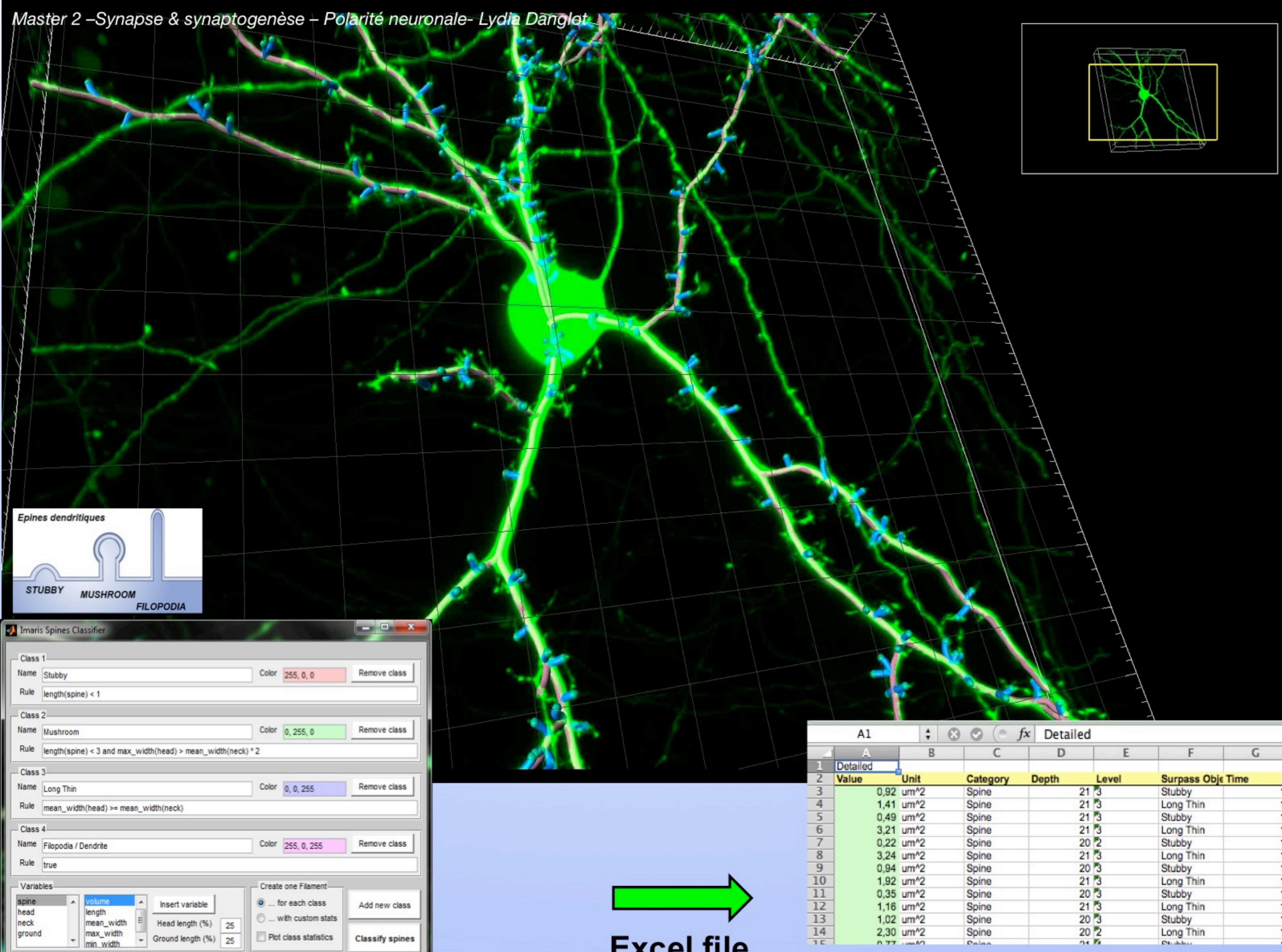
# Dendritic Spines (fluorescence)



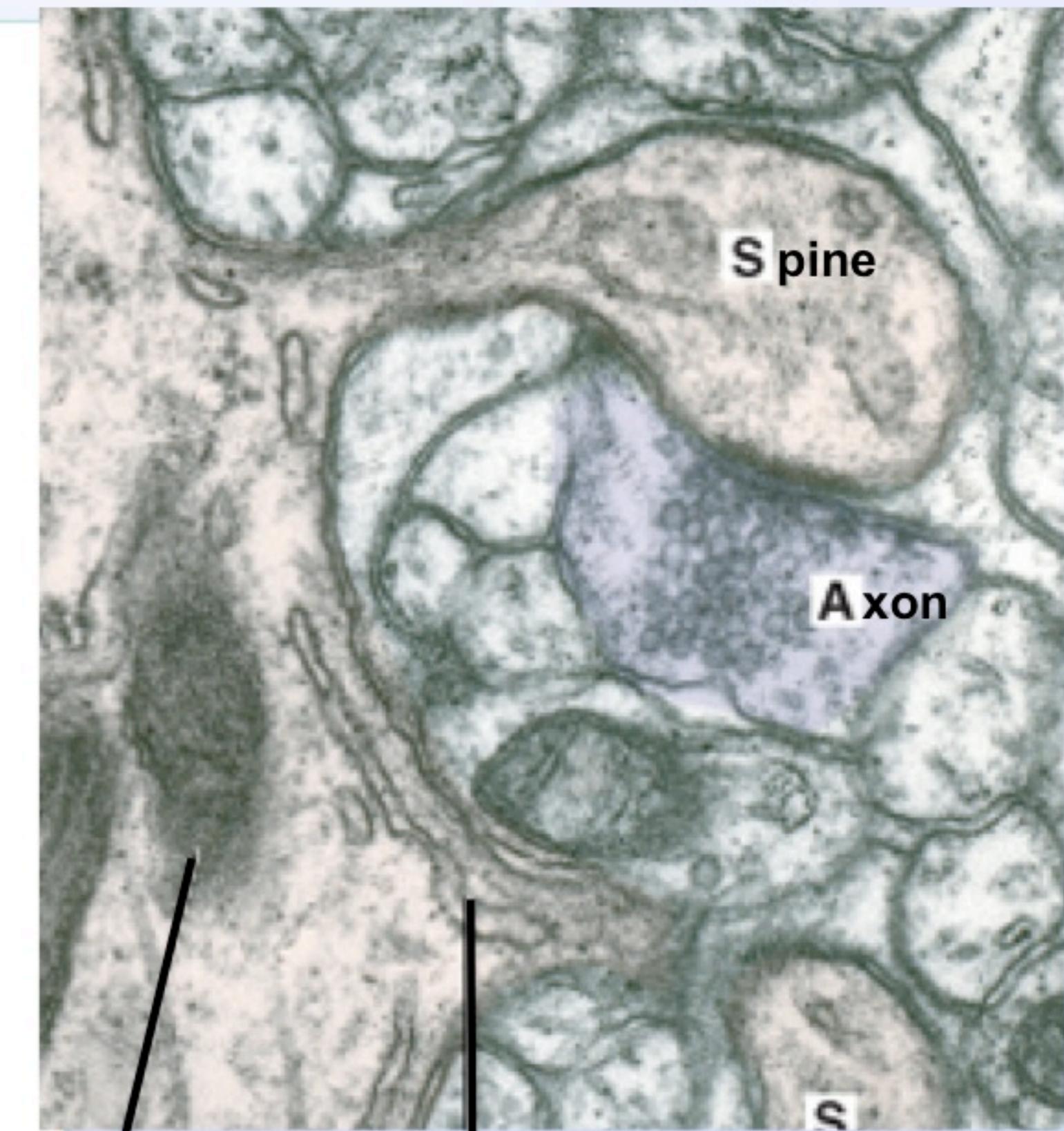
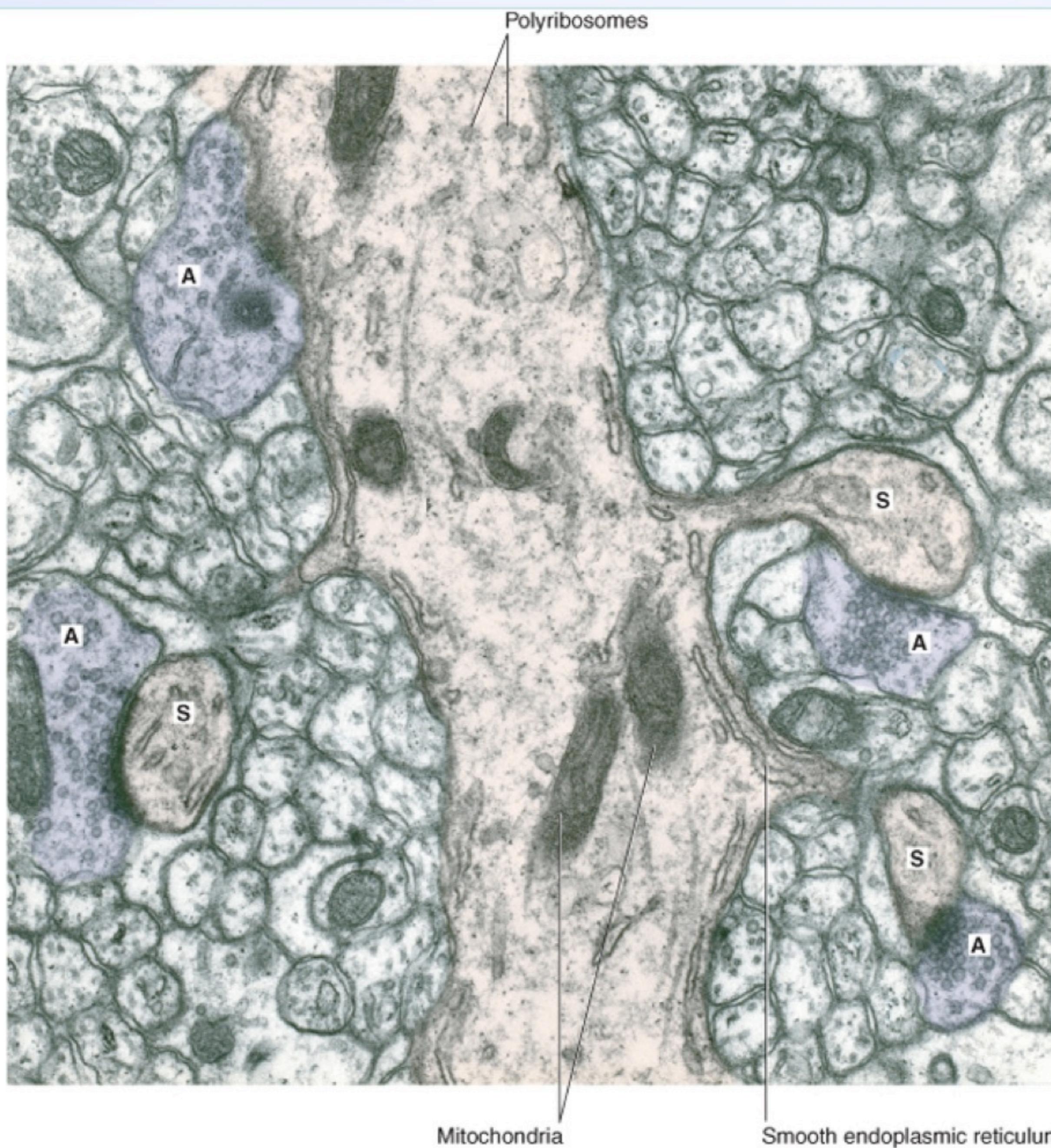
## Spine morphology?



Tile confocal imaging,  
3D reconstruction (Imaris)  
and spine quantification (Matlab).



# Dendritic Spines (electron microscopy)



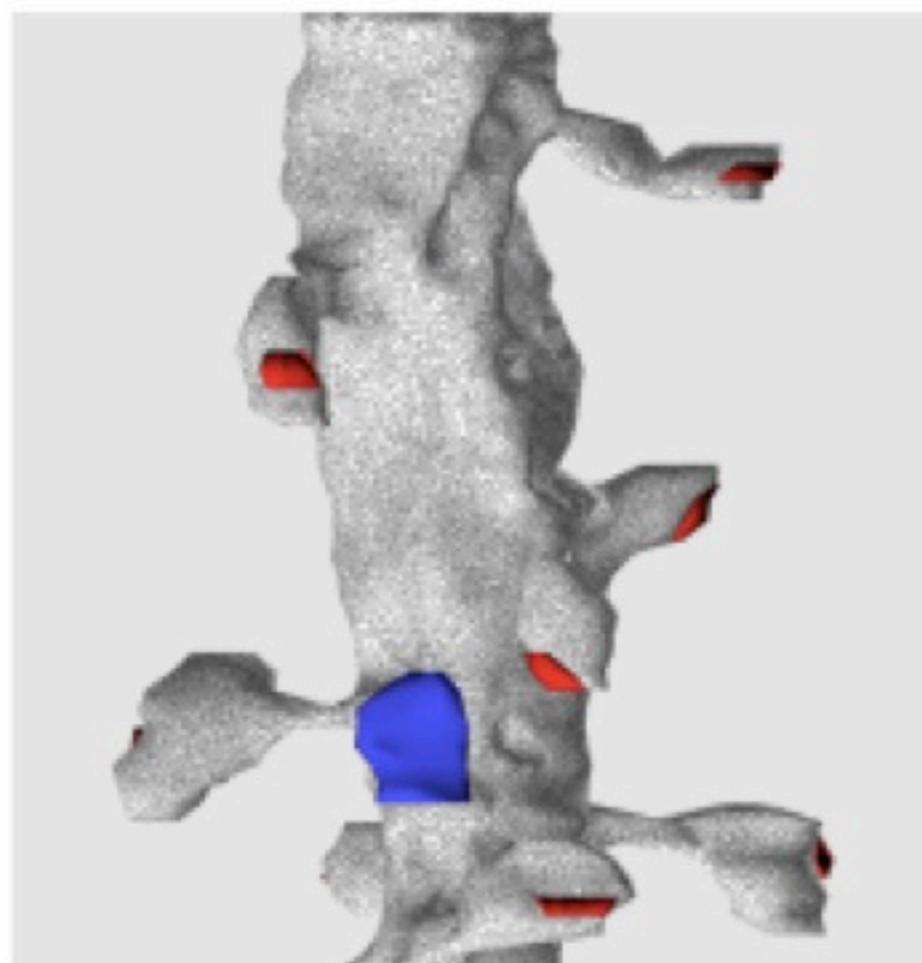
Reticulum Endoplasmique  
Lisse

# Different forms of spines

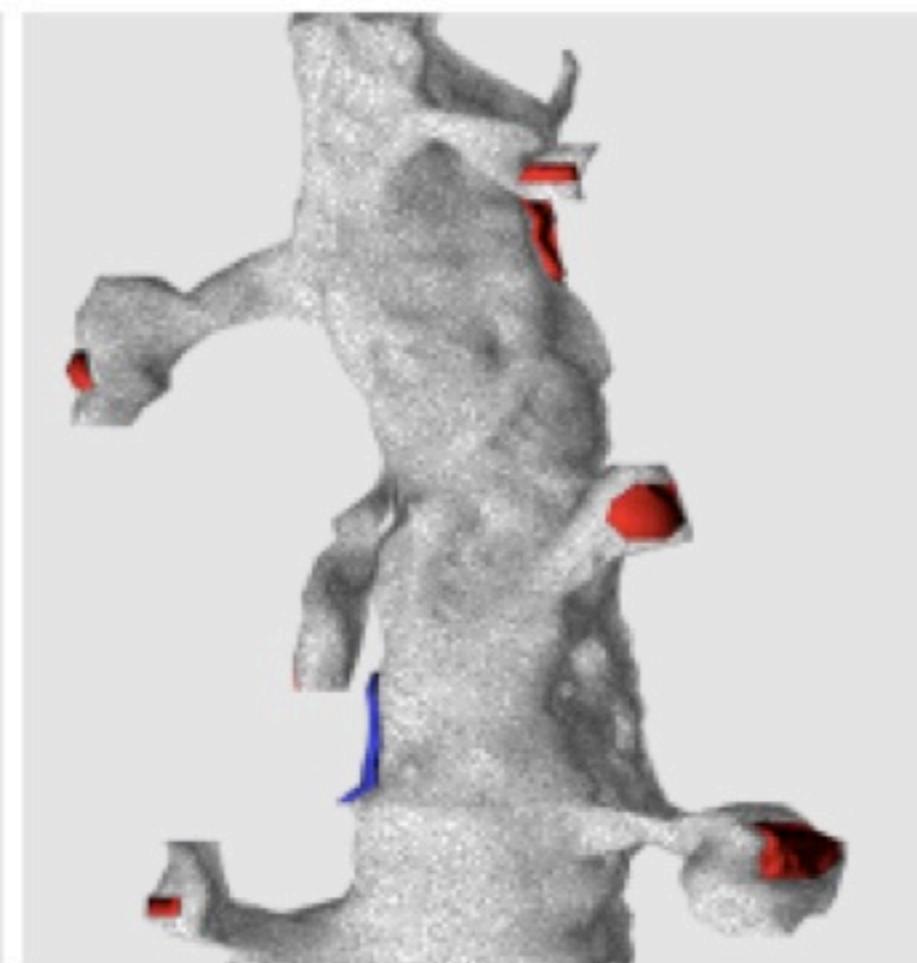
**SynapseWeb**

About      Anatomy      Links      Publications

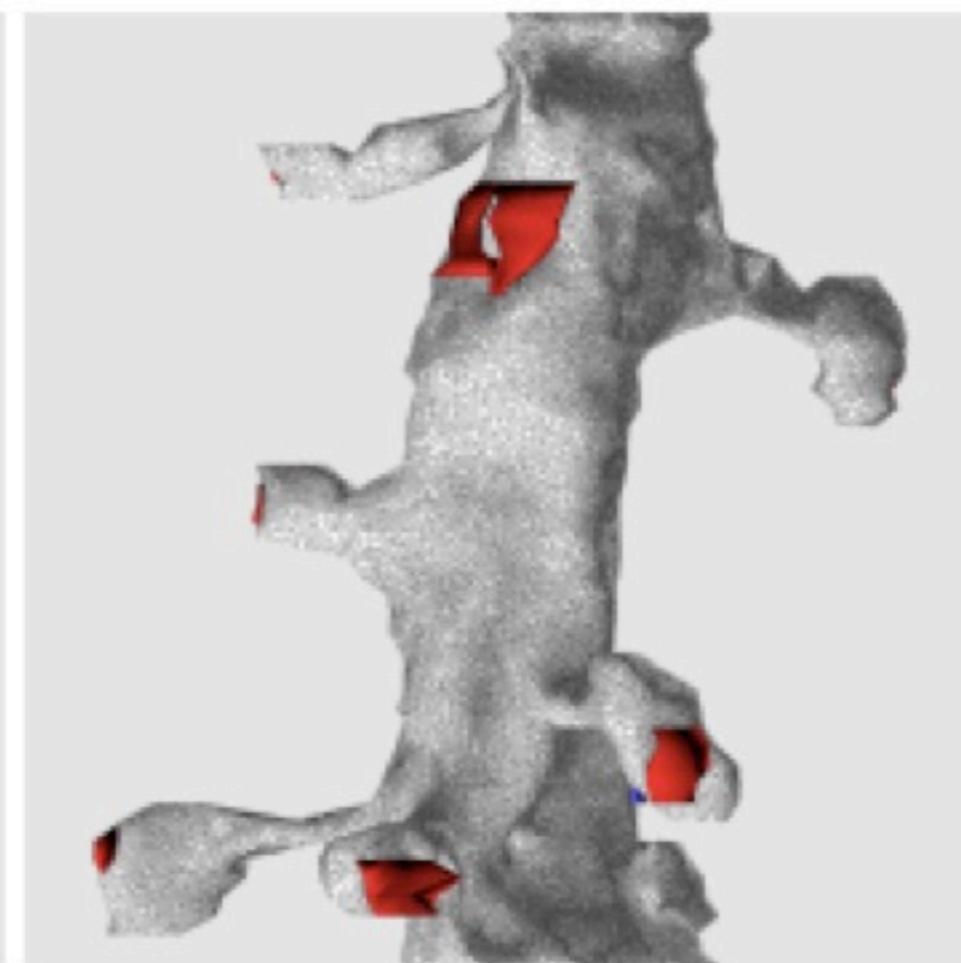
A spiny dendrite from CA1 stratum radiatum (series K24).



Front View



Side View



Back View

1.0 μm

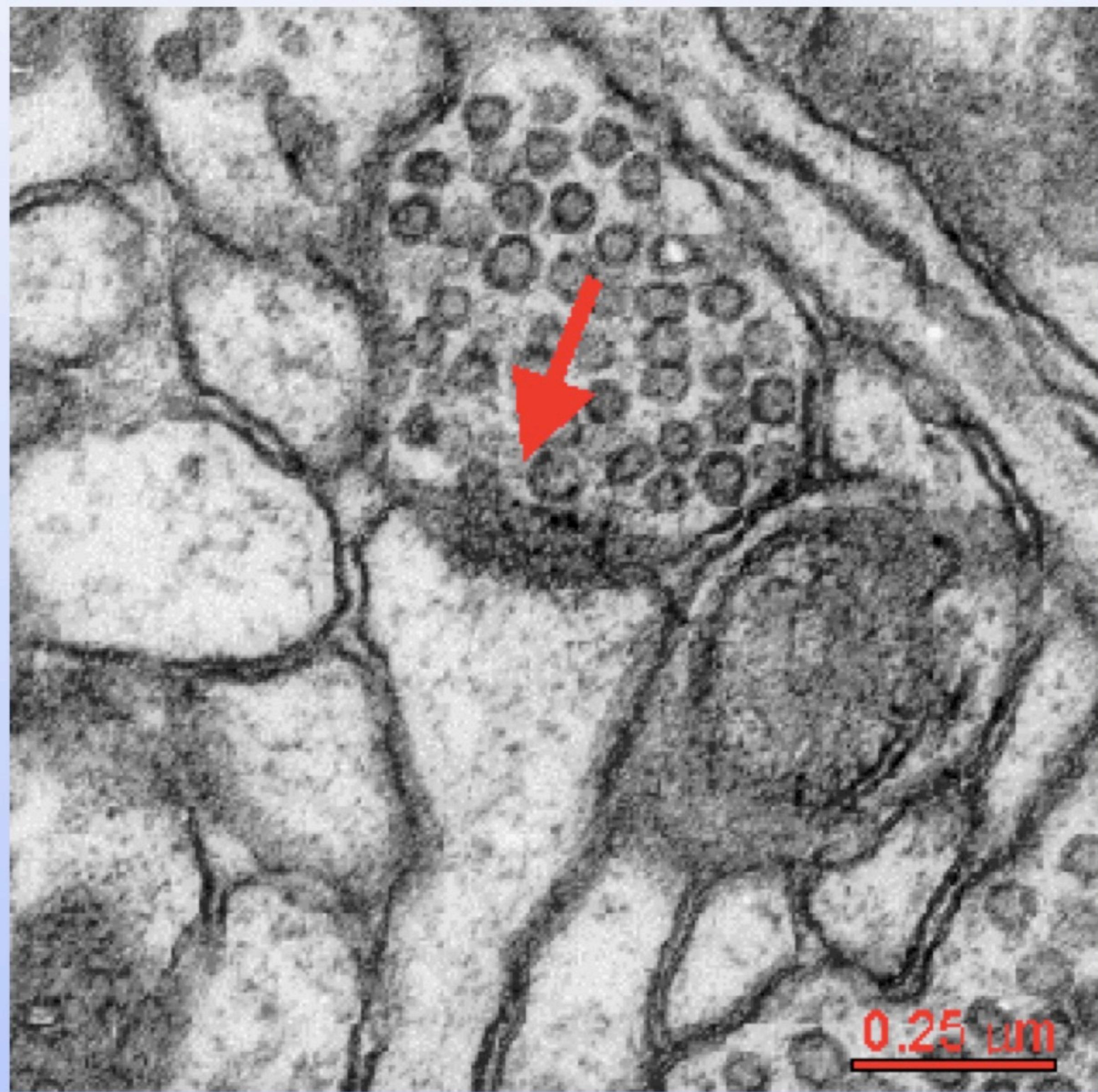
Excitatory synapses are shown in red. Inhibitory synapses in blue.

Excitatory synapse

Inhibitory synapse

**3D** View this dendrite as a [flip book of serial section images](#) (2.62Mb Javascript). View a three-dimensional reconstruction of this dendrite as a [rotating GIF](#) (126Kb), as a [VRML 1.0 object\\*](#) (450Kb). You may also [download](#) (4.65Mb) the data which appears in the flipbook (46 sections) as an IGL Trace\*\* series of Windows Bitmap images.

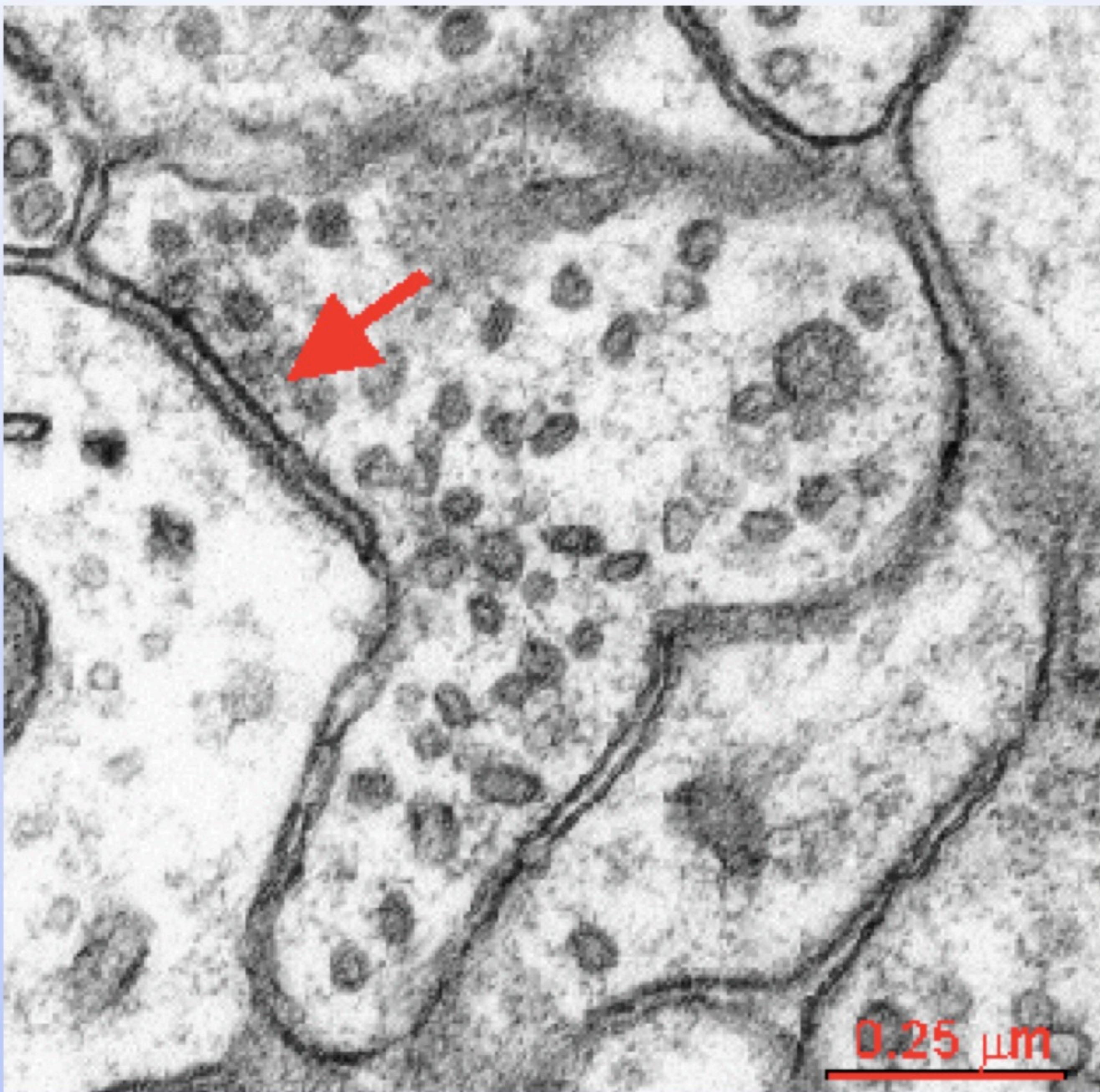
# Asymmetric synapse (GRAY'S TYPE I) excitatory



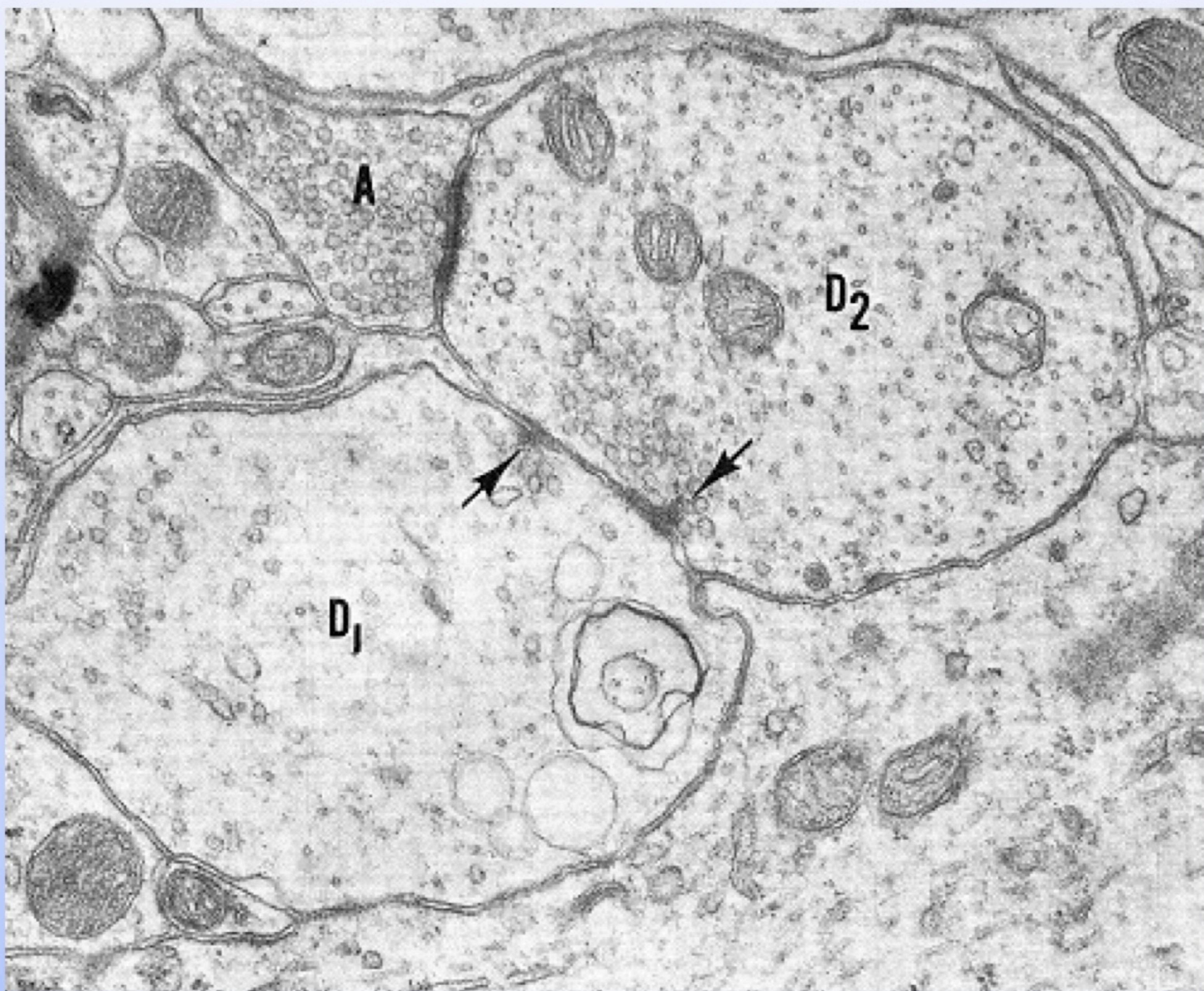
# WHAT TYPE OF SYNAPSE?



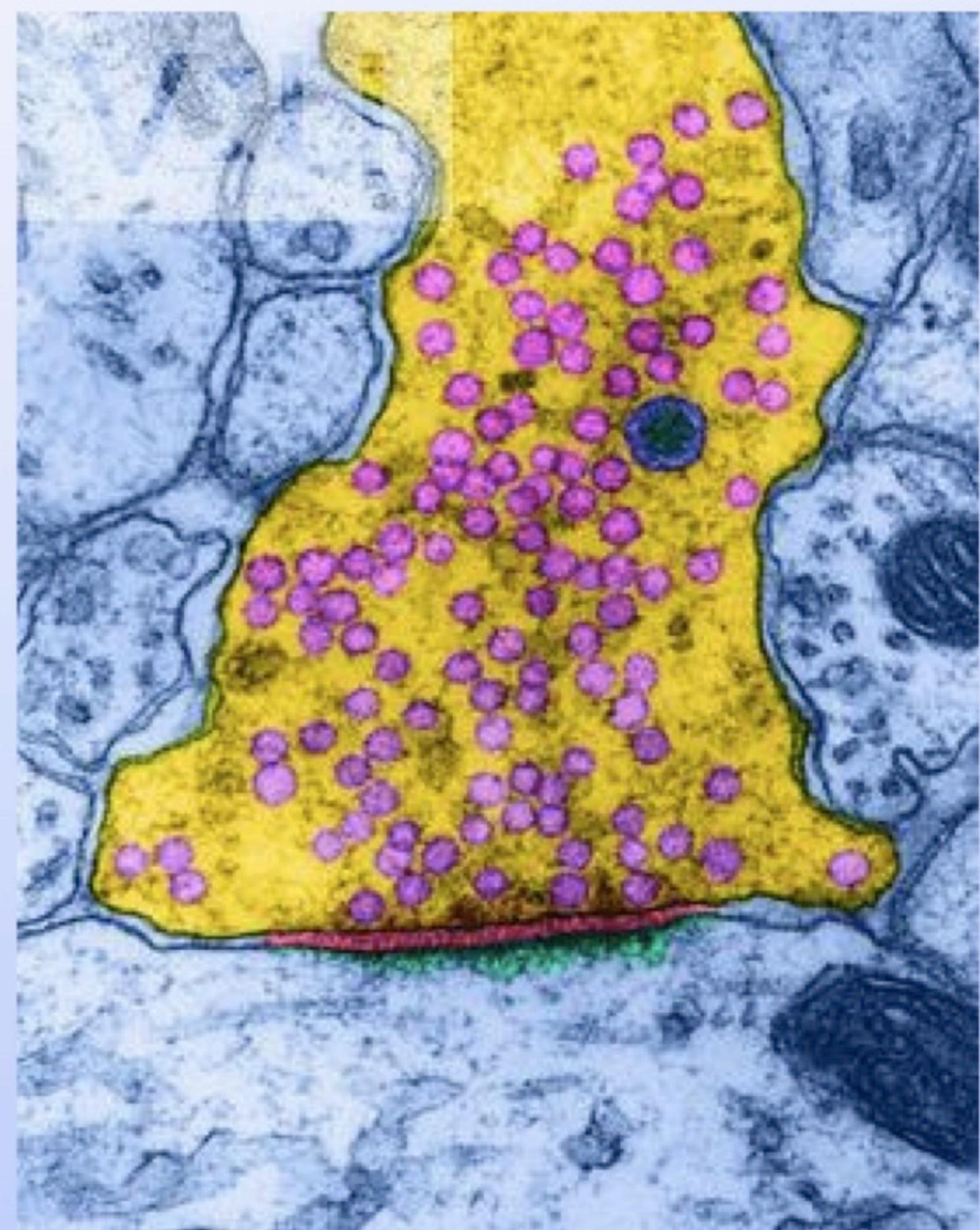
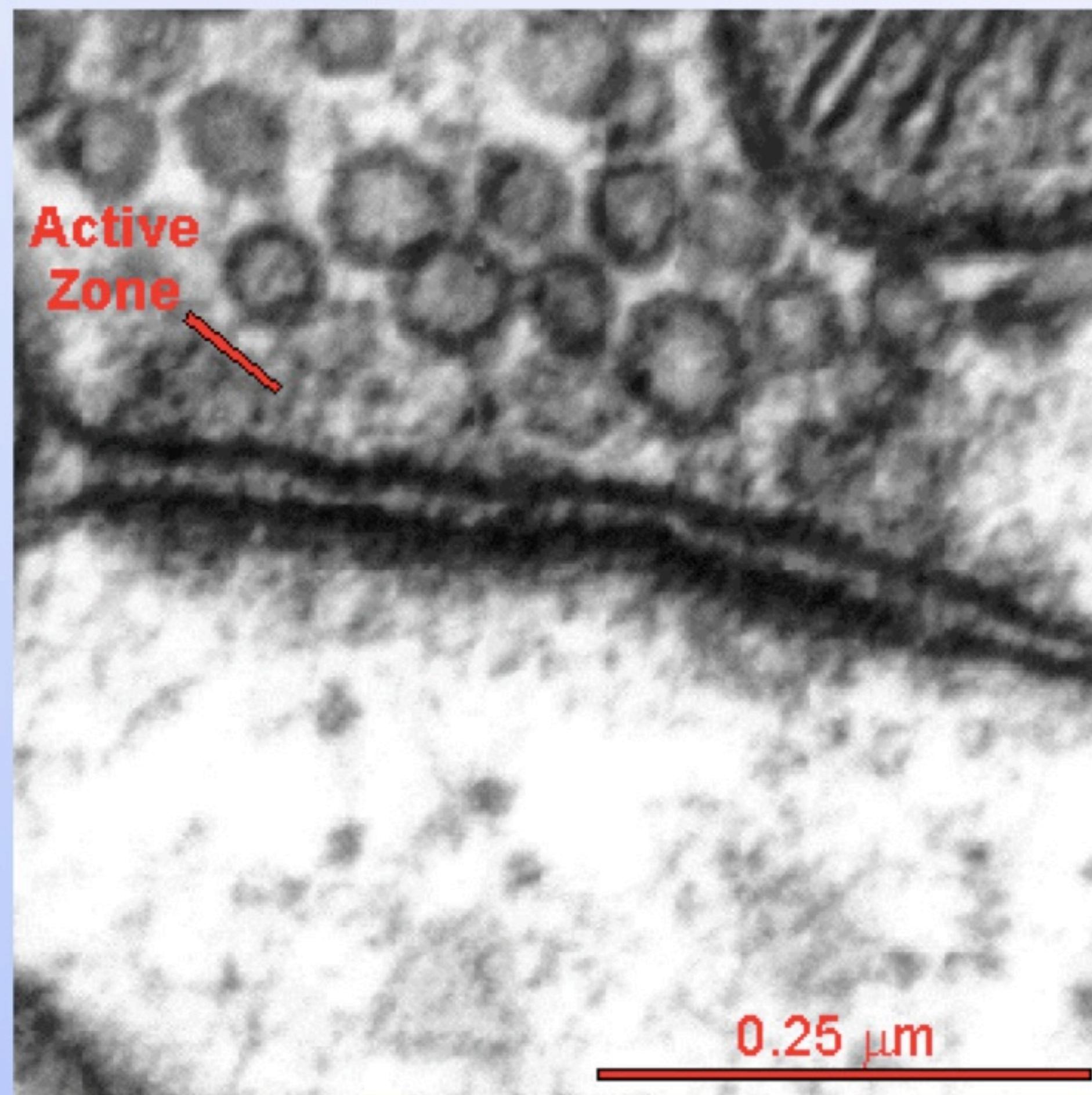
# SYMMETRIC (GRAY'S II) : INHIBITORY



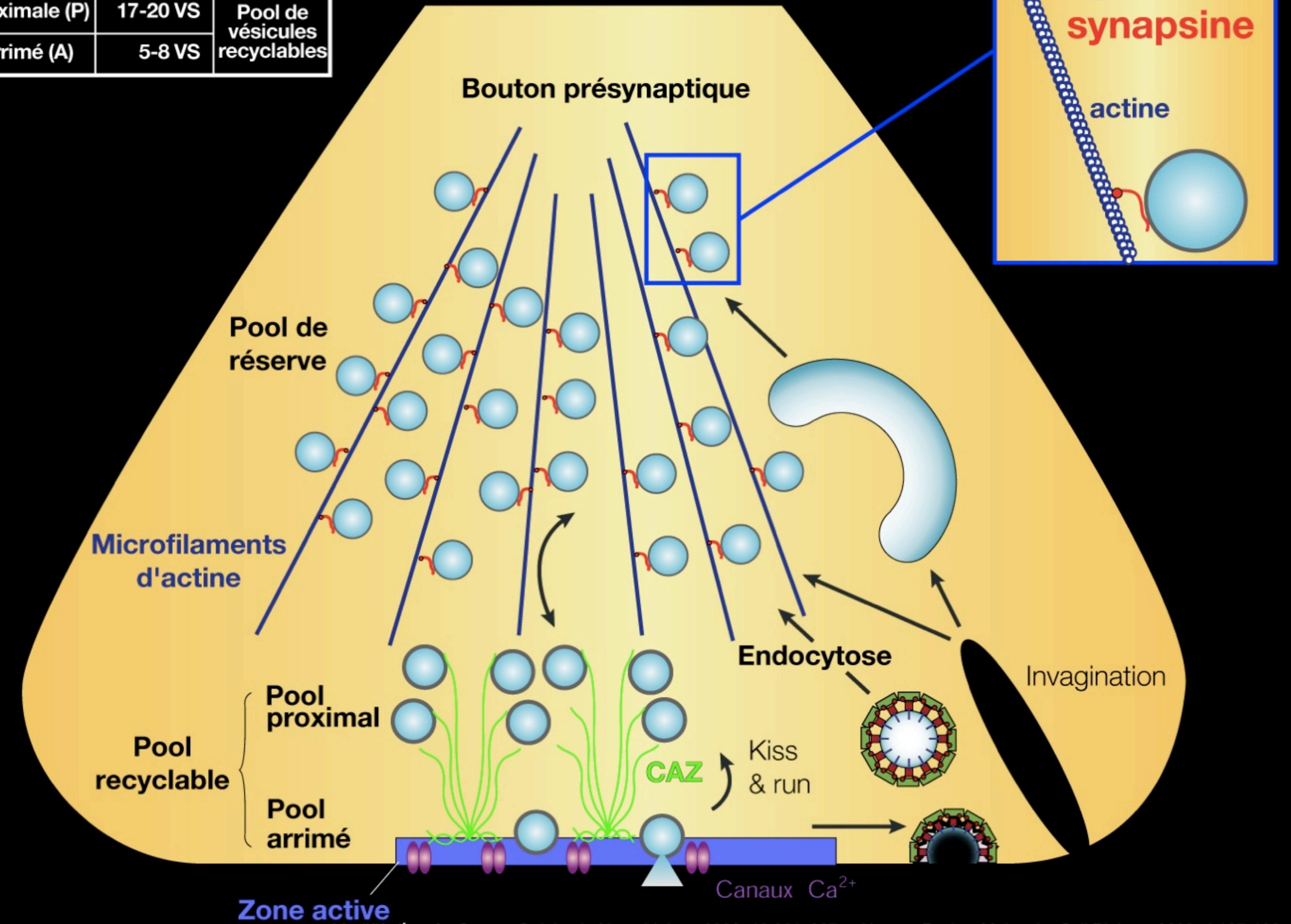
# DENDRO-DENDRITIC SYNAPSE



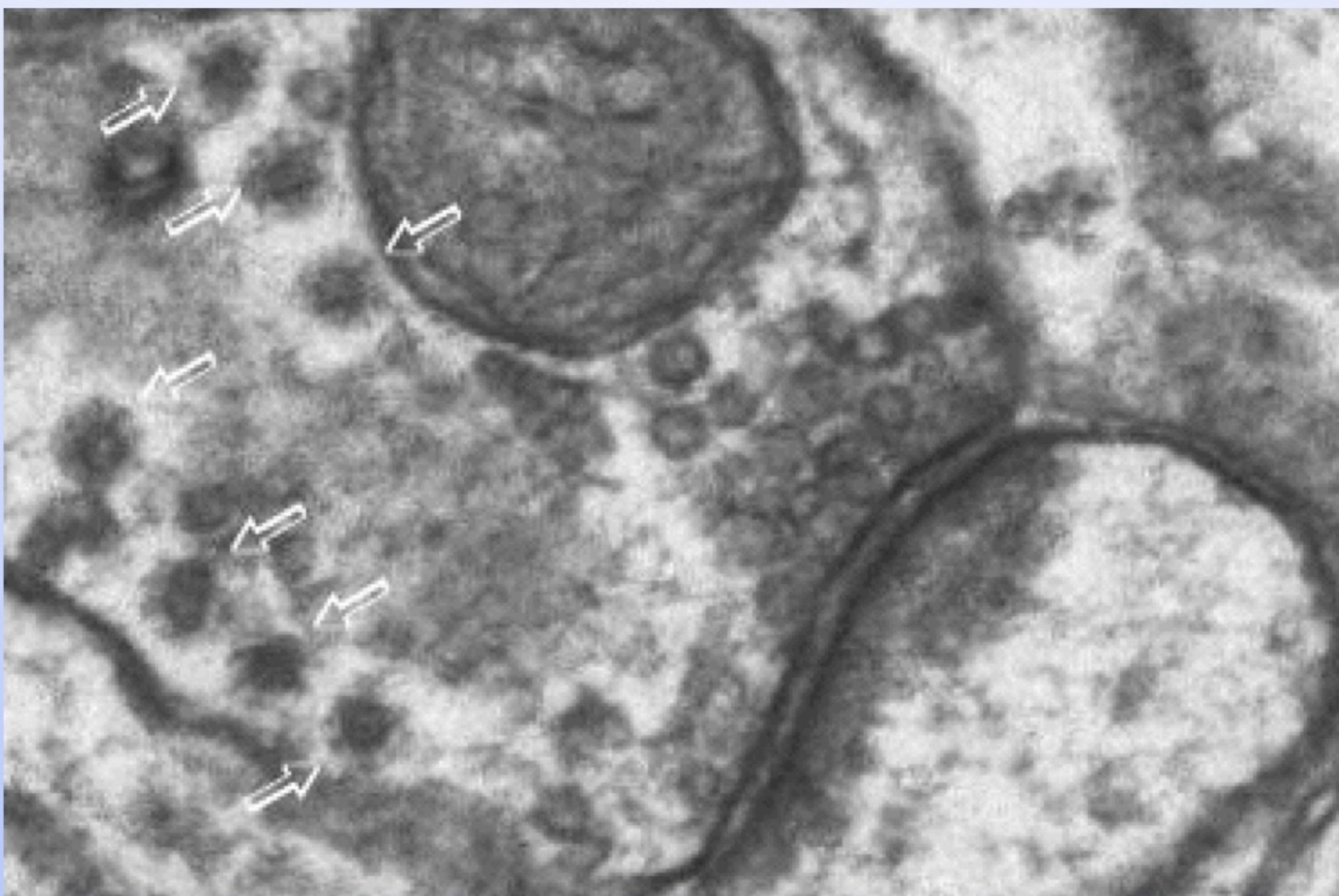
# Presynaptic Active Zone



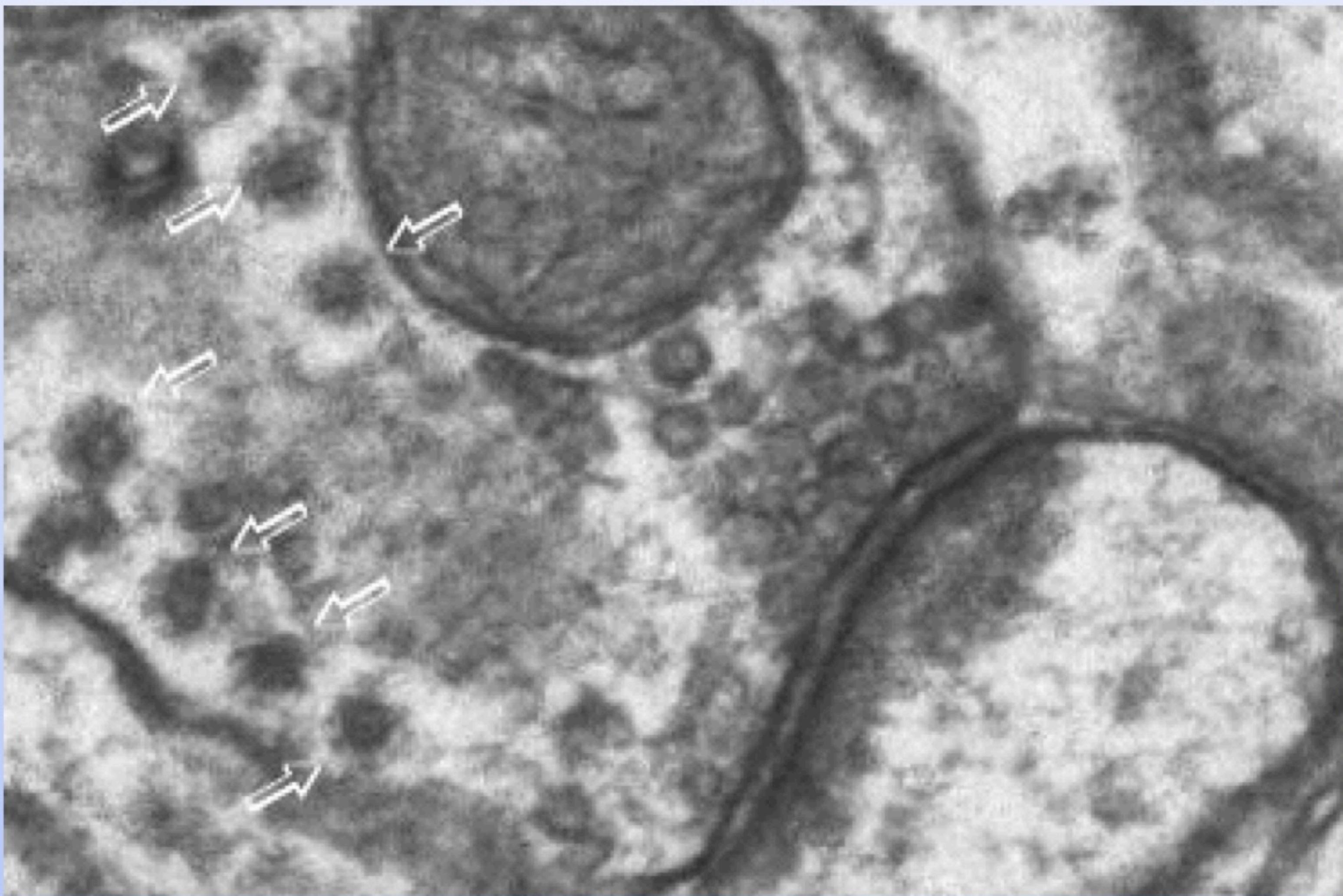
Type de pool	Nombre de vésicules	
de réserve (R)	180 VS	
proximale (P)	17-20 VS	Pool de vésicules recyclables
arrimé (A)	5-8 VS	



?



# COATED VESICLES (CLATHRIN) MEMBRANE RECYCLING AT THE TERMINAL

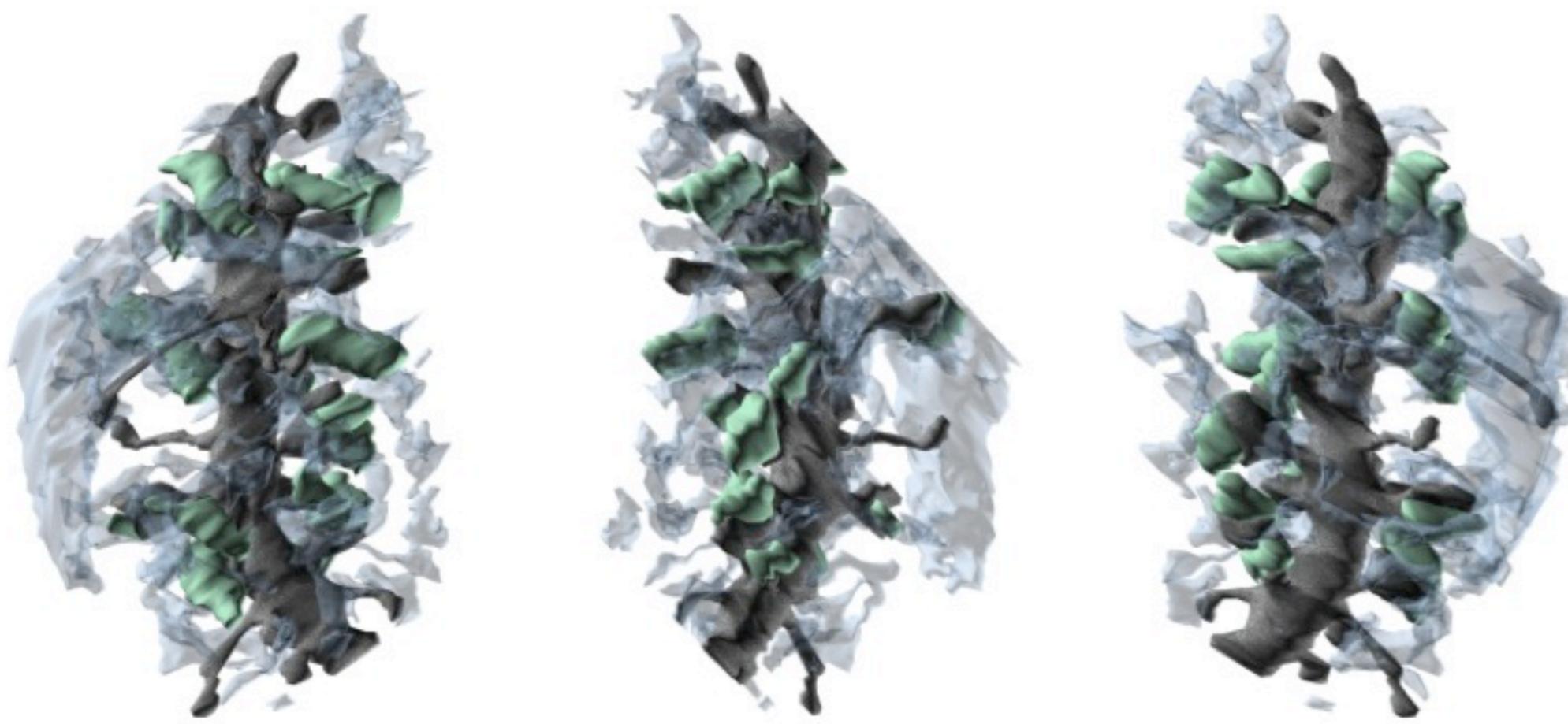
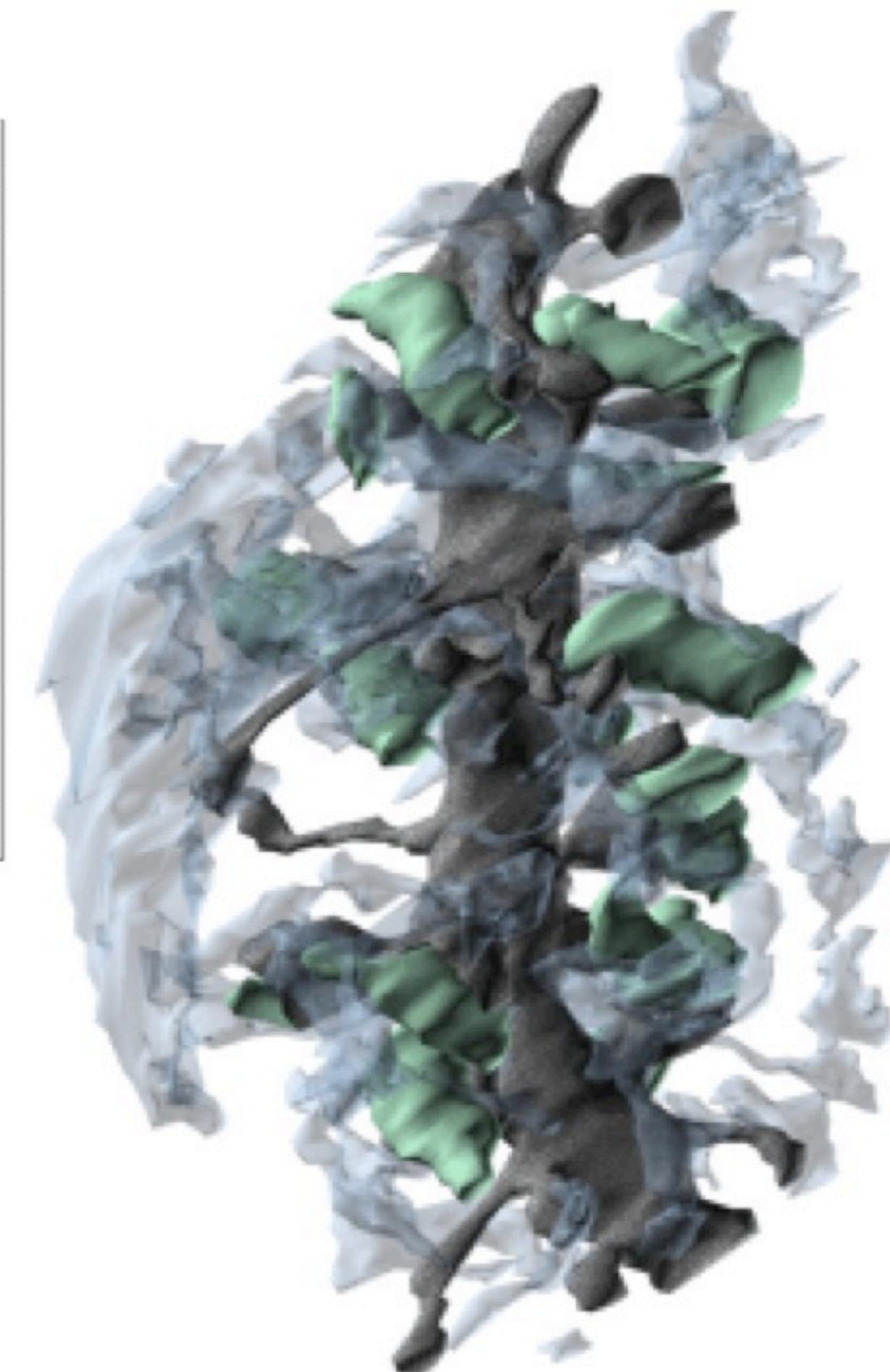
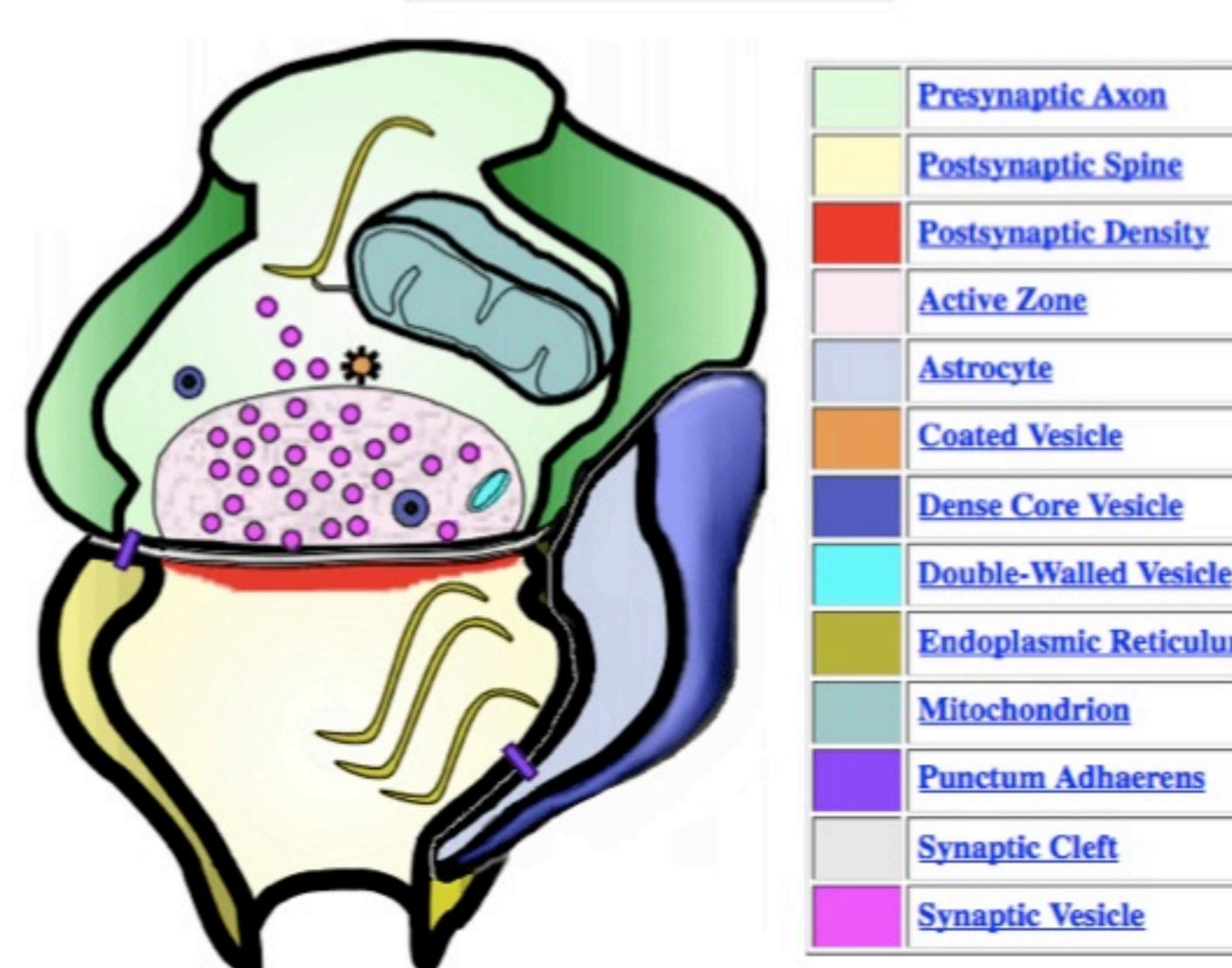


# Tripartite synapse with astrocyte



1 micron

A dendrite with synapsing boutons and neighboring astrocyte from CA1 stratum radiatum (series K18).



Presynaptic boutons

Dendrite & spines

Astrocyte

Front View

Side View

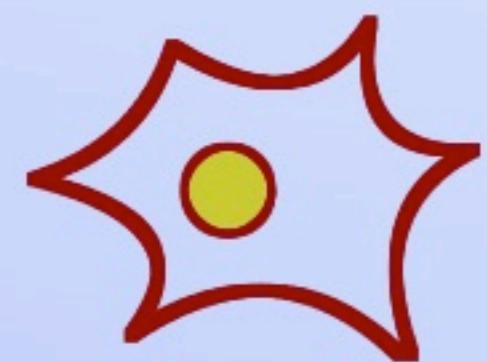
Back View

# 4. Role of membrane trafficking in neuritogenesis

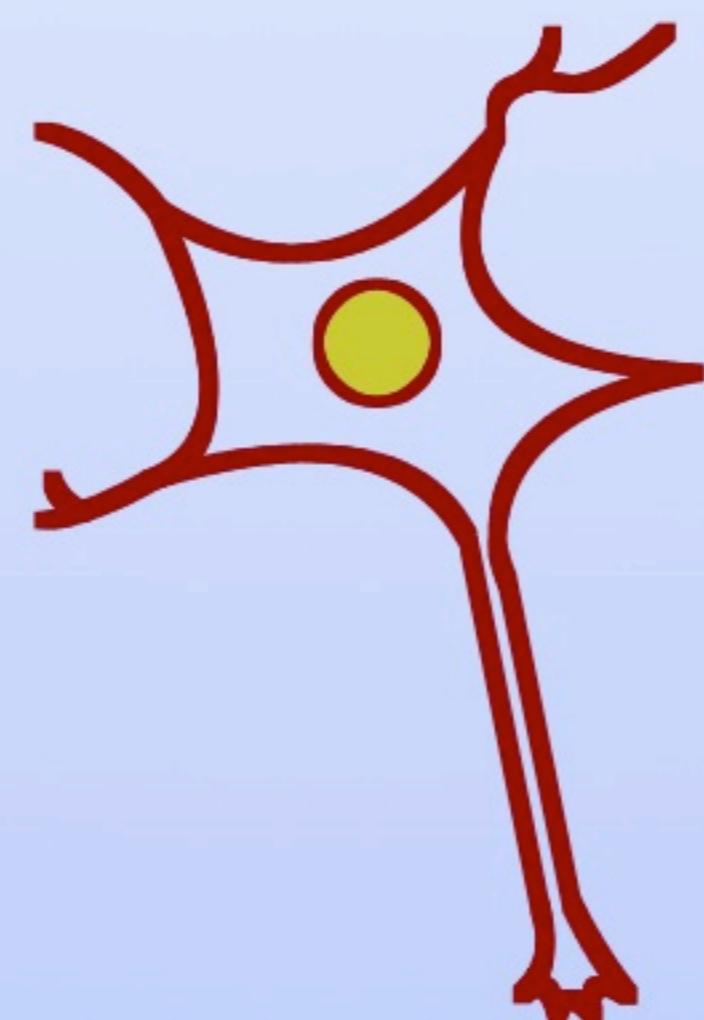
## I. Croissance membranaire

## II. Ciblage polarisé des molécules

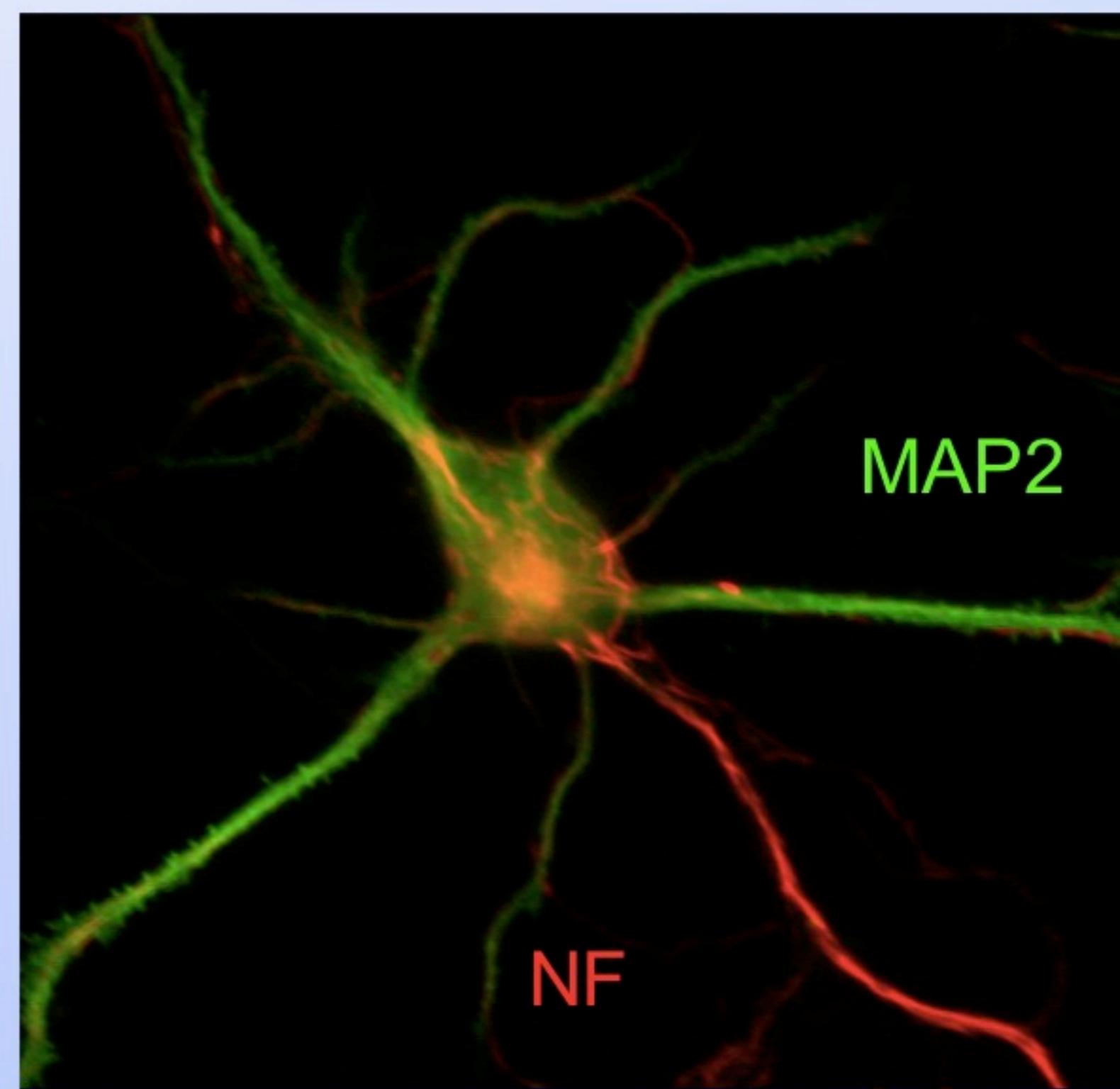
Précurseur



Neurone immature



Neurone mature



# 5. Role of membrane trafficking in neuritogenesis

## I. Croissance membranaire



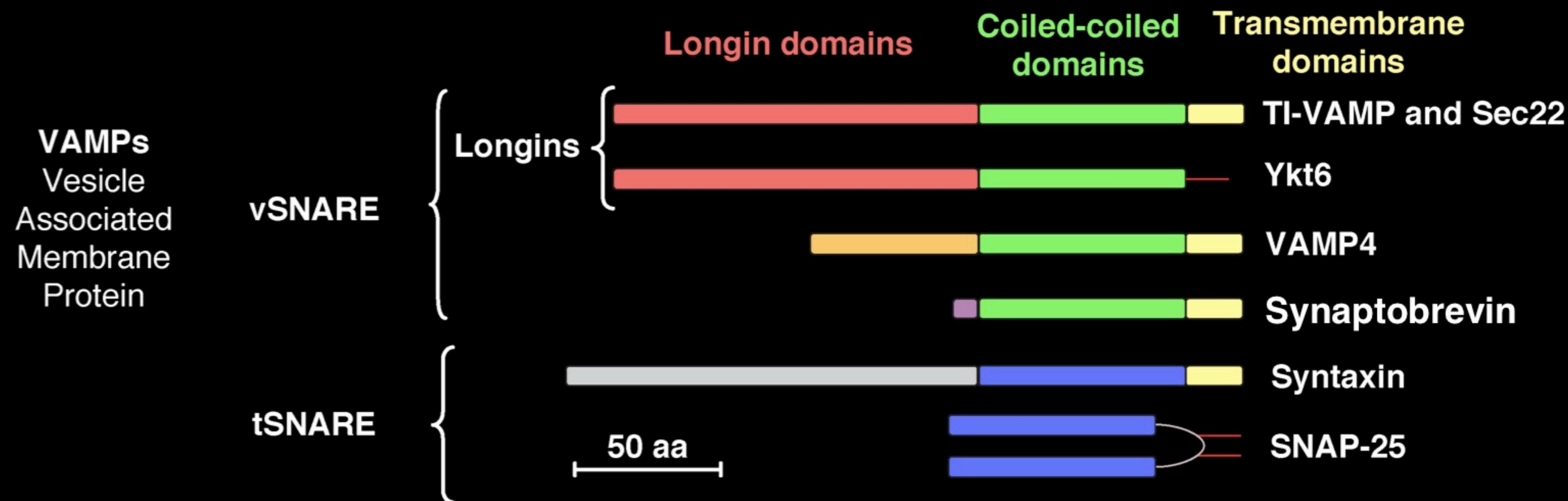
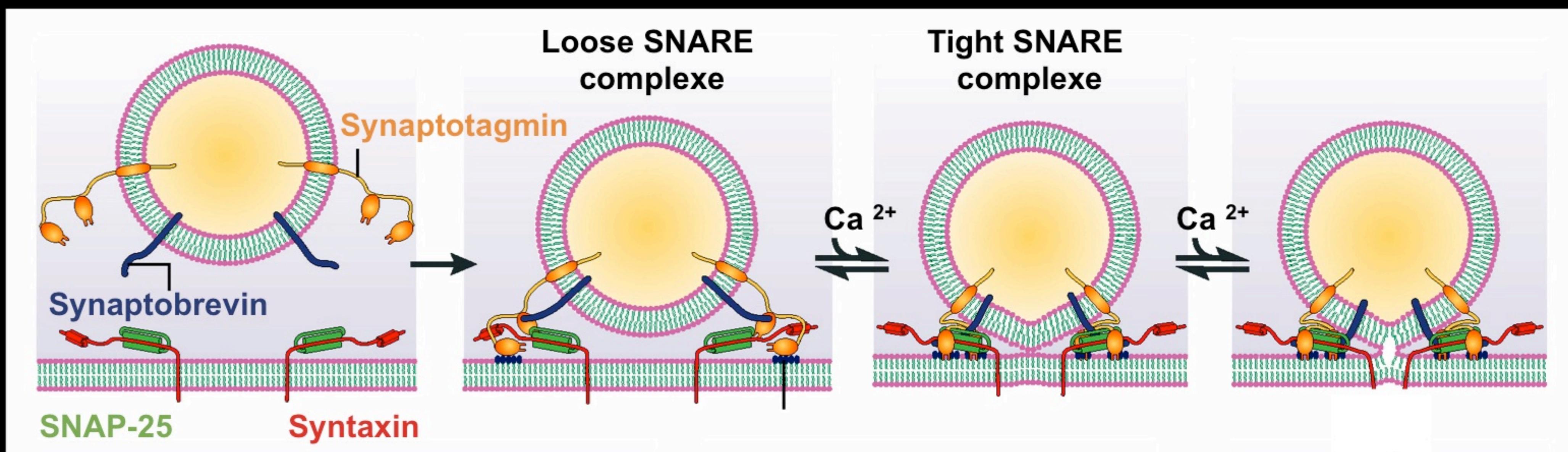
Phase contrast time-lapse recording of a neuron forming an axon. The video shows a rapid playback of a 16 hour recording with one image taken every 10 minutes.

[http://www.silvermanlab.org/gal\\_mov.html](http://www.silvermanlab.org/gal_mov.html)

Movie from <http://www.silvermanlab.org/>

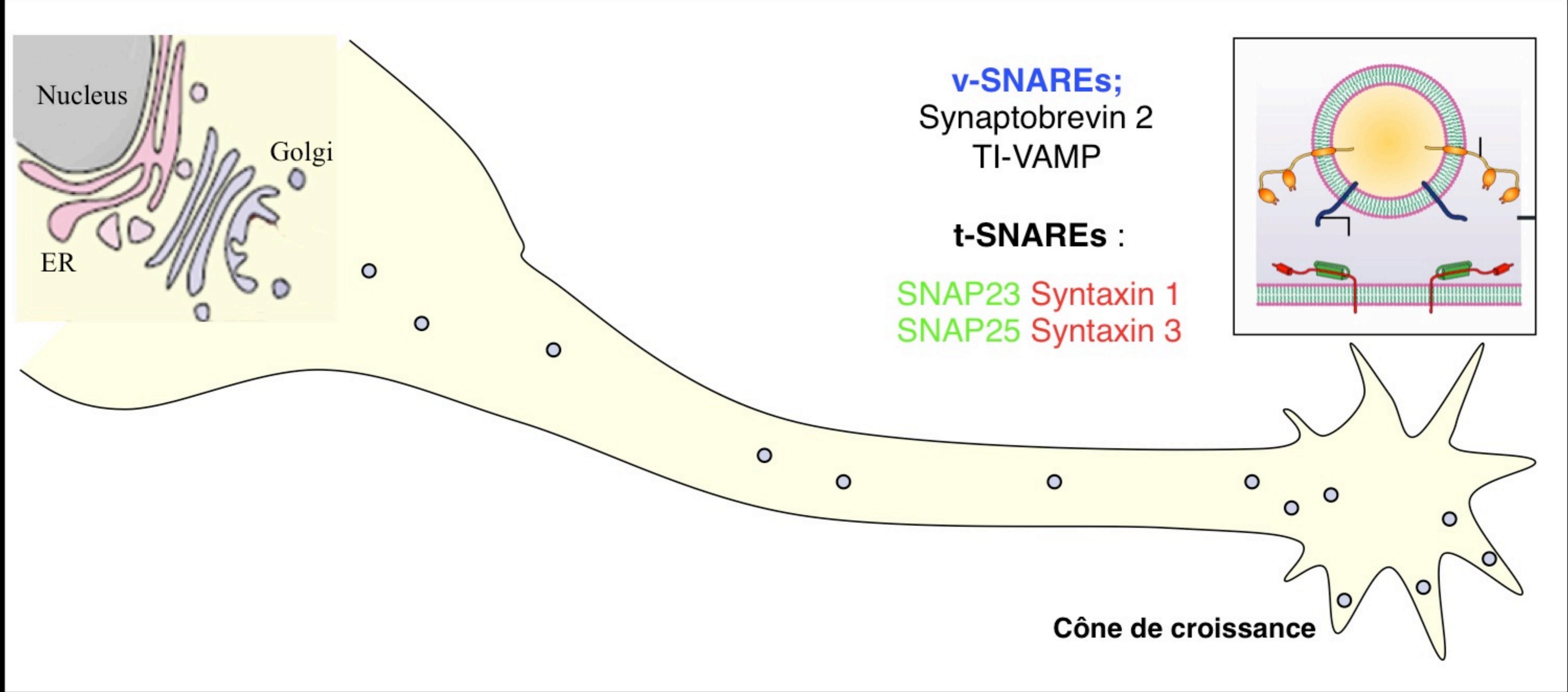
Quels sont les mécanismes permettant l'expansion membranaire ?

# Le complexe SNARE



Adapted from Rossi et al. (2004) TRENDS in Biochemical Sciences Vol.29 No.12:682-688.

# SNAREs in neurite outgrowth



## SNAREs à la synapses

v-SNAREs:      t-SNAREs :  
Synaptobrevin 2      Syntaxin 1  
                          SNAP25

Perte de Syb2 ou SNAP25:  
Perte de sécrétion évoquée  
Devpt cerveau normal, croissance neur. normale

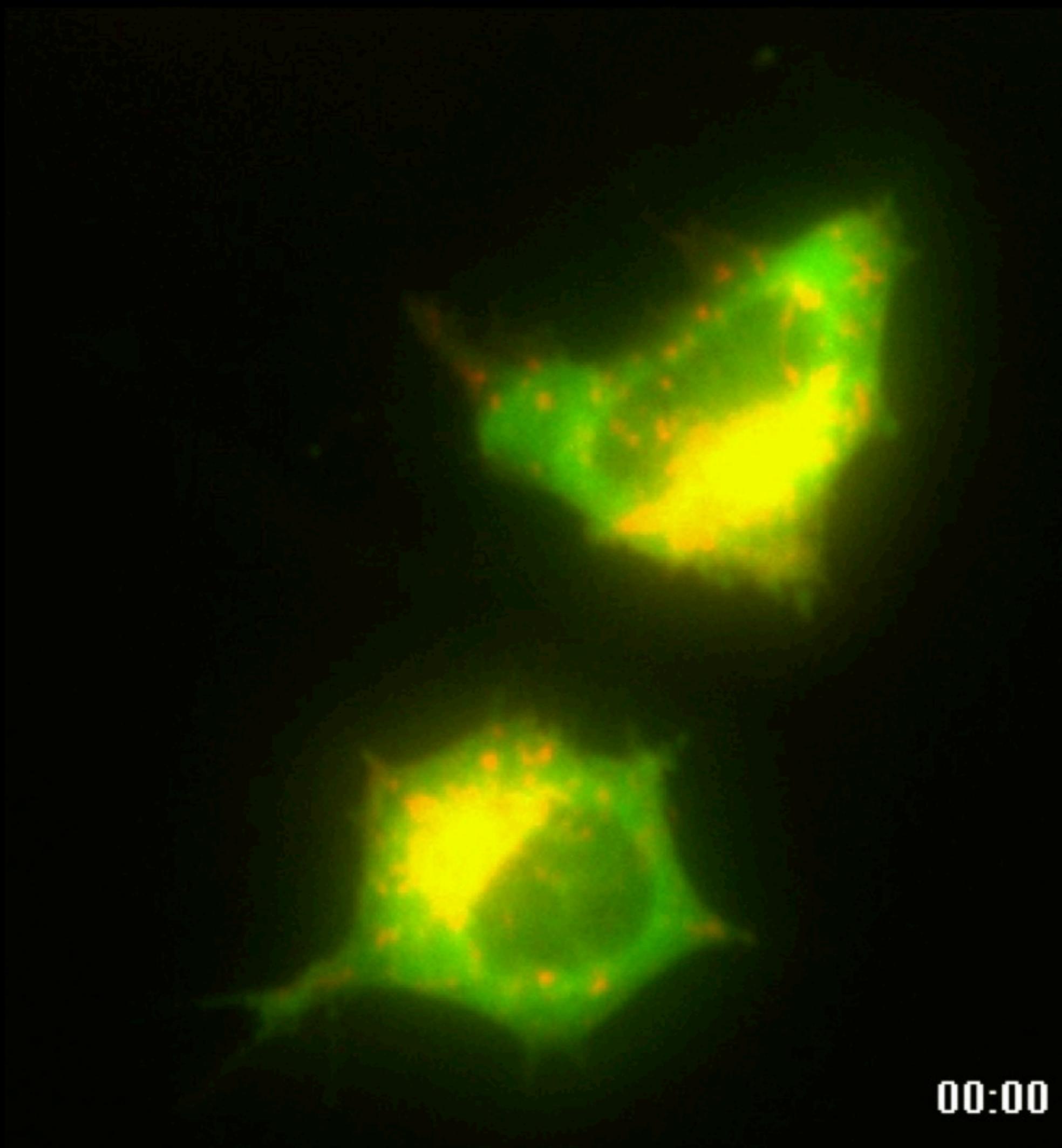
## SNAREs au cône de croissance

v-SNAREs:      t-SNAREs :  
Synaptobrevin 2      Syntaxin 1,3  
                          TI-VAMP      SNAP23,25

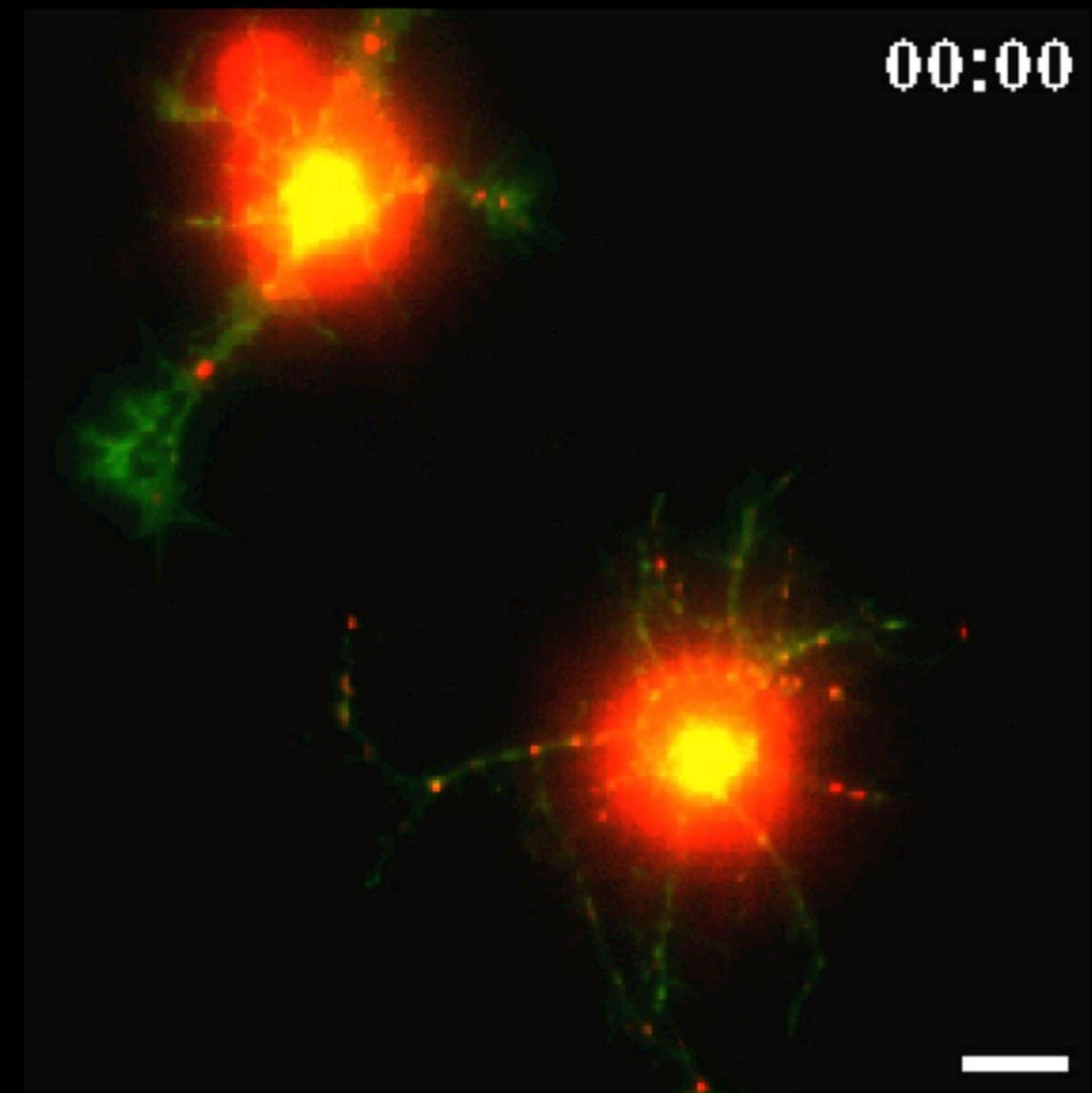
Croissance neuritique:  
Résistante à la TeNT qui clive Syb2  
Nécessite TI-VAMP et stx3

# TI-VAMP et la neuritogenèse

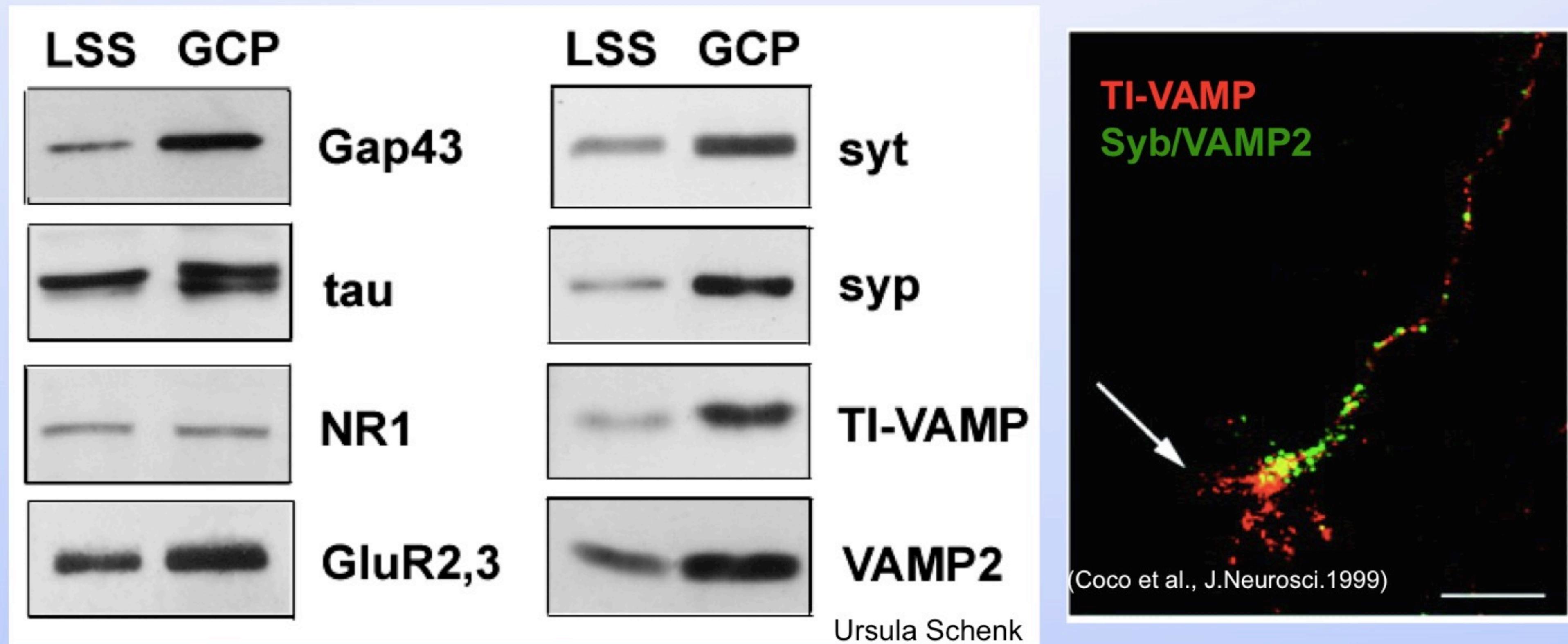
*PC12 cells*



*Hippocampal neurons*

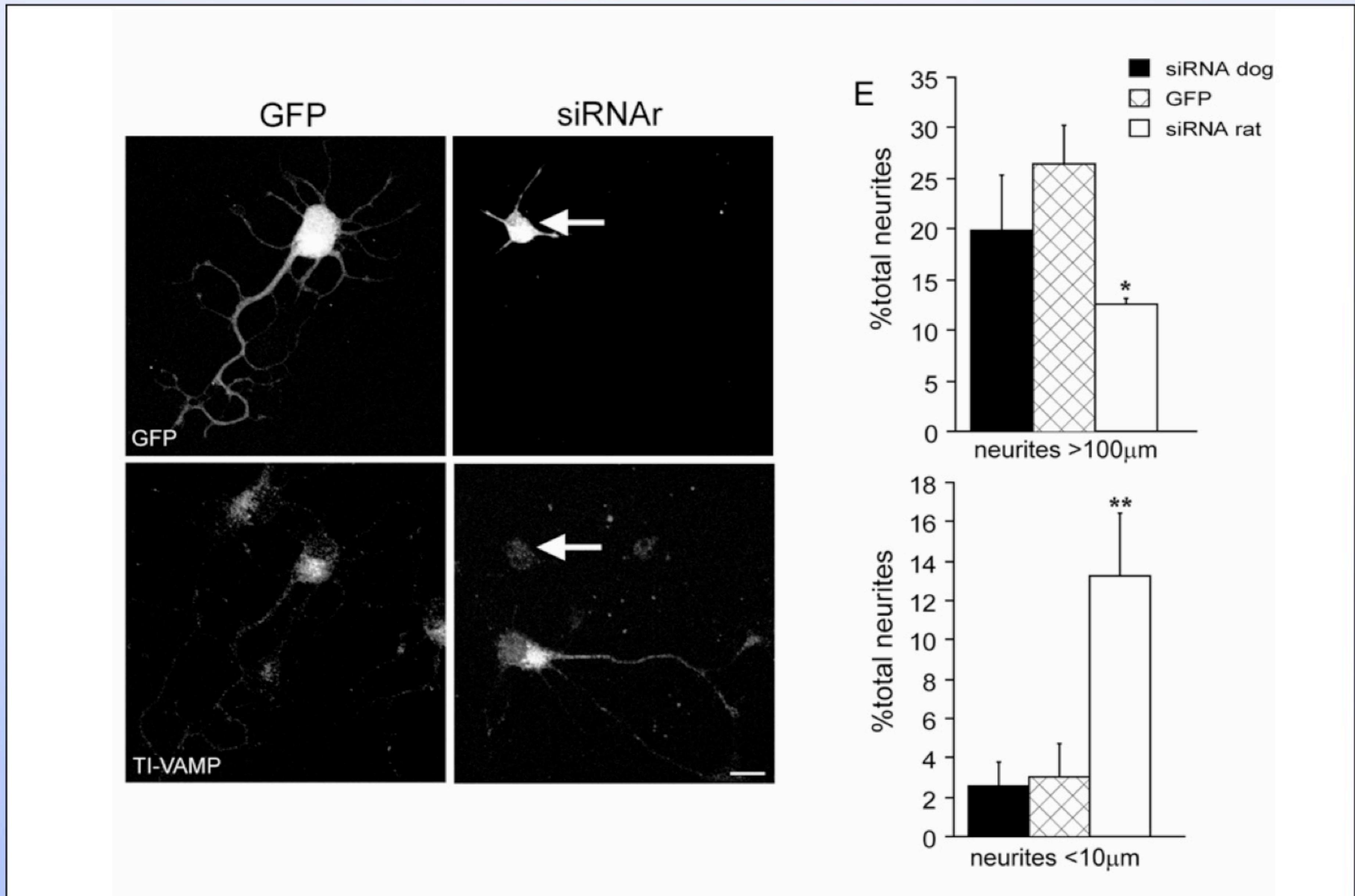


# TI-VAMP dans les cônes de croissance

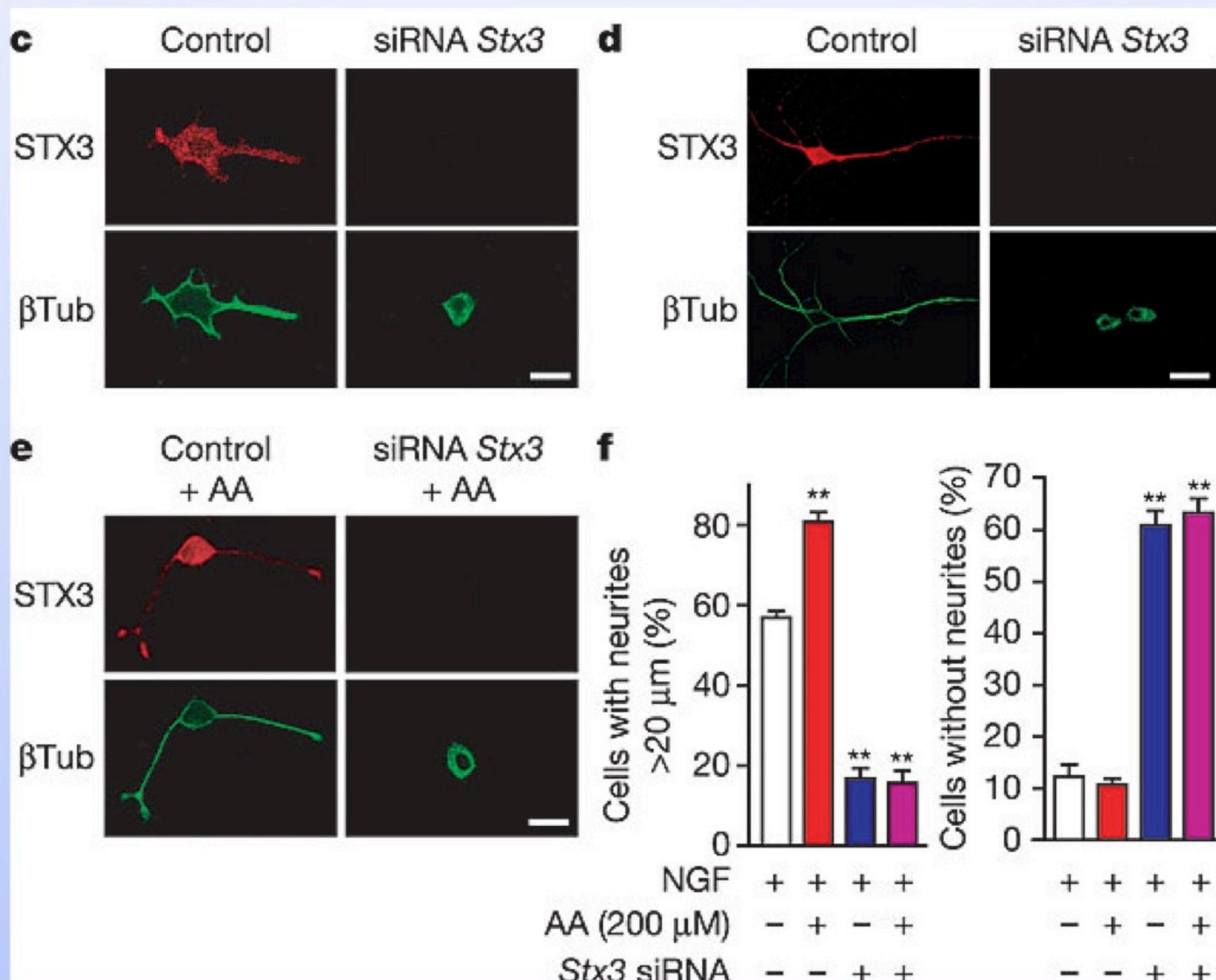


Les marqueurs présynaptiques sont enrichis dans les préparations de cones de croissance (GCP).

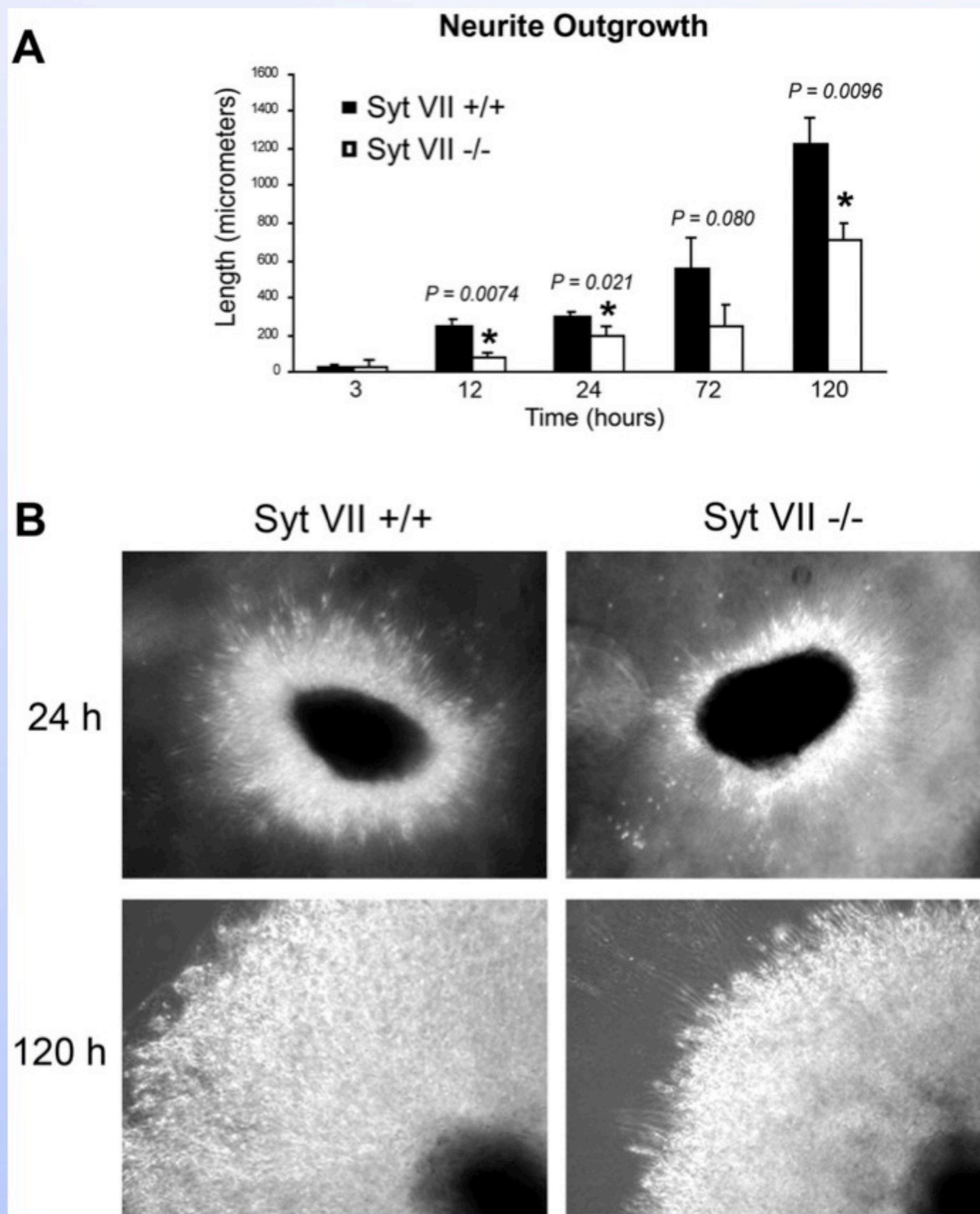
# TI-VAMP est essentiel à la croissance neuritique



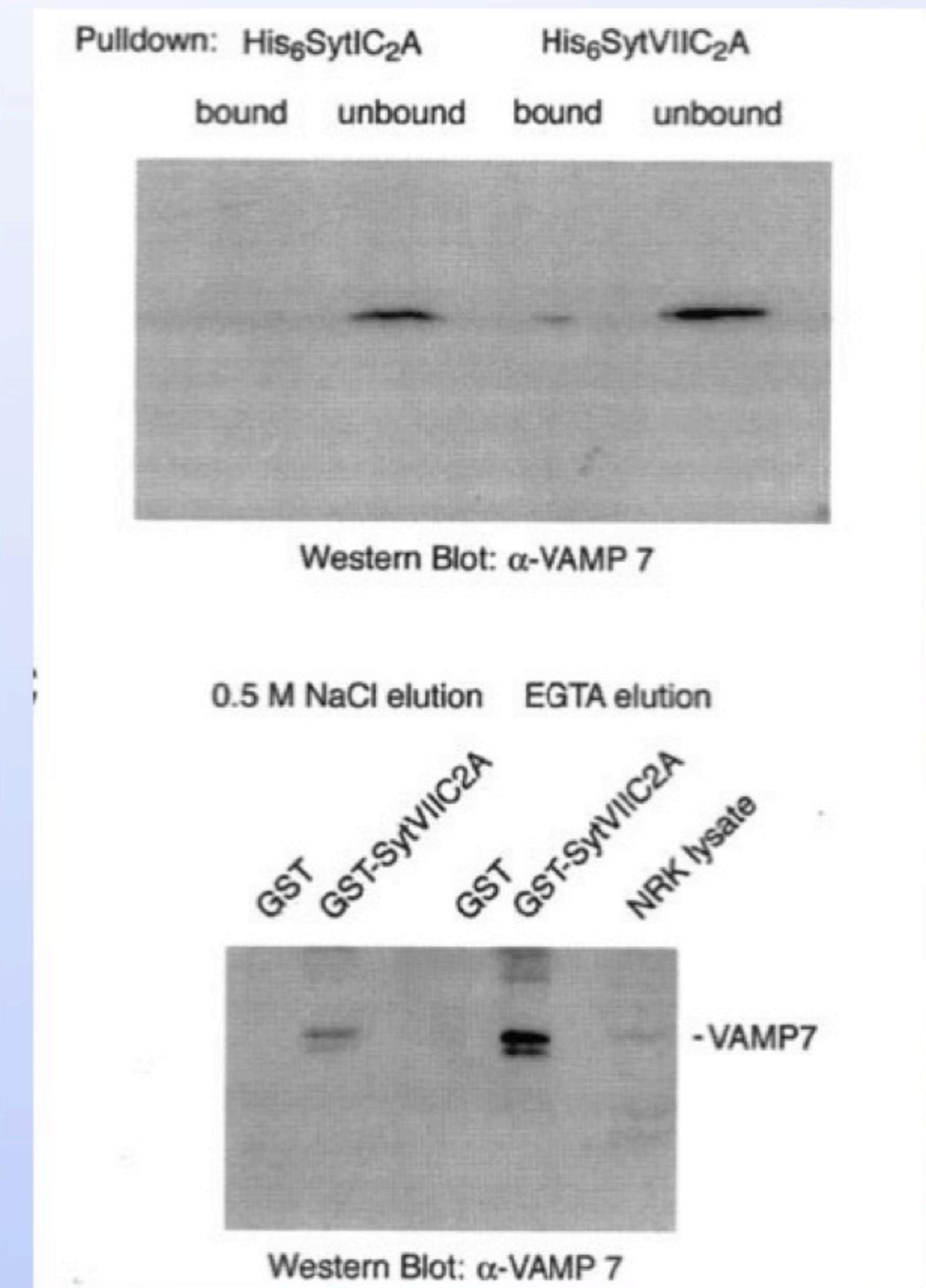
# Implication de la syntaxine 3 dans la croissance neuritique



# La croissance neuritique est diminuée dans les explants de ganglions spinal de souris SytVII -/-



Arantes & al J. Neurosci. 2006

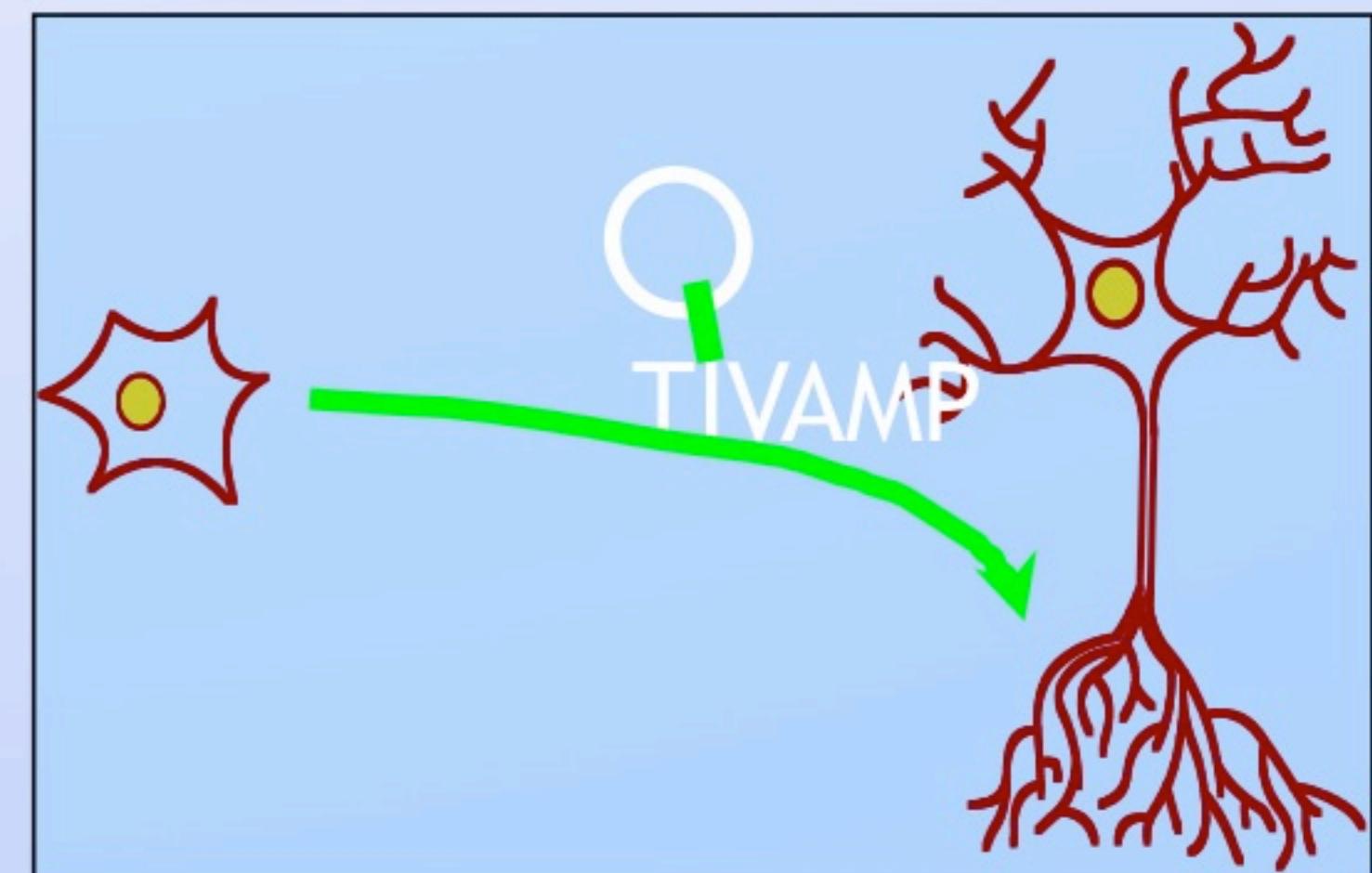
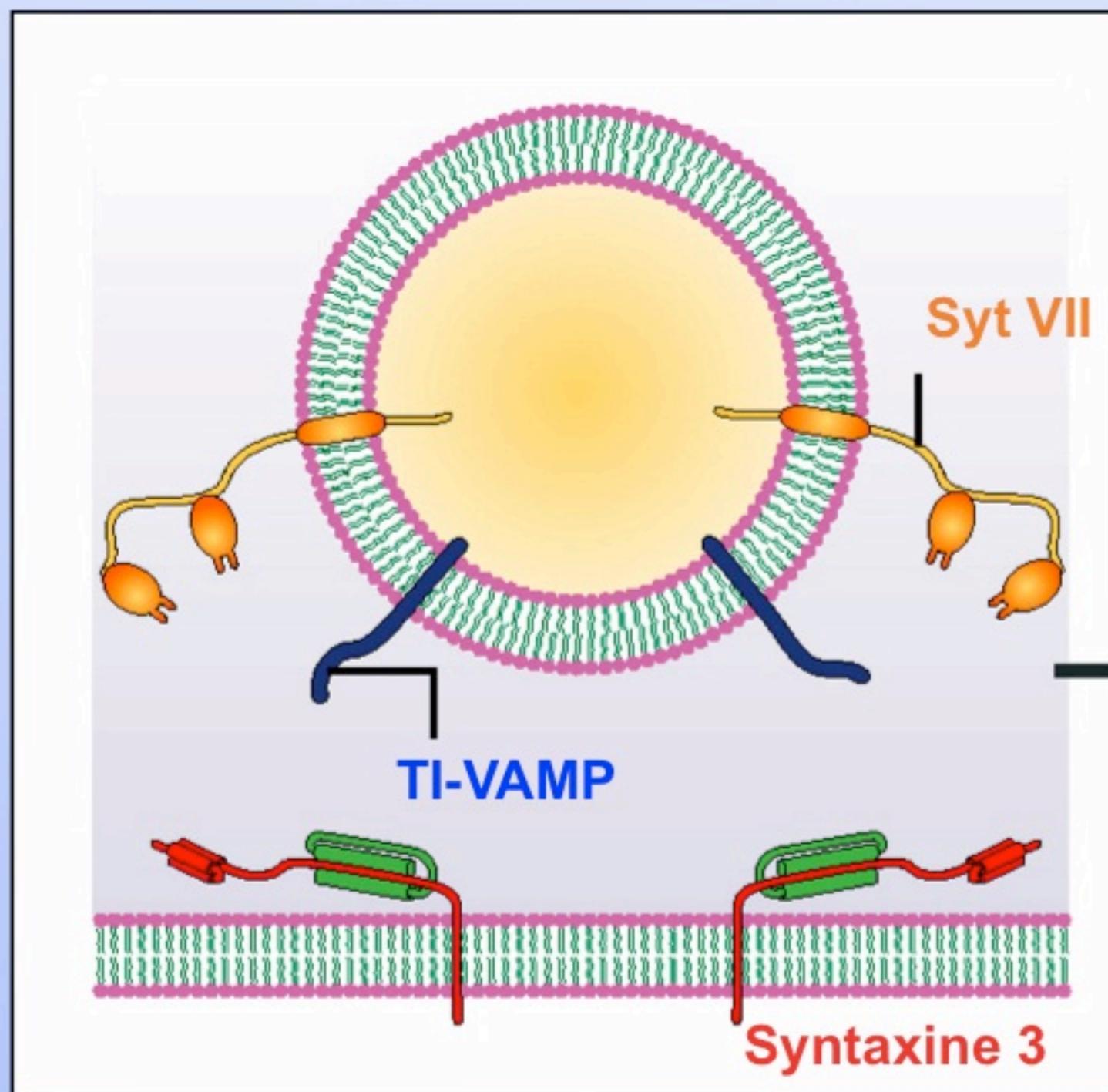


Rao & al JBC 2004

# Role de l'exocytose dans la morphogenèse neuronale

Mécanisme moléculaire impliquant:

- TIVAMP comme v-SNARE
- Syntaxine 3 comme t-SNARE
- Synaptotagmin VII



# Cargo de TI-VAMP: L1

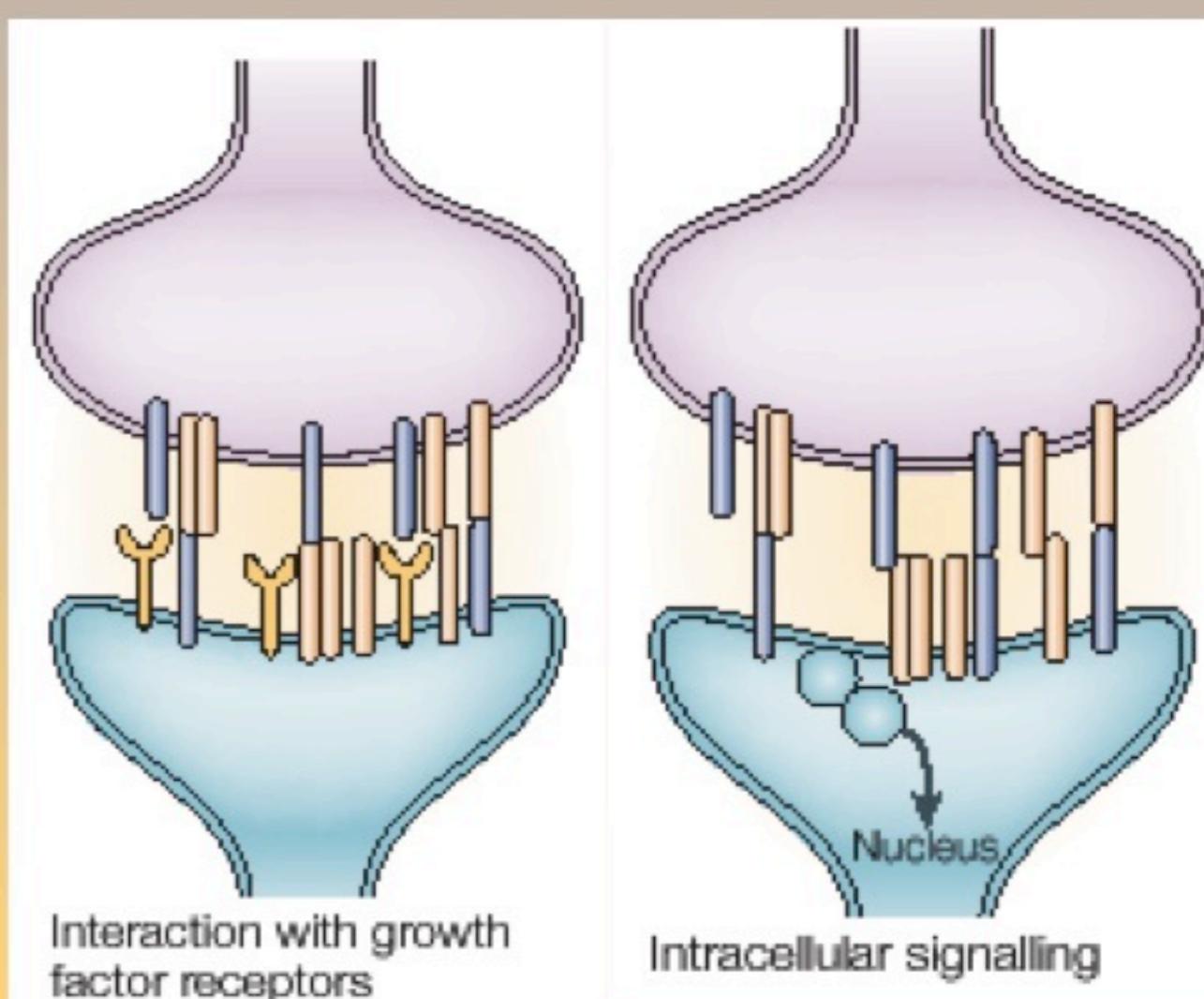
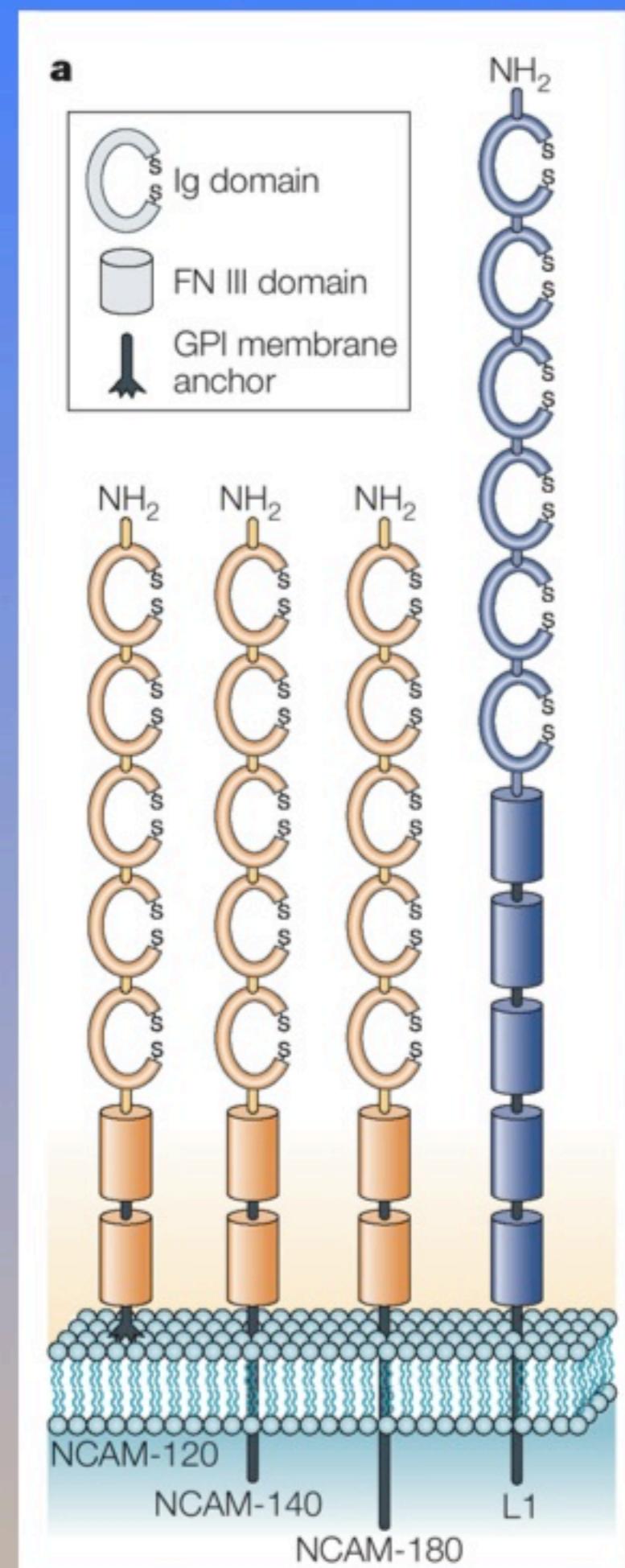
- Appartient à la famille des Ig-CAM:
- Maladies associées:
  - Large domaine extracellulaire avec 6 motifs Ig et 5 FIII
  - Un segment transmembranaire helicoïdal
  - Un domN cytoplasmique
- Rôle de L1:
  - Interactions cellulaires:
    - homo (L1-L1)
    - et hétérophilliques (L1-IgCAMs)
  - Liaison integrines et Recépteurs à tyrosine phosphatases.
  - Au cours du développement :
    - Différenciation cellulaire, survie.
    - Croissance neuritique, guidage axonal et synaptogenèse.
  - Chez l' adulte:
    - Maintien des contacts cellulaires
    - Plasticité synaptique et processus de mémorisation (LTP).

Rôle de L1:

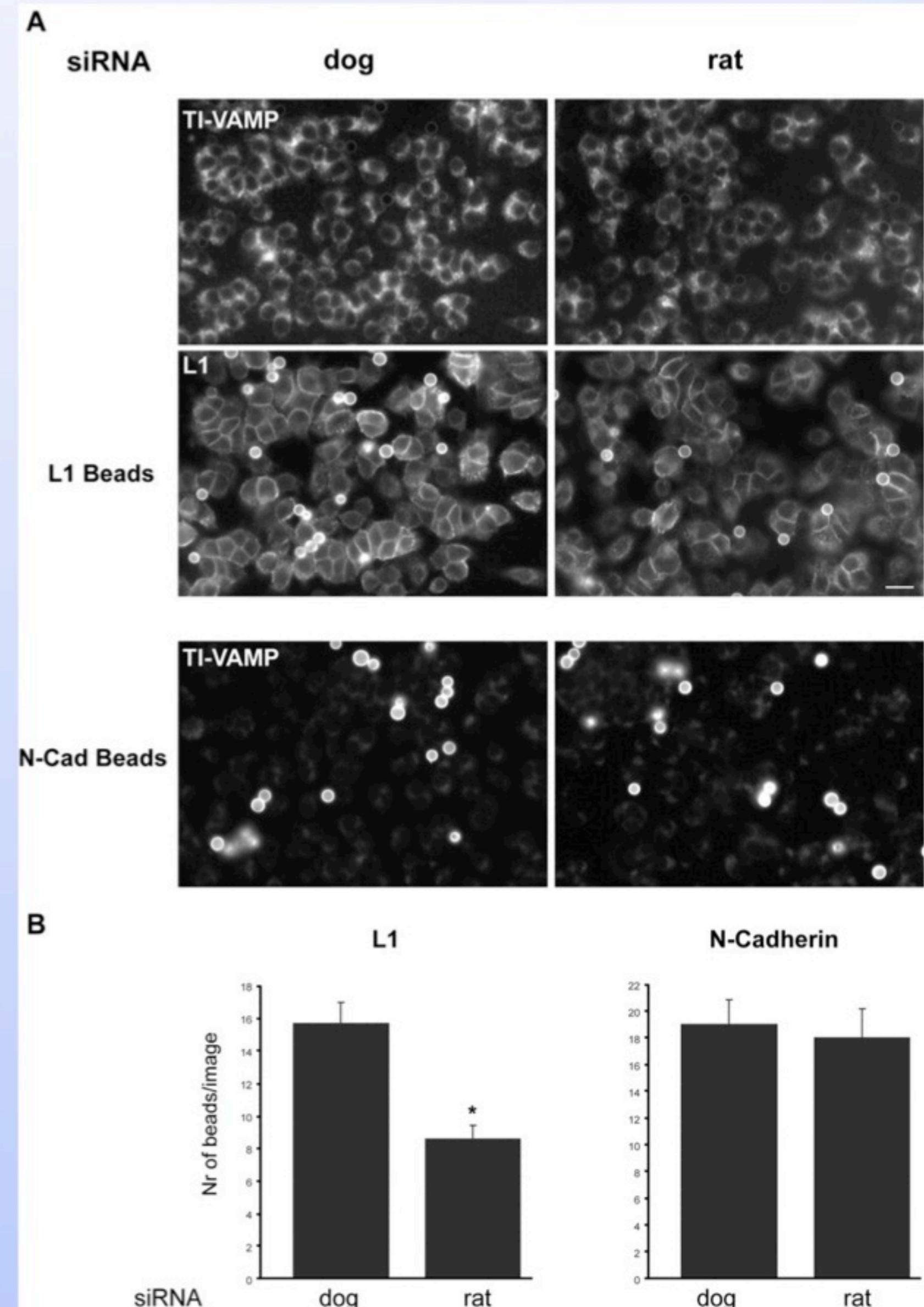
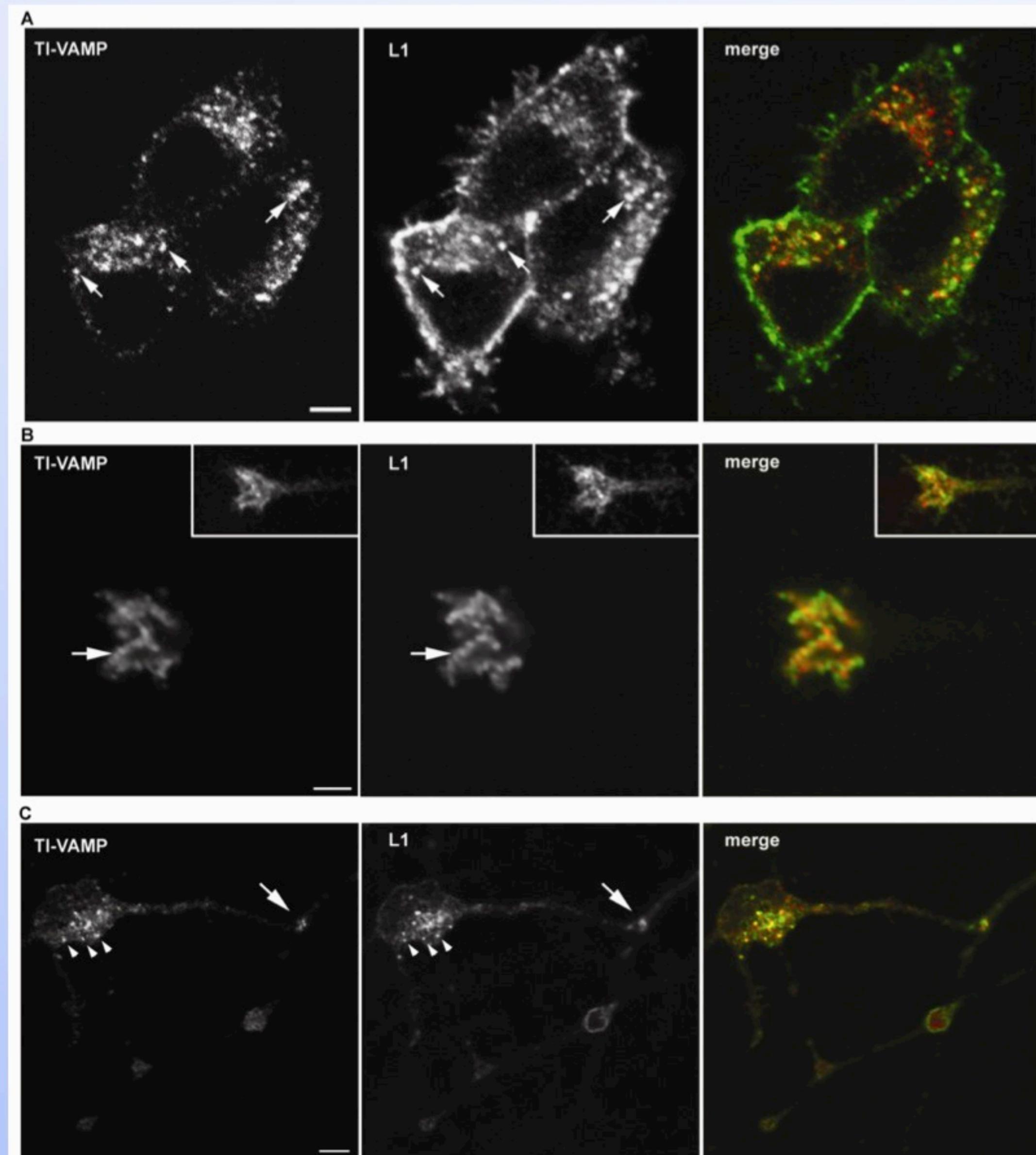
- Maladies associées:
  - Mutations chez l' homme (MASA syndrom):
 

dysfonctions neurologiques sévères:

    - Agenesis du tractus corticospinal, du corps calleux
    - Paraplégie spastique
    - Retard mental
  - Mutation chez la drosophile (neuroglian):
    - Lethalite embryonnaire, défaut de ciblage axonal.

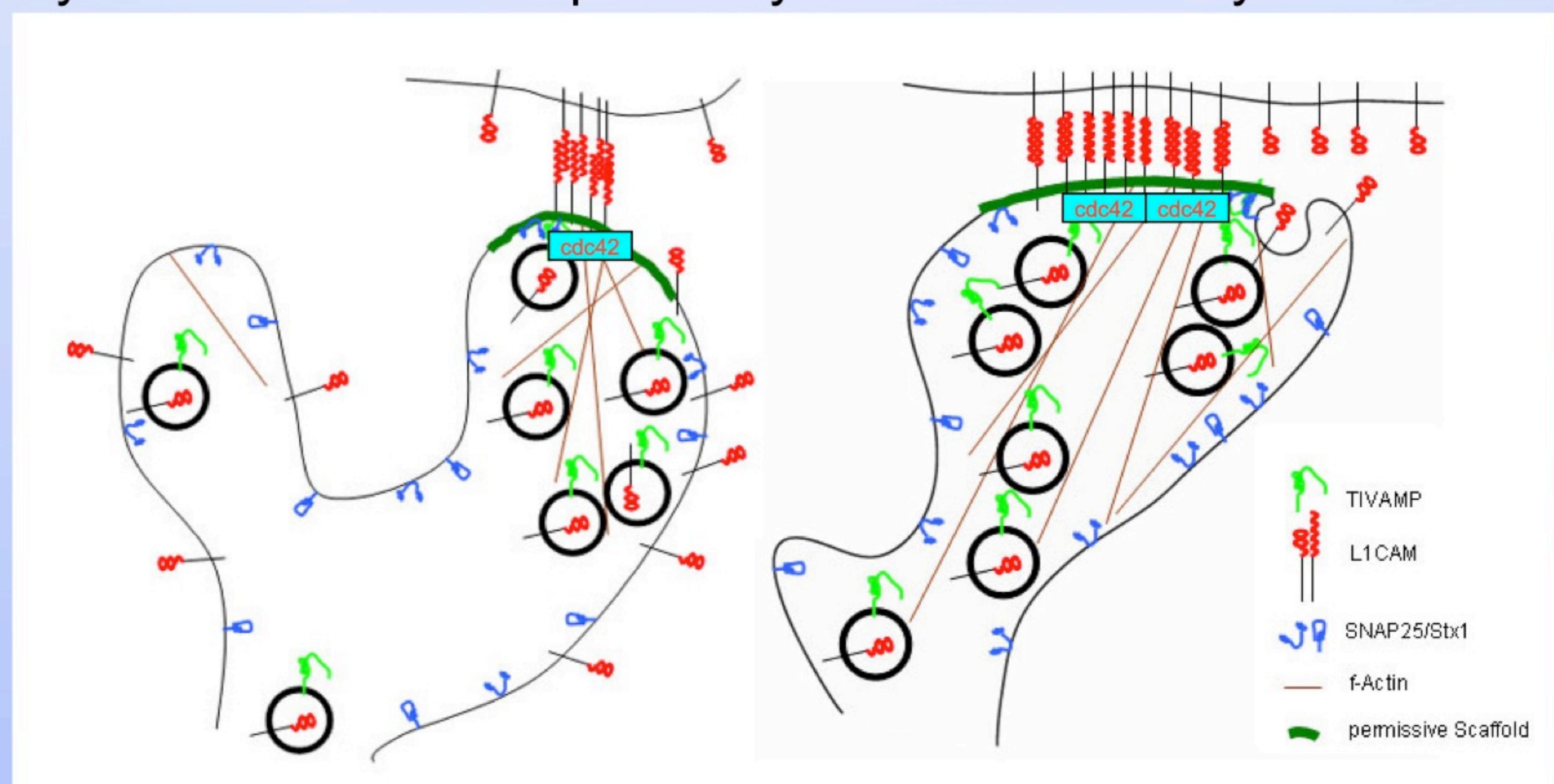


# *L1-CAM est une cargo de TI-VAMP*

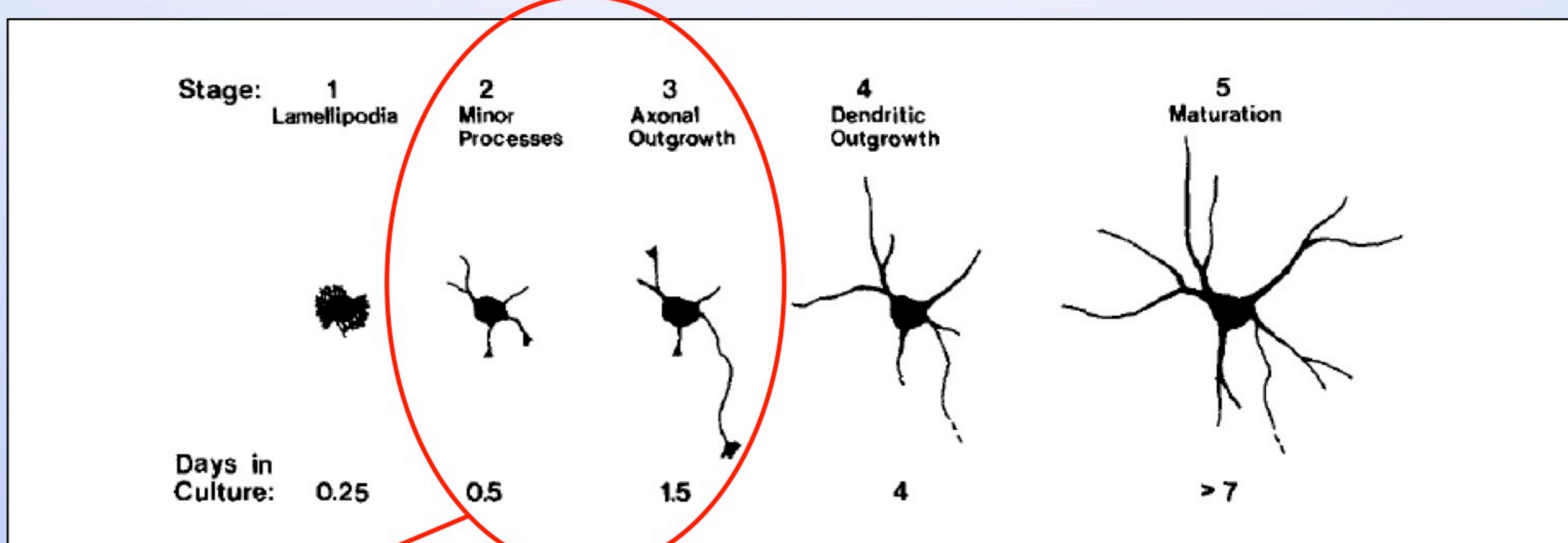


# Conclusion: Integration of signaling, actin, and exocytosis in neurite outgrowth

- TI-VAMP is a vesicular SNARE that is necessary for neurite outgrowth
- TI-VAMP transports the IgCAM L1. The TI-VAMP dependent membrane trafficking regulates the stability of L1-dependent adhesive contacts
- L1 mediated adhesion induces a polarization of TI-VAMP vesicles to sites of contact
- The exocytosis of TI-VAMP is positively controlled actin dynamics and cdc42



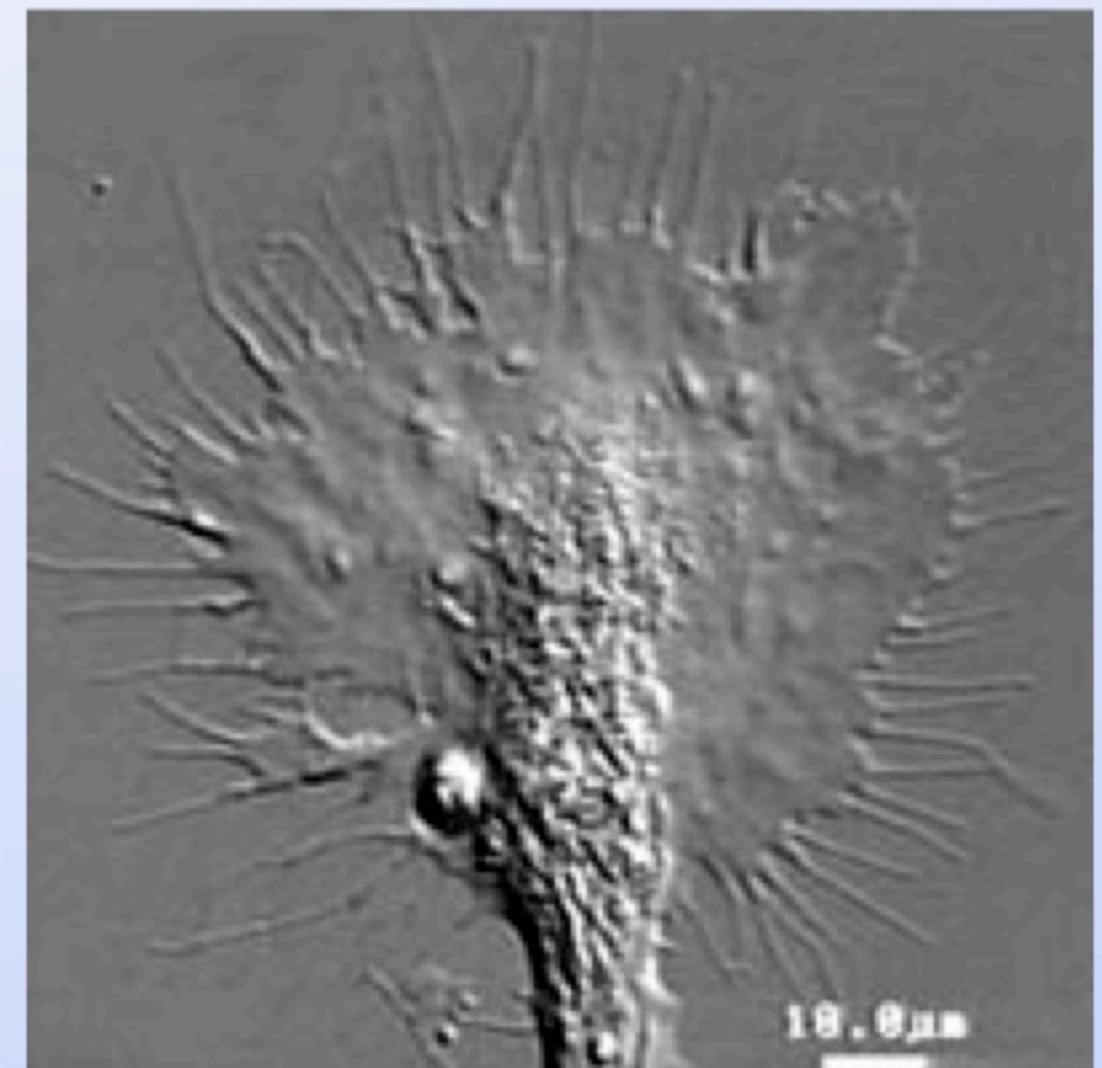
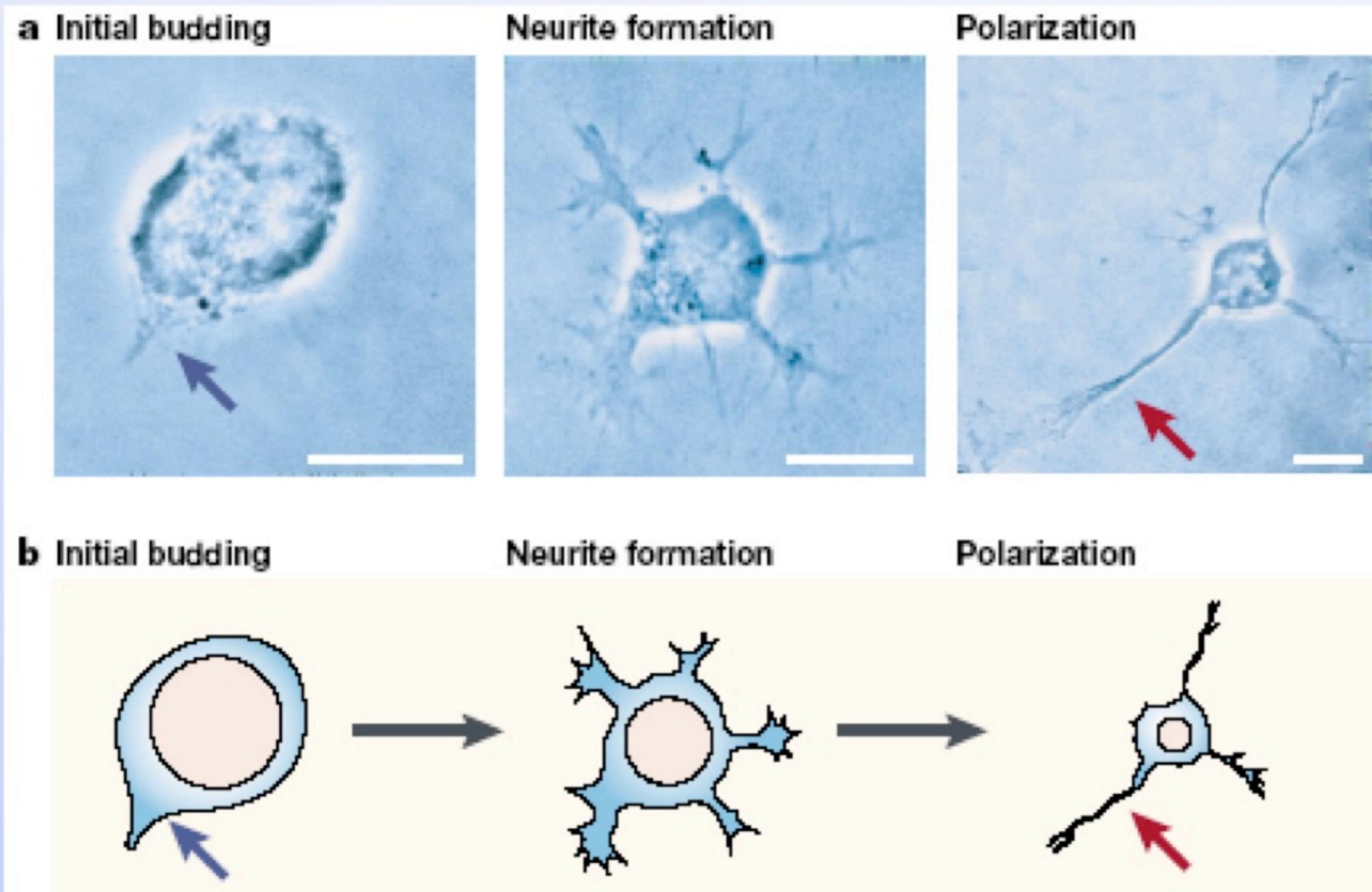
# Etudes pionnières *in vitro* par Gary Banker dans les années 1980's-1990's



Craig AM, Banker G. Annu Rev Neurosci. 1994;17:267-310.

L'acquisition de la polarité réside en **la sélection d'un neurite**, qui après une **croissance accélérée**, donnera l'axone pendant la transition **du stade 2 au stade 3**.

# Acquisition de la polarité entre les stades 2 & 3 dans les neurones d'hippocampe



Cône de croissance

da Silva JS, Dotti CG. Nat Rev Neurosci. 2002 Sep;3 (9):694-704.

Des études extensives ont montré que la croissance du neurite était étroitement contrôlée par la structure de son extrémité : **le cone de croissance**. La question de la polarité pourrait se simplifier à **la sélection du cône de croissance**.

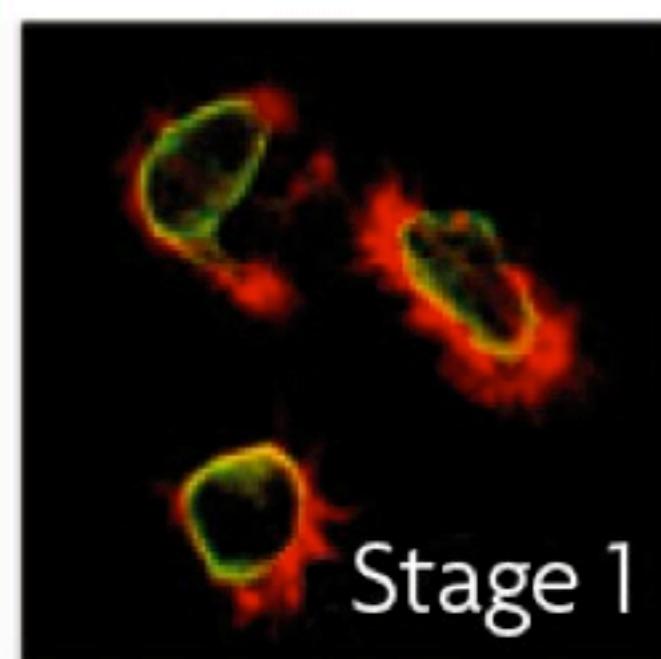
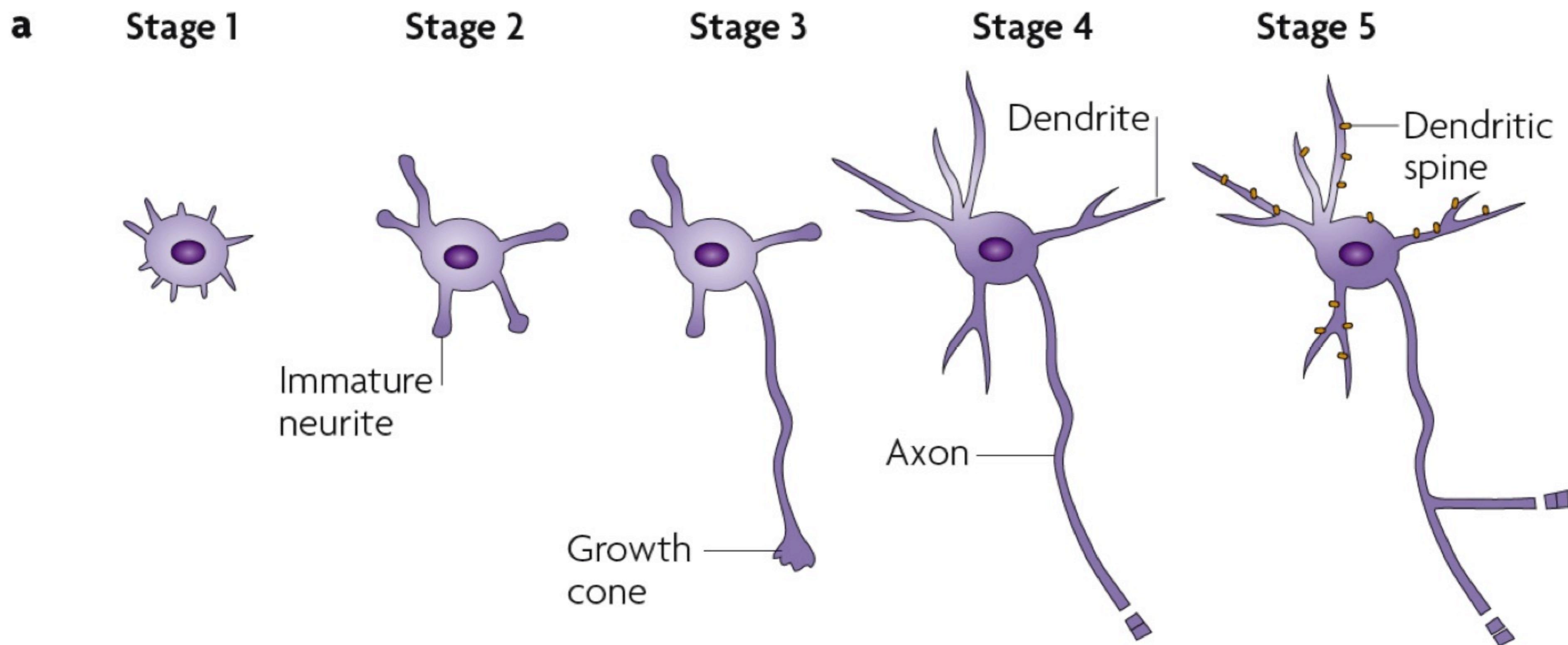
# Croissance neuritique:



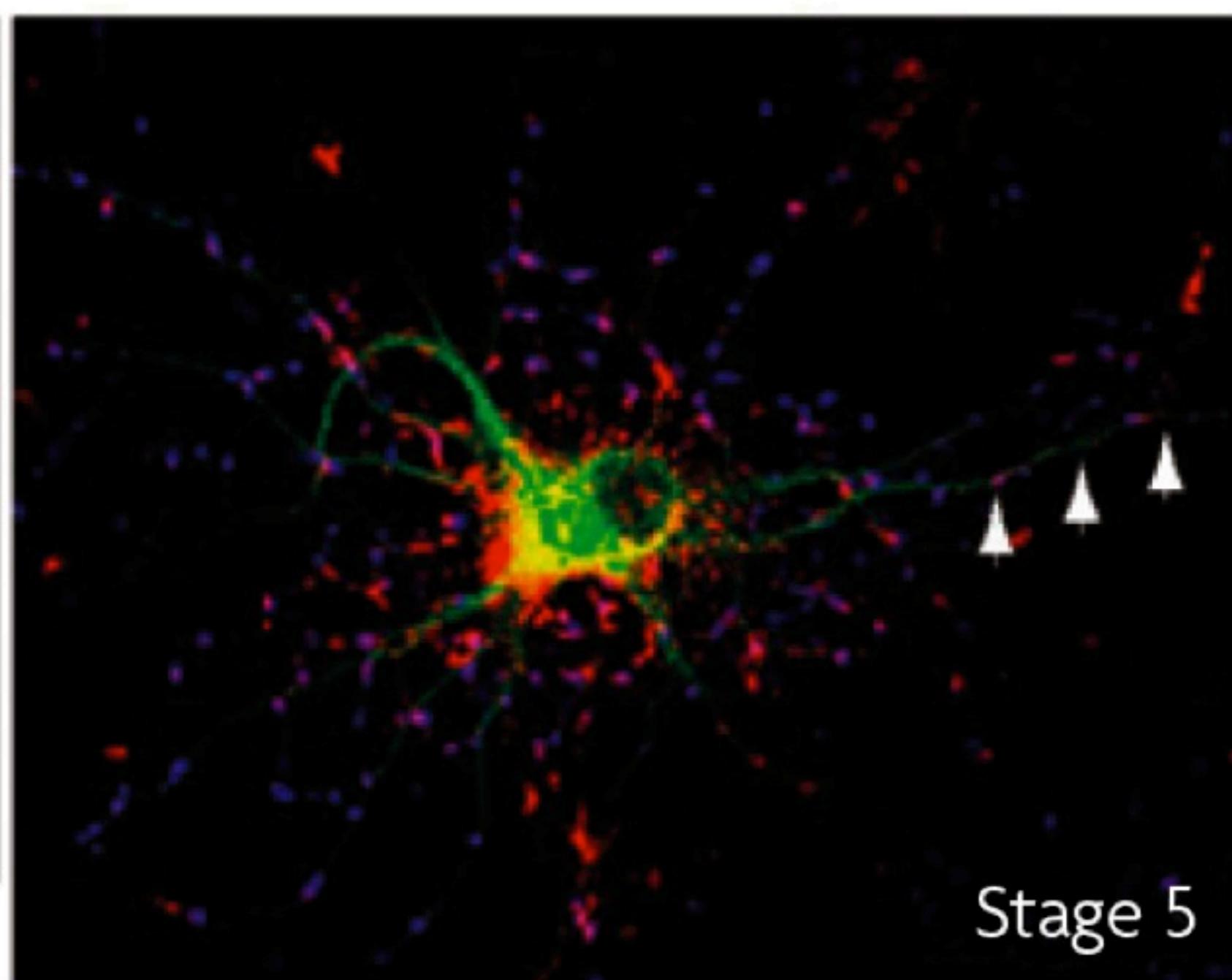
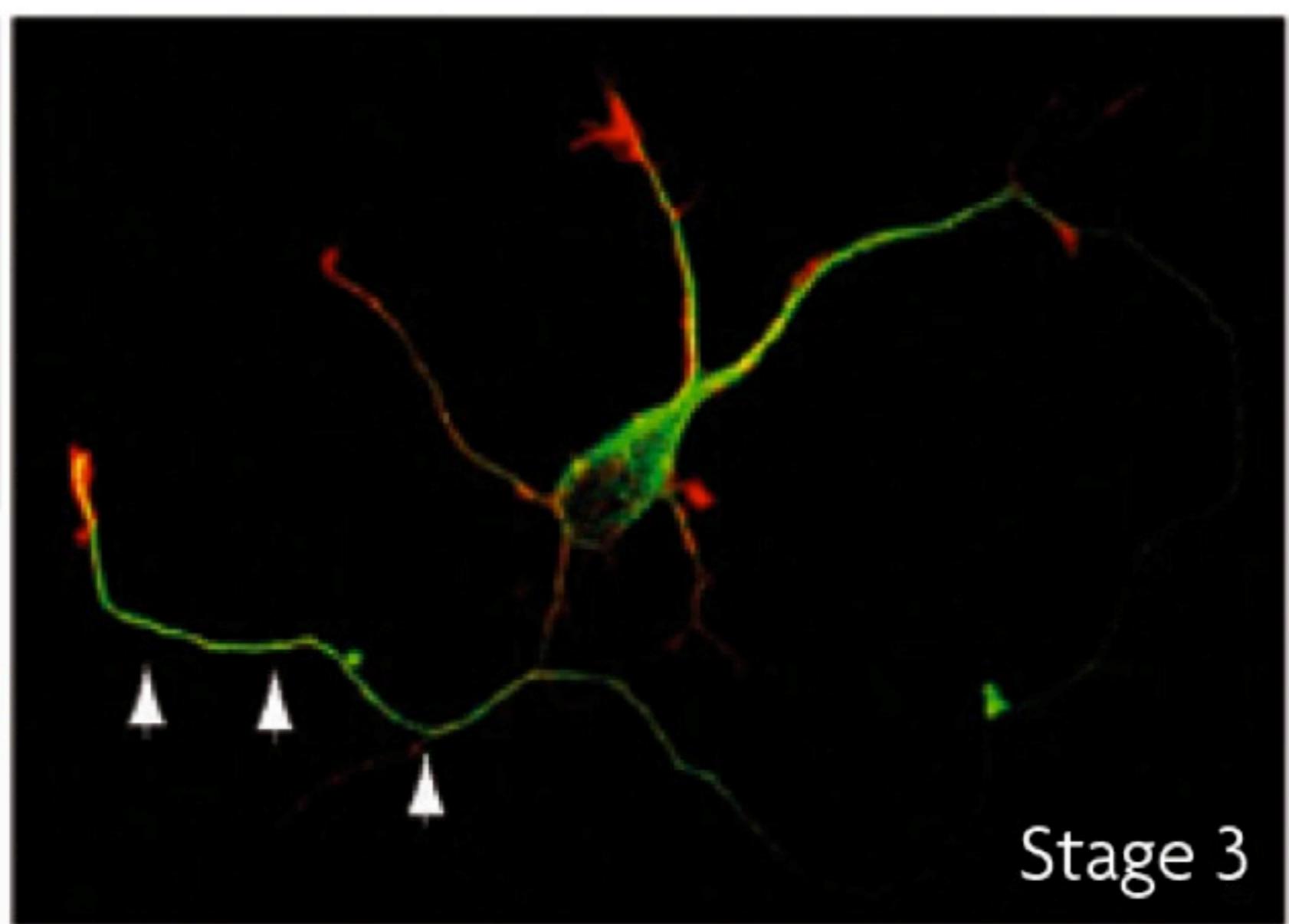
00:30:00

Elle est menée par 4 grandes étapes:

- Une augmentation de la membrane plasmique (recrutement de vésicules et fusion)
- Augmentation de la dynamique des filaments d'actine
- Augmentation de la formation des microtubules
- Augmentation locale de molécules de signalisation (PI3K, Rho GTPase,...)



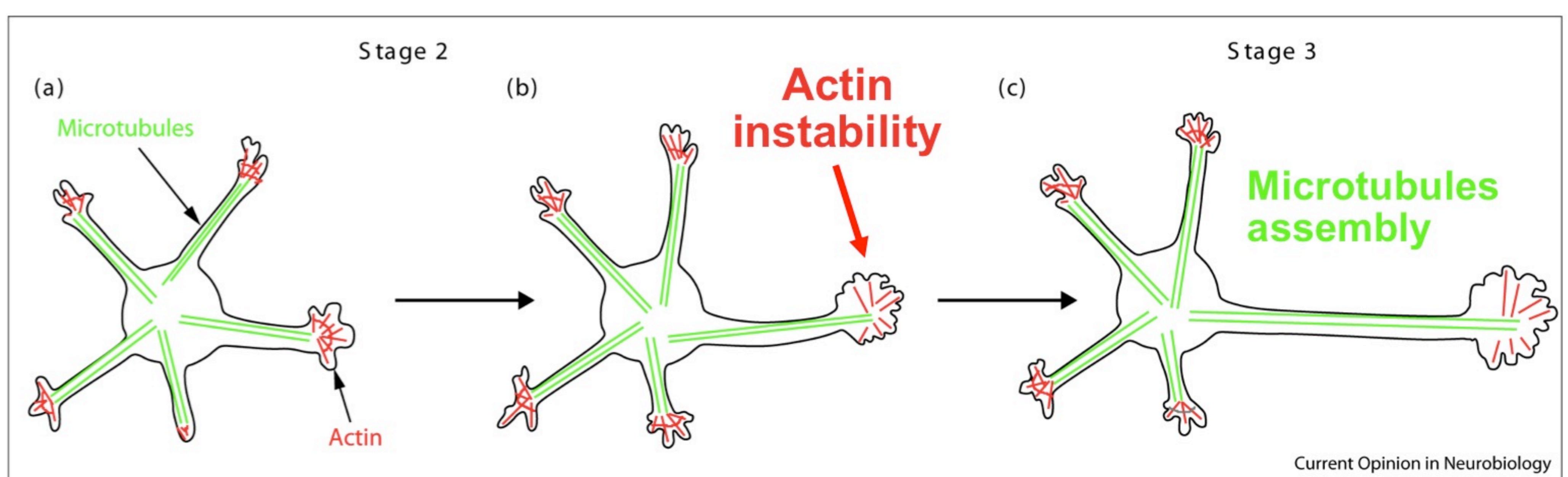
Actine  
Microtubules  
Synapsine



-The Role of Local Actin Instability in Axon Formation. Frank Bradke and Carlos G. Dotti. (1999) *Science* 283: 1931-1934

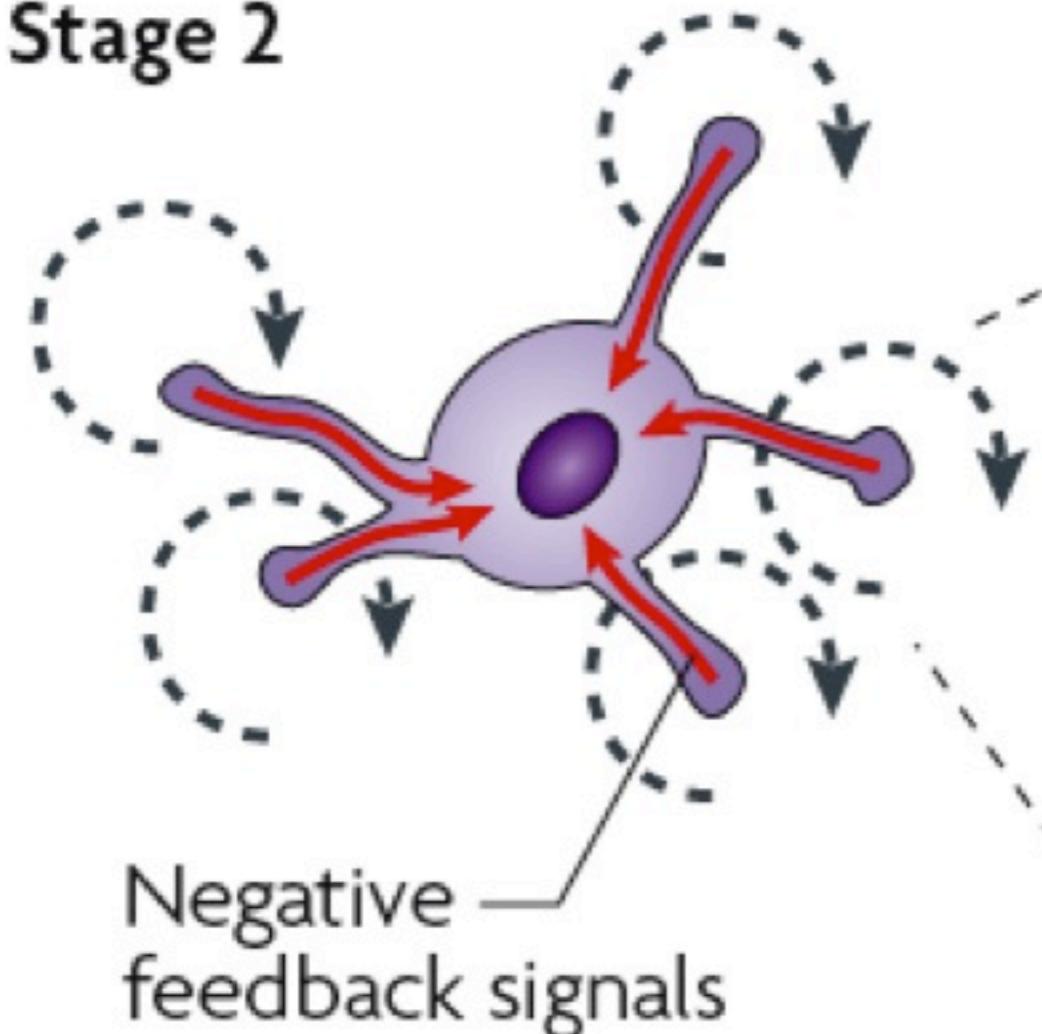
-Establishment of neuronal polarity: lessons from cultured hippocampal neurons, Frank Bradke and Carlos G Dotti (2000). *Curr Opin. Neurobiol.* 10: 574-581

Figure 3



Actin instability causes axon formation. (a) The growth cones of the different neurites of unpolarized stage 2 cells show similar activities over time early in development. The actin filaments comprise the peripheral area of the growth cones whereas the microtubules enter only at the central domain of the growth cone. (b) One of the growth cones of the neurites of morphologically unpolarized stage 2 cells

becomes highly dynamic and displays a less-stable actin cytoskeleton. The actin filaments appear to restrict the protrusion of the microtubules to a lesser extent. Thus, the microtubule may protrude and polymerize more distally. (c) Eventually, this neurite elongates by further polymerization/protrusion of microtubules enabled by a growth cone that has a high actin turnover. Thus, neuronal polarization takes place.

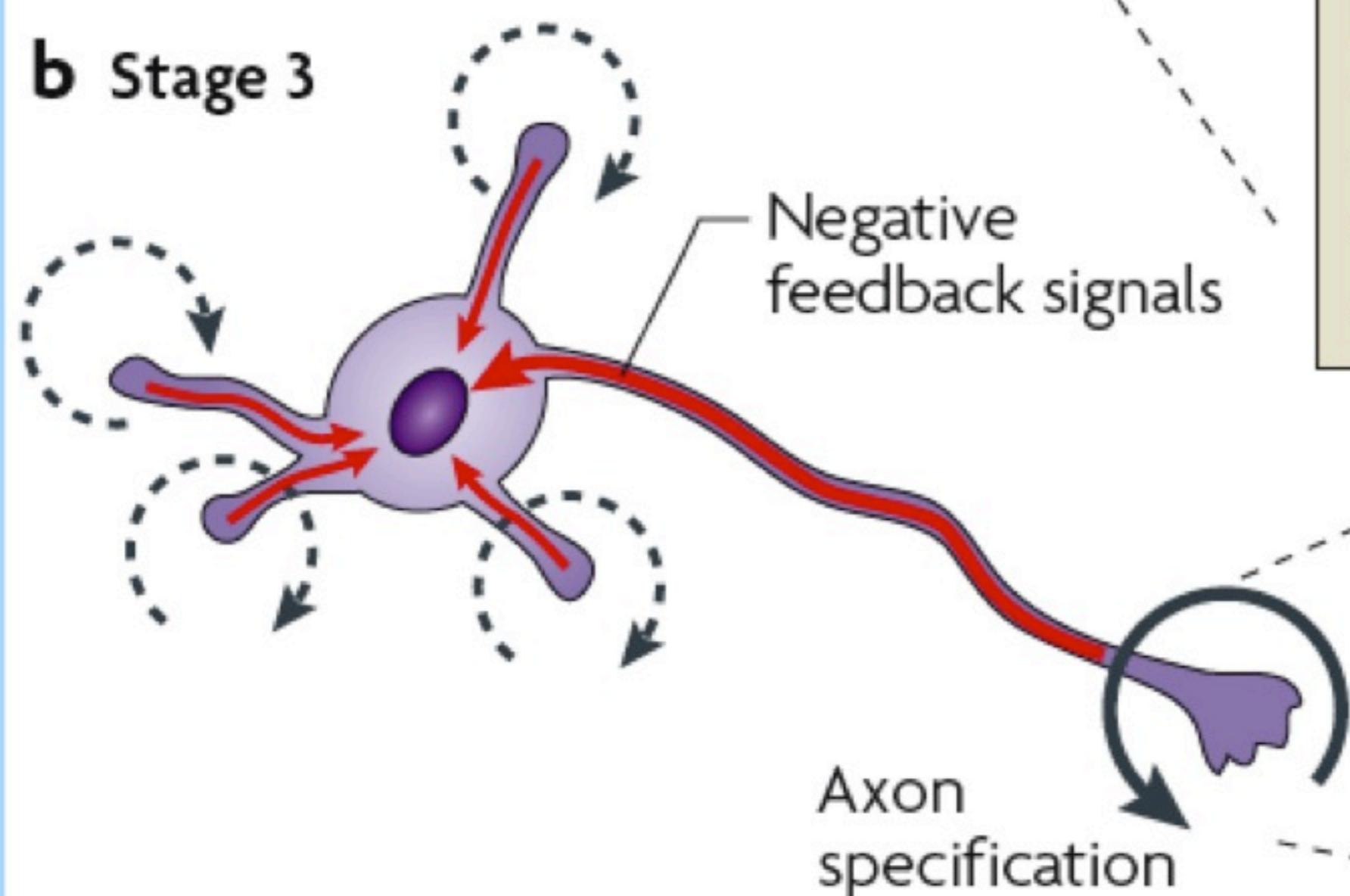
**a Stage 2****Negative regulation**

- Membrane elimination
- Degradation of proteins
- Decrease in dynamics of F-actin
- Microtubule catastrophe

**Retraction**

Phosphatase  
Rho GAP

Rho GTPases and GEF  
PI3K  
Centrosome

**Extension****b Stage 3****Positive regulation**

- Membrane recruitment
- Protein transport
- Increase in dynamics of F-actin
- Microtubule assembly

Extracellular signals,  
receptors,  
adhesion molecules.  
Transport of  
key regulators

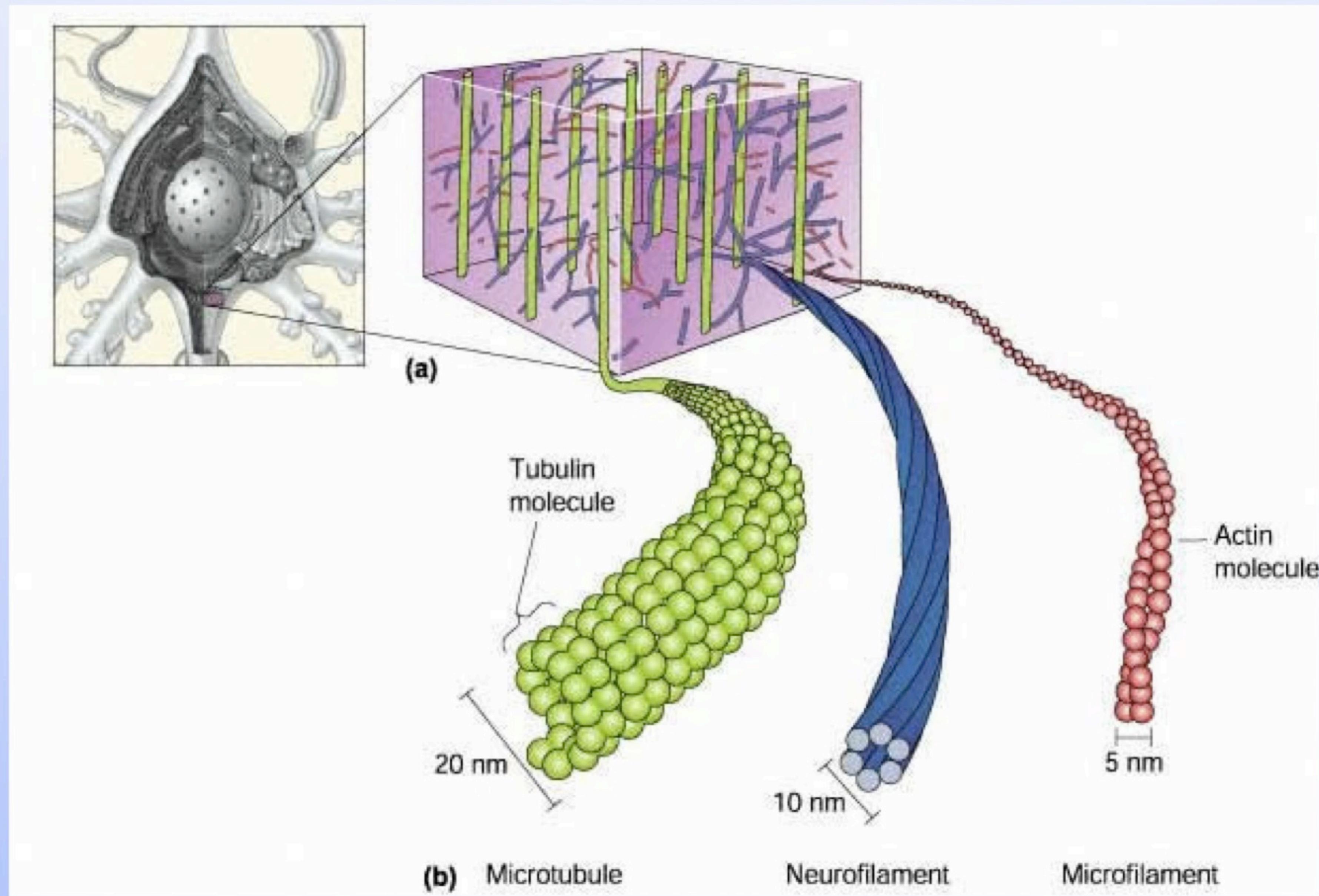
Positive  
feedback  
loop

Figure 2 | A tentative model for axon specification in neuronal polarization.

# Molécules régulant la polarité

	Protein (function)	Subcellular localization	*Effect of upregulated activity	*Effect of down-regulated activity	Criteria used to score effects	Association with other molecules	Refs lot
+	PI3K (kinase; produces PtdIns(3,4,5)P <sub>3</sub> )	Activated at the tip of future axon (st.2,3)	Myr-p110: multiple axons	Inhibitor (LY294002 etc.): no axon, or delay of polarization	Morphological and molecular (tau-1)	PtdIns(3,4,5)P <sub>3</sub> , Akt	9,25, 38,40
-	PTEN (PtdIns(3,4,5)P <sub>3</sub> degrading enzyme)	Uniform (EGFP-PTEN)	WT: unpolarized	siRNA: increased number of axons	Morphological and molecular (PAR3, tau-1, MAP2, pAkt, synapsin 1, GAP43)	PI3K, Akt, GSK3β	25,38
+	R-Ras (small GTPase)	Single neurite (st.2); axon (st.3)	WT, DA (Q87L): multiple axons	siRNA: unpolarization; R-RasGAP-Myr: prevention of axon formation	Morphological and molecular (tau-1, synapsin 1, GAP43, FM4-64 uptake, MAP2, pAkt, pGSK3β)	GSK3β, Akt	51
+	H-Ras (small GTPase)	ND	WT, DA (V12): multiple axons	DN(N17): no axon	Morphological and molecular (tau-1)	PI3K, MEK, GSK3β, CRMP2	40
+	RAP1B (small GTPase)	Tip of single minor process (st.2); tip of axon (st.3)	DA (V12): supernumerary axons, or reduced number of minor processes	siRNA: loss of polarity, no axon	Morphological and molecular (tau-1, MAP2, PAR3, p-aPKC, pAkt)	PI3K, Cdc42, PAR3, PAR6, aPKC, Akt	26
+	Cdc42 (small GTPase)	Tip of single minor process (st.2); tip of axon (st.3)	DA (L28): supernumerary axons; V12: unpolarized	siRNA: no axon; N17: no effect	Morphological and molecular (tau-1, MAP2, PAR3, p-aPKC, pAkt, synapsin 1)	PI3K, PAR3, PAR6, aPKC, Akt	26,63
+	Rac1 (small GTPase)	ND	DA (V12): increased the length of minor neurite, or unpolarized	DN (N17): reduced the length of minor process, or unpolarized	Morphological and molecular (tau-1, MAP2, synapsin 1)	STEF	26,63
+	Akt (mediates the signals of growth factors)	Tips of minor processes (st.2); tip of axon (st.3)	WT, Myr-Akt: multiple axons, or increased number of neurites	shRNA: neuronal death	Morphological and molecular (tau-1, MAP2, pAkt, synapsin 1, GAP43)	PI3K, Ras, GSK3β, tau, MAP1B	38,40, 42
-	GSK3β (glycogen synthesis)	Tips of all neurites (st.2); tip of axon (st.3); pGSK3β: tip of axon (st.3); cell body	WT, S9A: no axon or unpolarized	shRNA, inhibitor GID5-6 peptide: multiple axon GSK3α/β knock-in mice: no effect	Morphological and molecular (tau-1, MAP2, pAkt, synapsin 1, GAP43, synaptophysin)	PAR3, PI3K, Akt, CRMP2, tau, BDNF, NT3, MAP1B, APC	38,39, 41,42, 83
+	PAR3 (involved in asymmetric cell division and cell polarization)	Tips of minor processes (st.2); tip of axon (st.3)	WT, 4N-1: unpolarized, or multiple axon-like neurites	delC, delN: no axon, or increased length of minor processes	Morphological and molecular (tau-1, MAP2, synapsin 1)	PAR6, aPKC, STEF	25,26, 63
+	PAR6 (involved in asymmetric cell division and cell polarization)	Tip of axon (st.3)	ND	WT, delCREB: unpolarized	Morphological and molecular (tau-1, MAP2, synapsin 1)	PAR3, aPKC, Cdc42, STEF	25,26, 63
+	aPKC (involved in asymmetric cell division and cell polarization)	ND	ND	Inhibitor (Bis): unpolarized	Morphological and molecular (tau-1, synapsin 1, MAP2)	PAR3, PAR6	25,26, 63
+	SAD (presynaptic differentiation)	Diffuse (st.3)	ND	KO: immature axon formation, enhancement of dendritic formation	Morphological and molecular (tau-1, MAP2)	Tau	94
-	MARK2 (microtubule affinity-regulating kinase)	All neurites, or tip of longest neurite (st.2); tip of axon (st.3)	WT or kinase dead mutant: unpolarized	siRNA: multiple axon-like neurites	Morphological and molecular (tau-1, synapsin 1)	Tau, aPKC	89
+	CRMP2 (mediator in semaphorin 3A signalling; cargo receptor)	Diffuse (st.2); distal part of axon (st.3)	WT: multiple axons, or increased length of axon	siRNA: unpolarized, or decreased length of axon	Morphological and molecular (tau-1, synapsin 1, synaptophysin, MAP2)	Tubulin, kinesin 1, numb, GSK3β, Akt, PI3K, BDNF, NT3	39,98, 99,104, 132

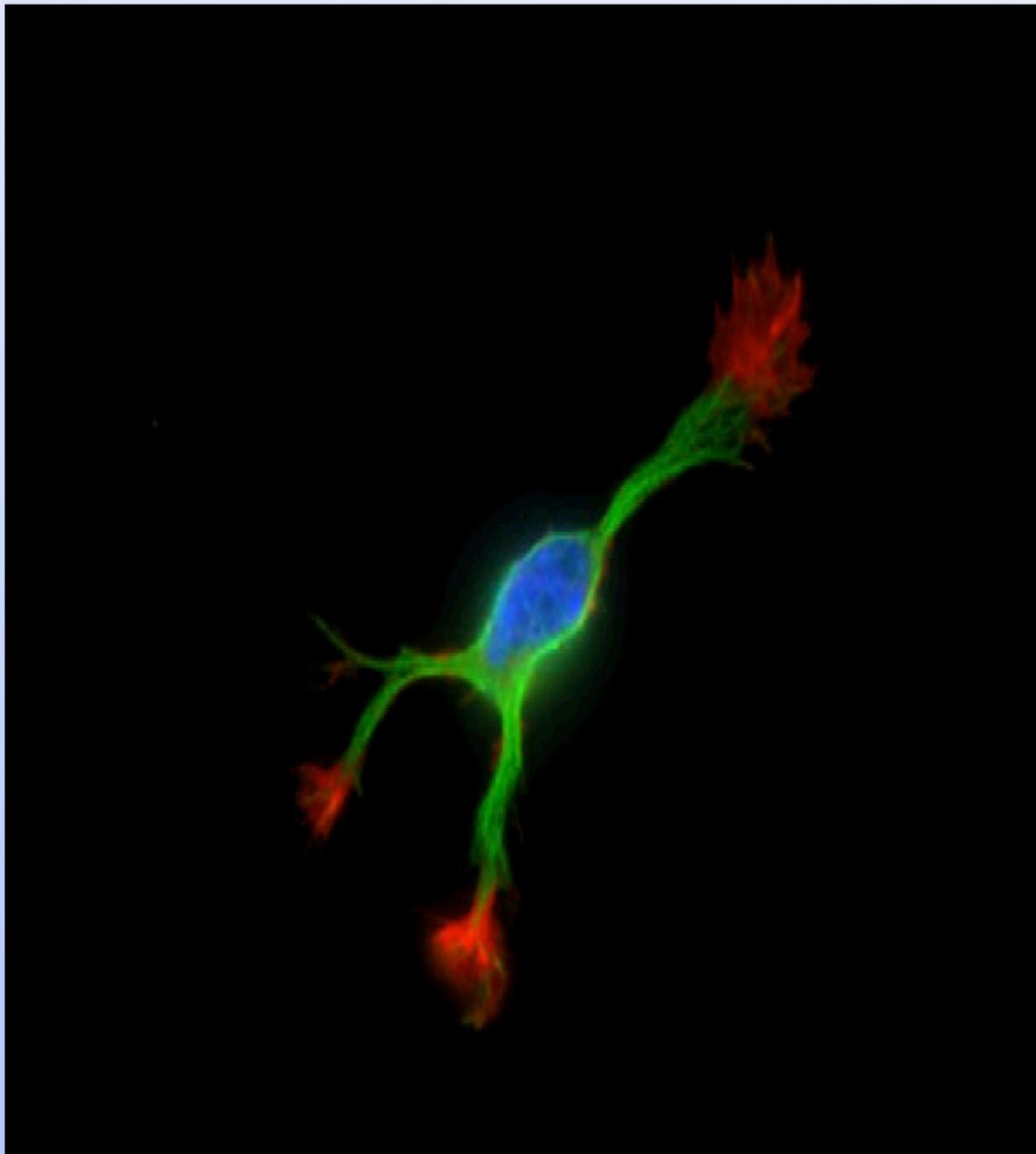
## 5. Le cytosquelette et la polarité neuronale



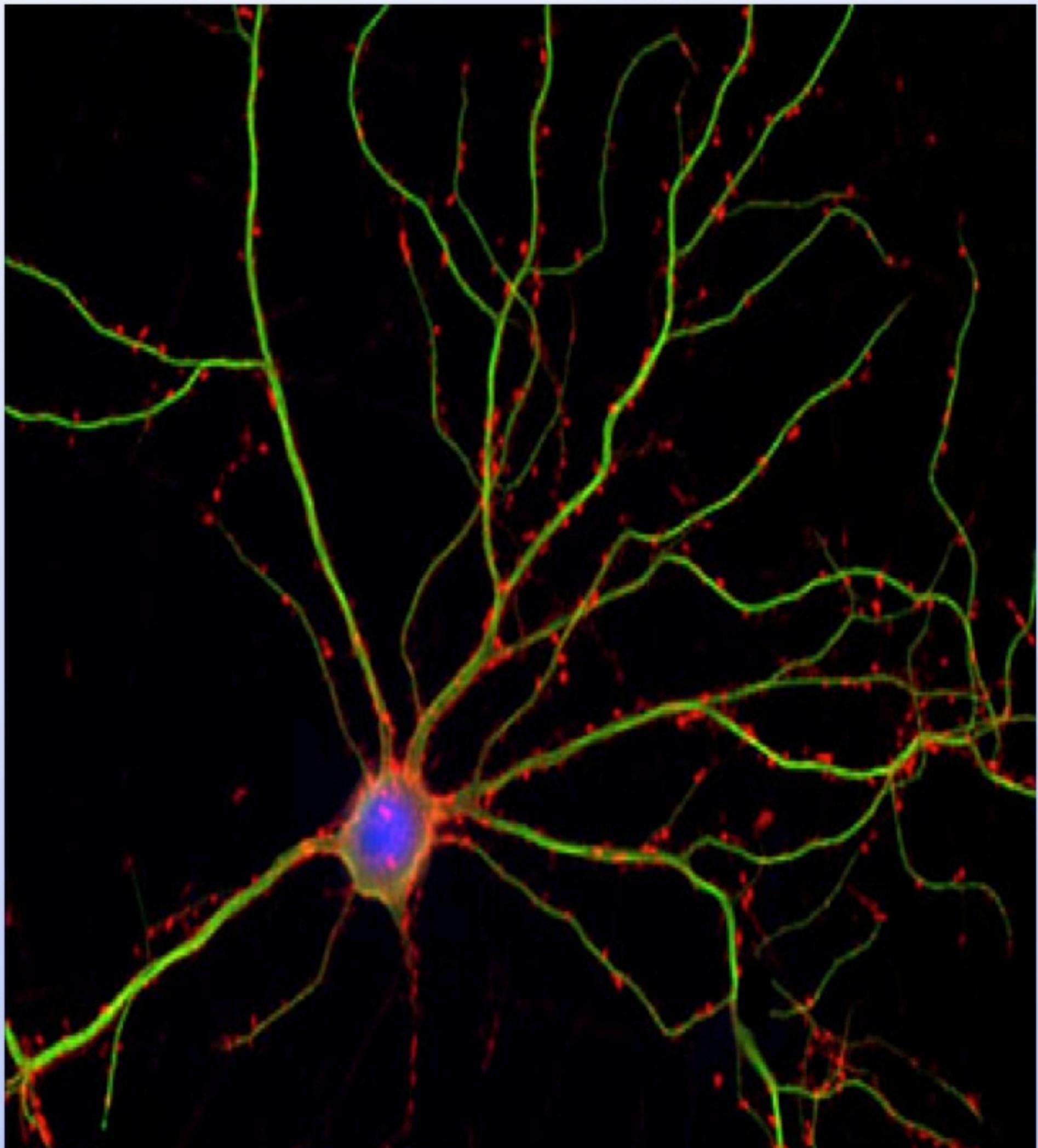
# 5. Cytosquelette et polarité

## a) Le rôle de l'actine

HPC neuron, 24h



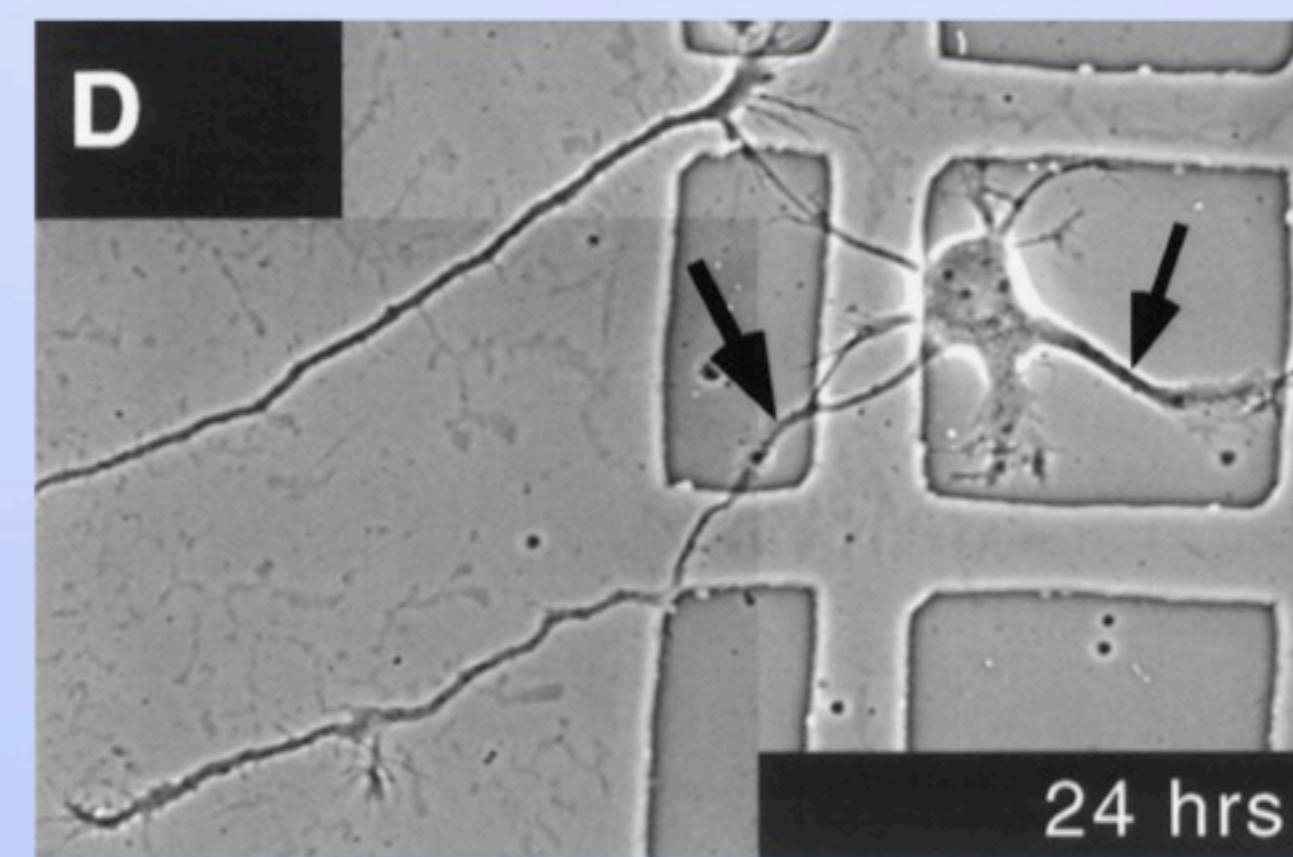
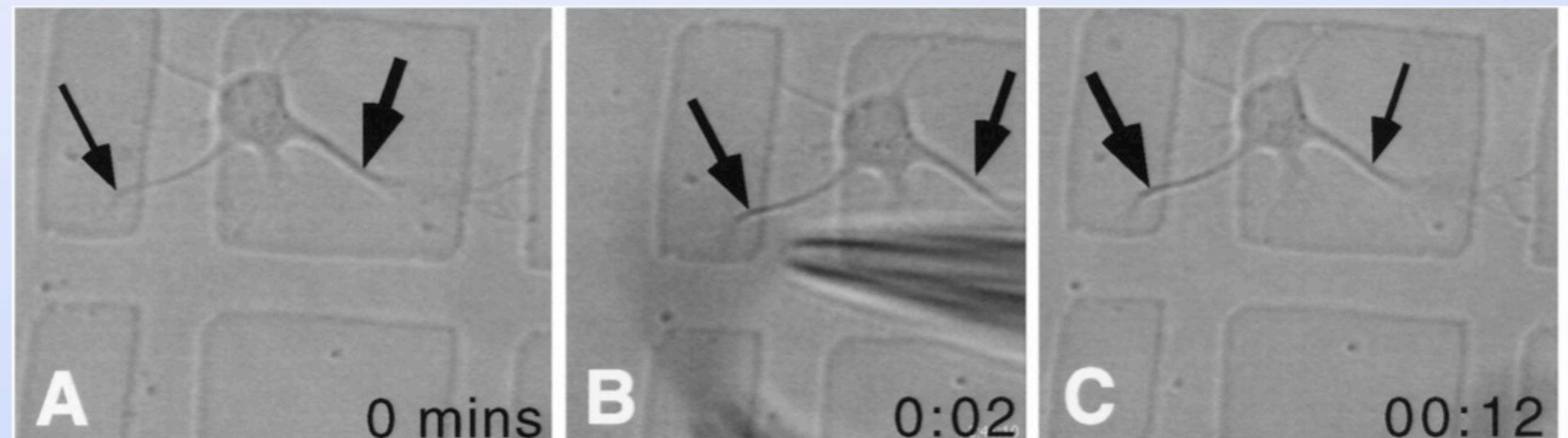
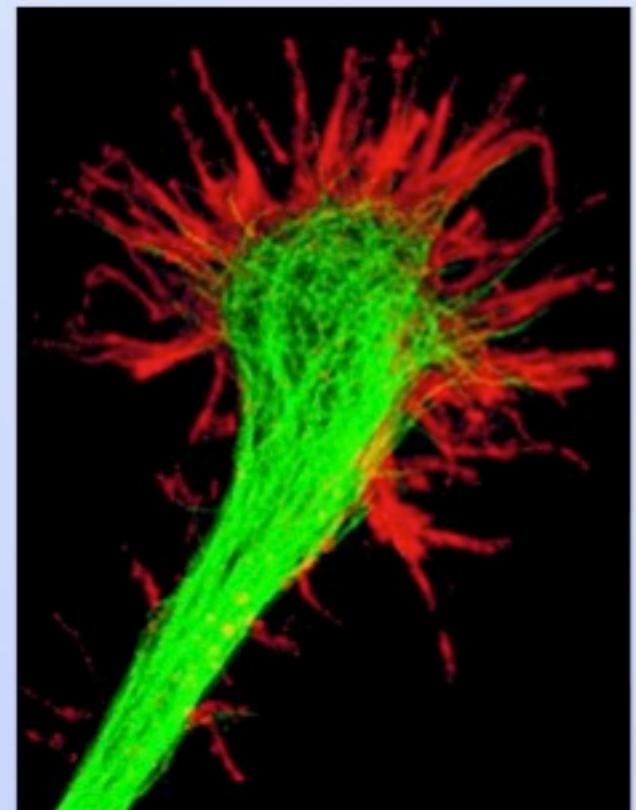
HPC neuron, 3 weeks



DNA- blue;  $\mu$ tubules- green; actin- red

# Instabilité de l'Actin dans le cone de croissance

Local perfusion of cytochalasin D onto a growth cone induces it to grow as an axon, indicating that actin destabilization is sufficient for axon formation. These data strengthen the proposed hypothesis that polarized actin-filament instability determines initial neuronal polarization

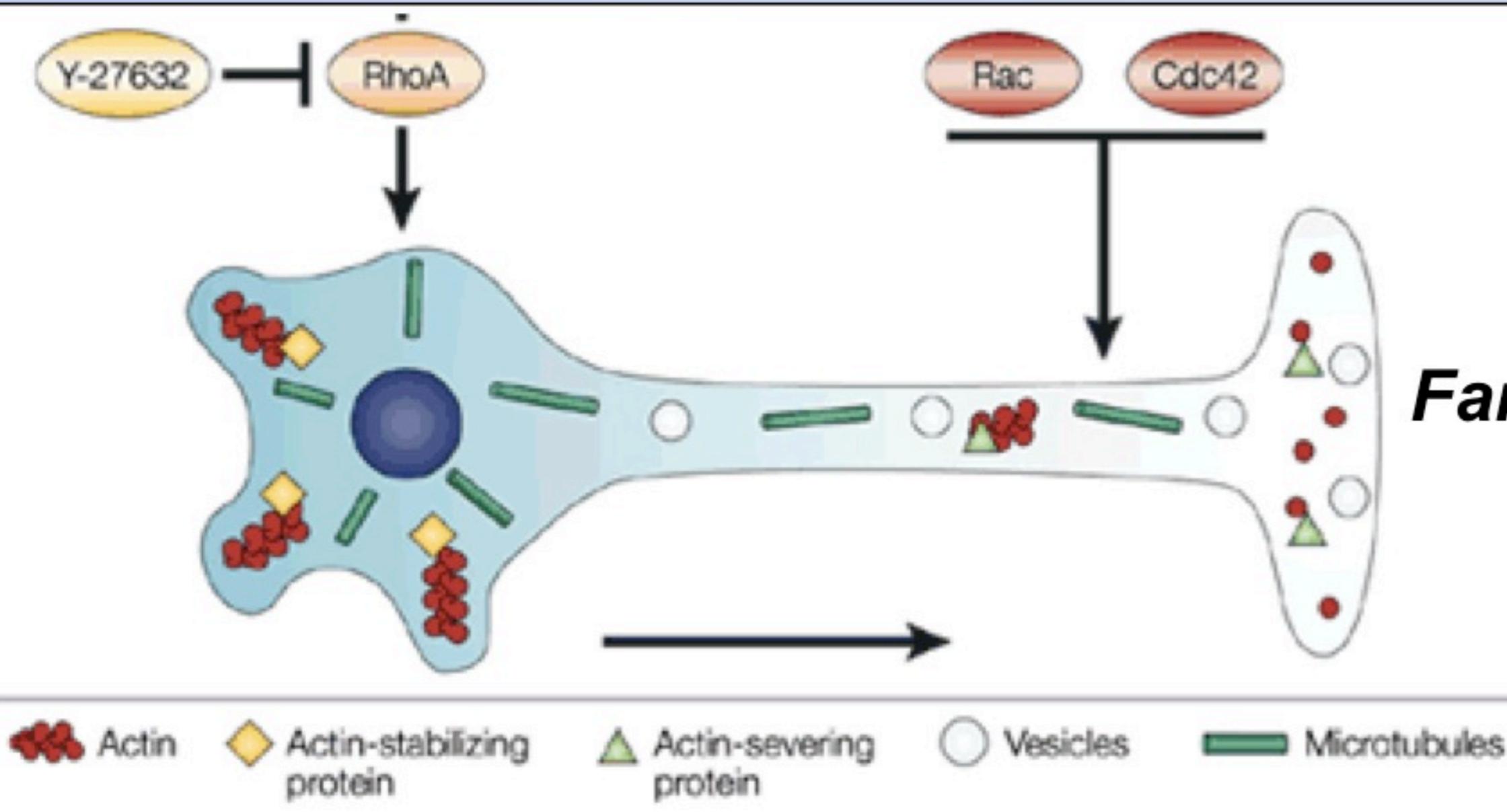
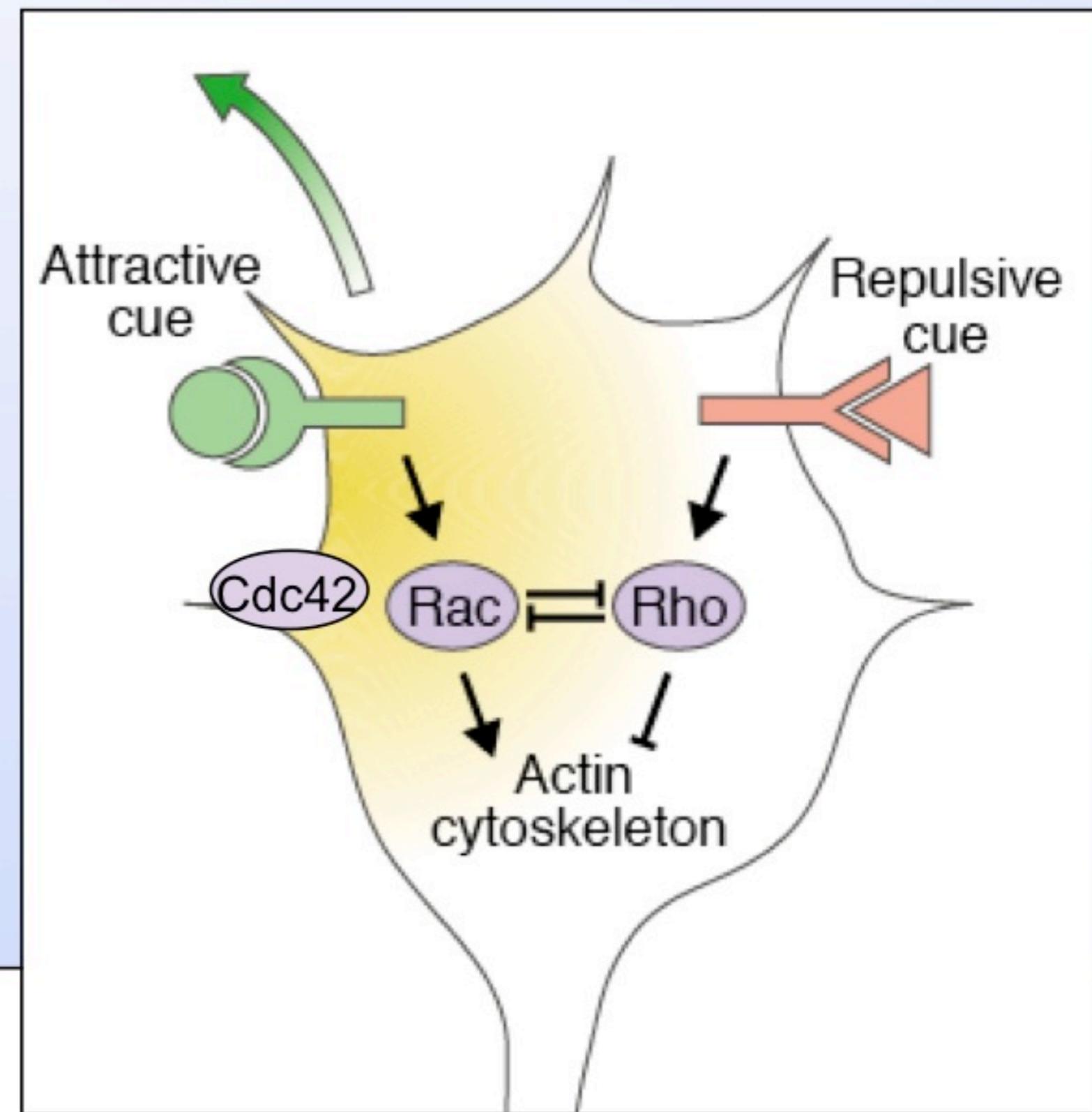
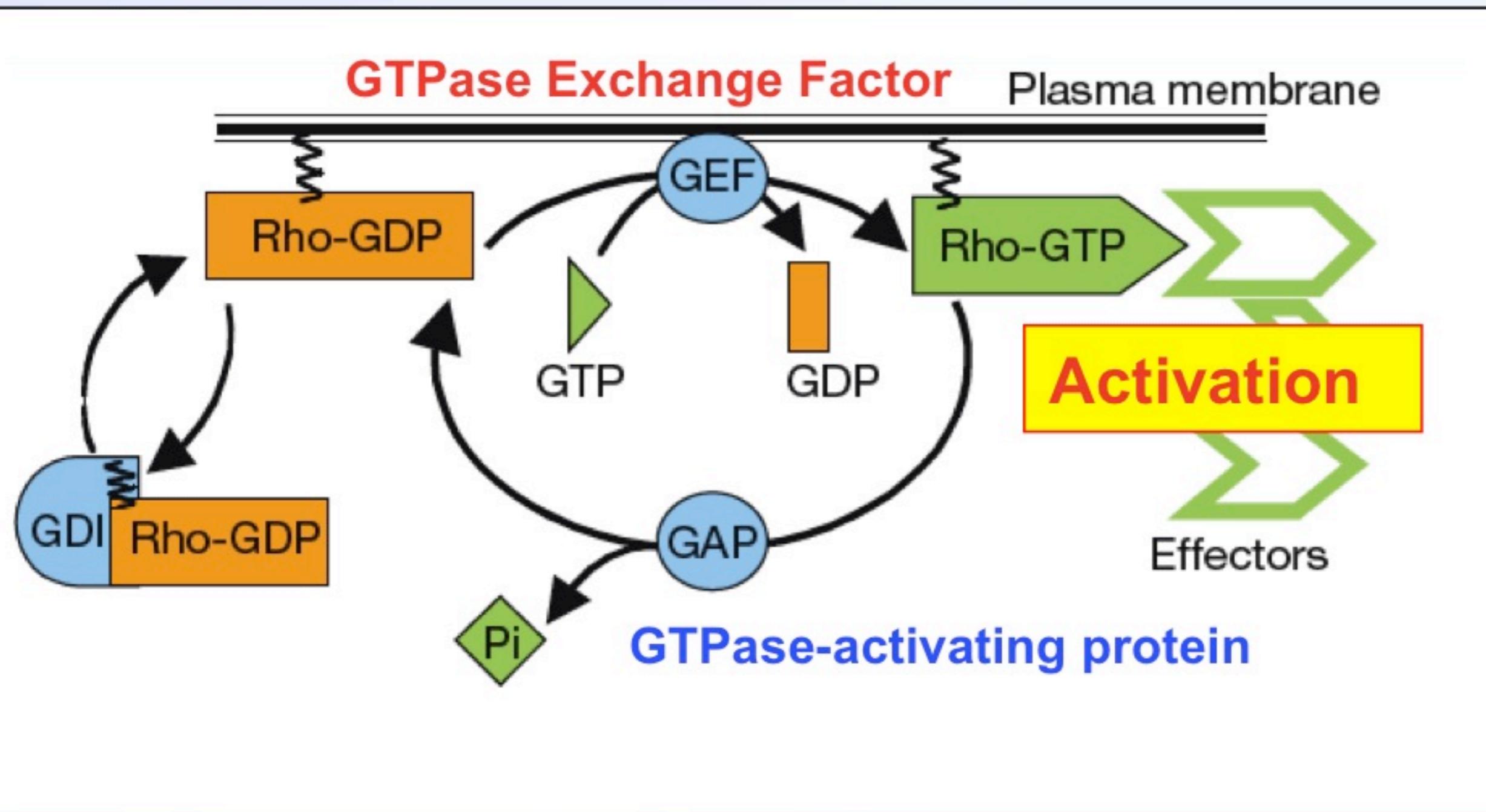


**Red: actin**

**Green: microtubule**

Bradke F, Dotti CG. Science. 1999 Mar 19;283(5409):1931-4.

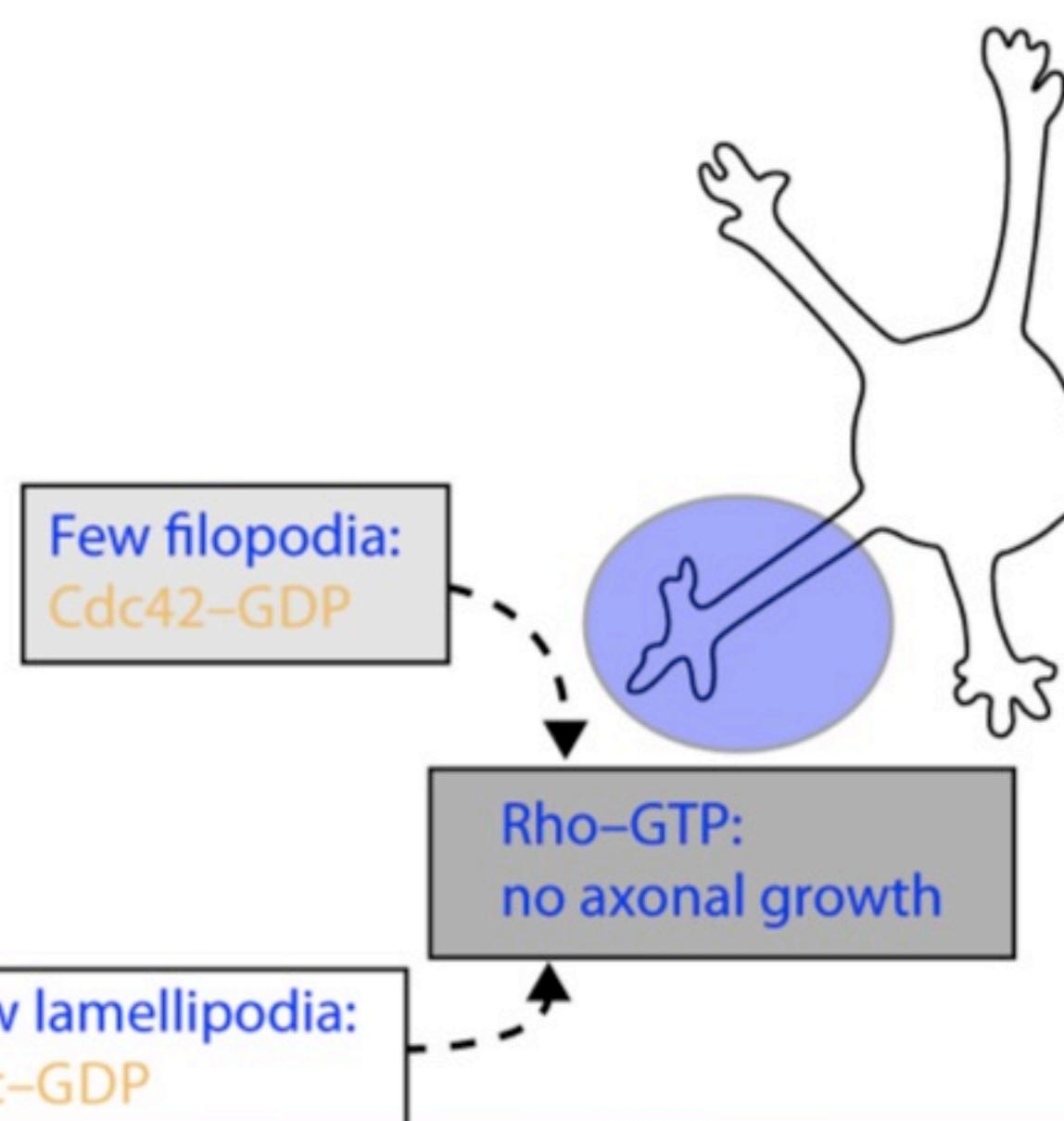
# Role du cytosquelette d'actine dans la polarité



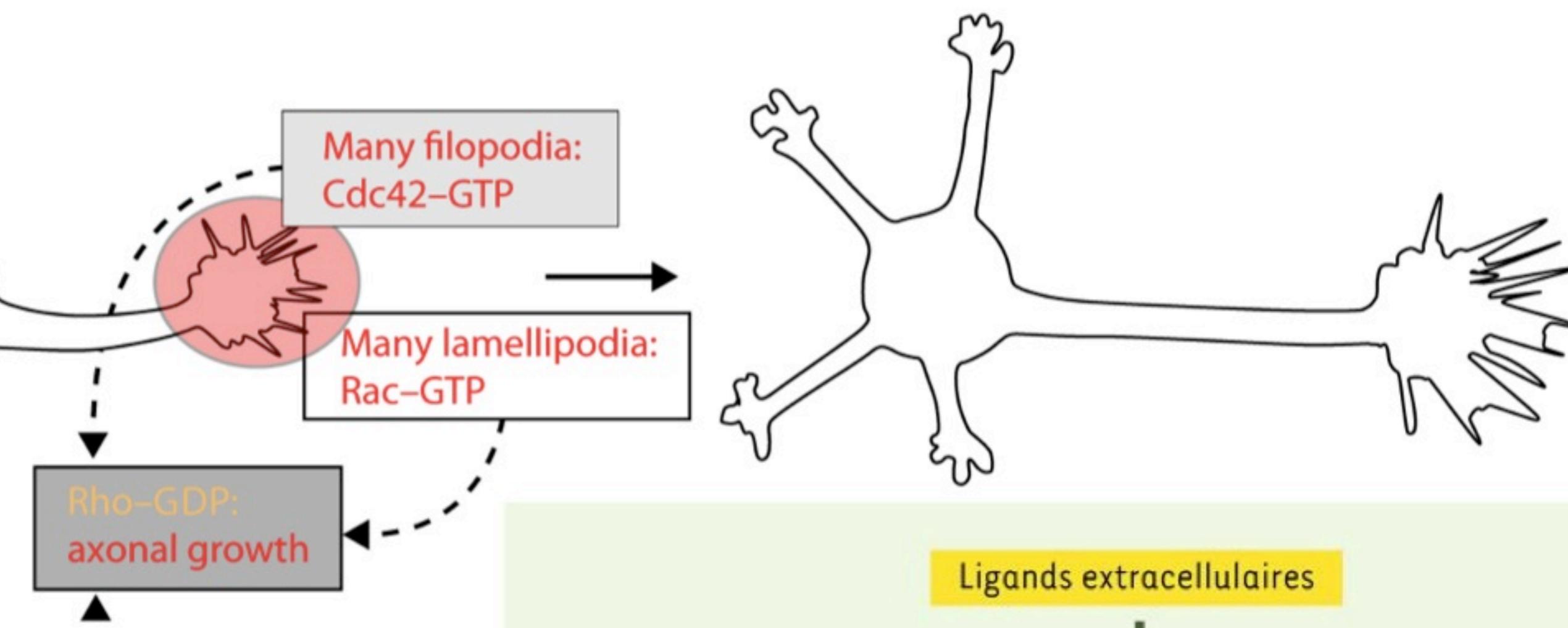
Late Stage 2

Stage 3

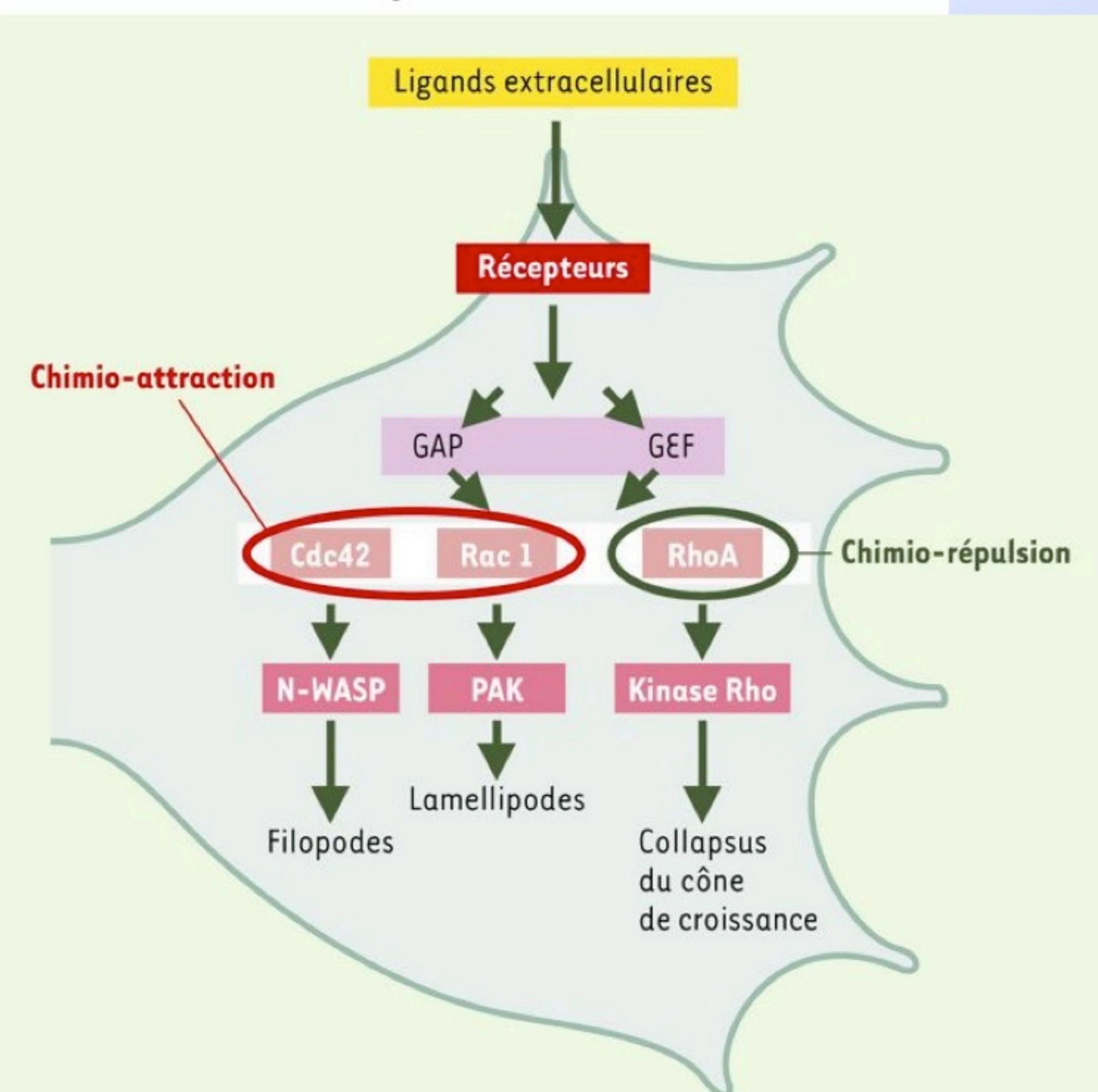
Without polarizing signal



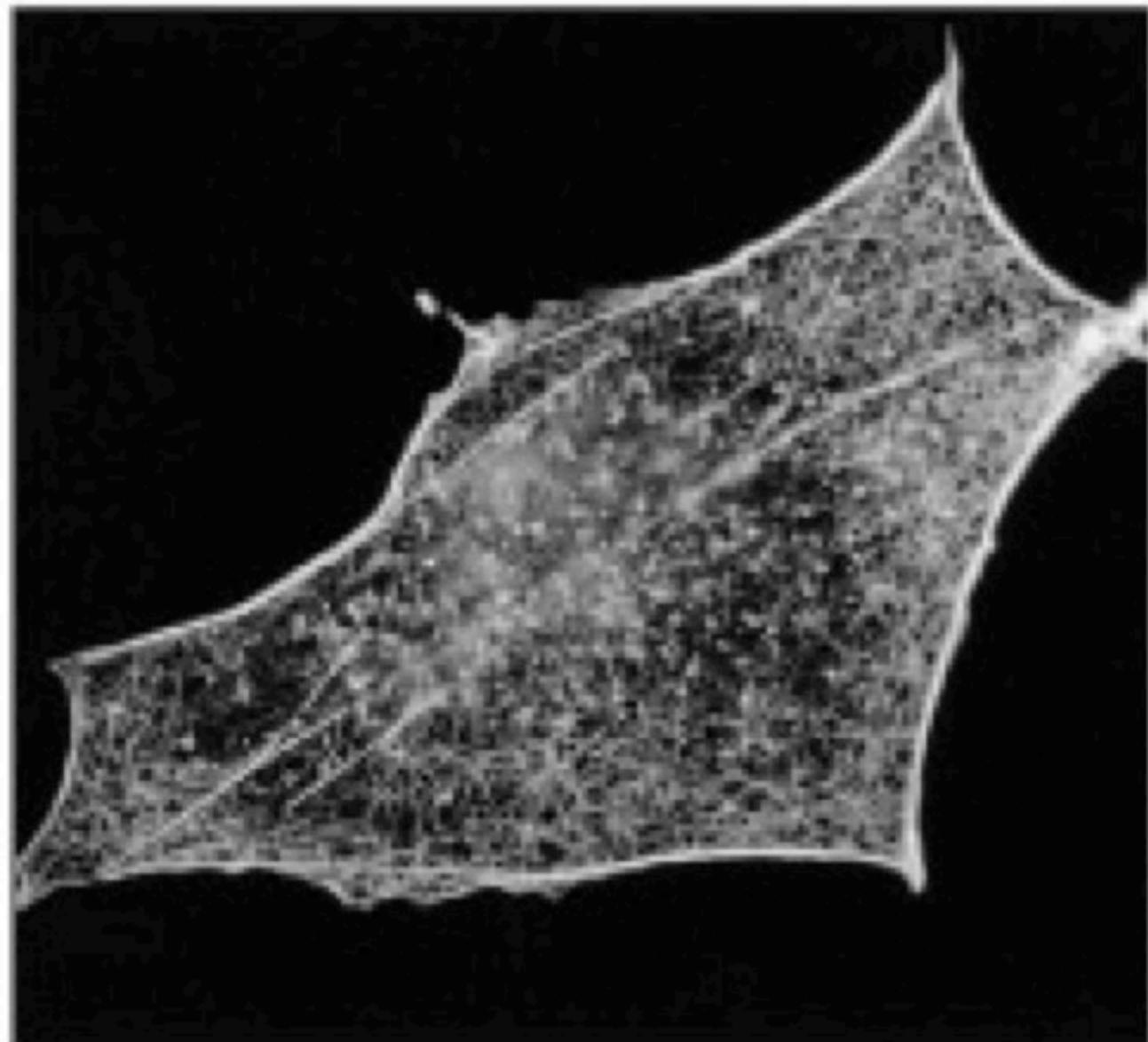
With polarizing signal



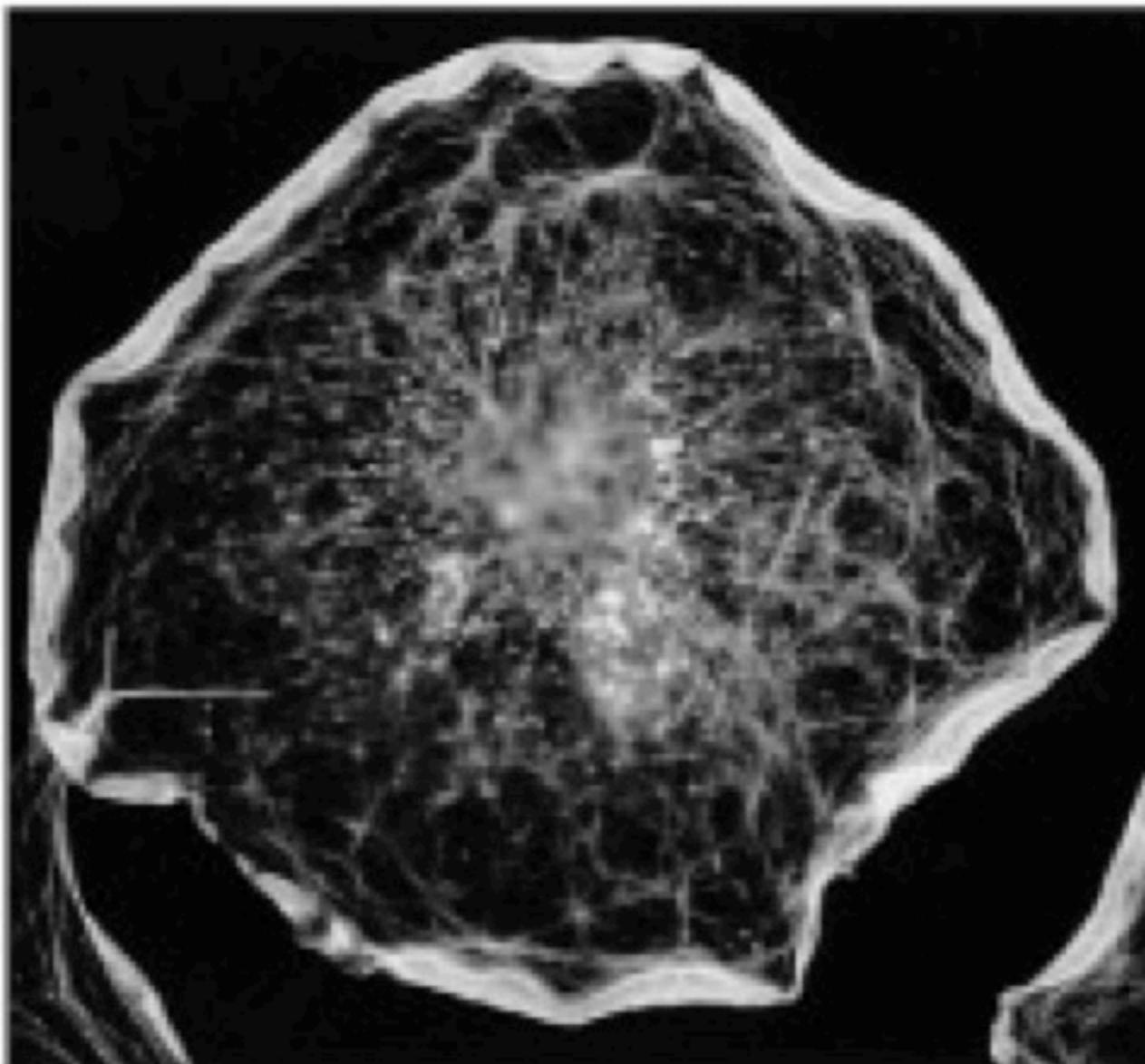
# Rho GTPases



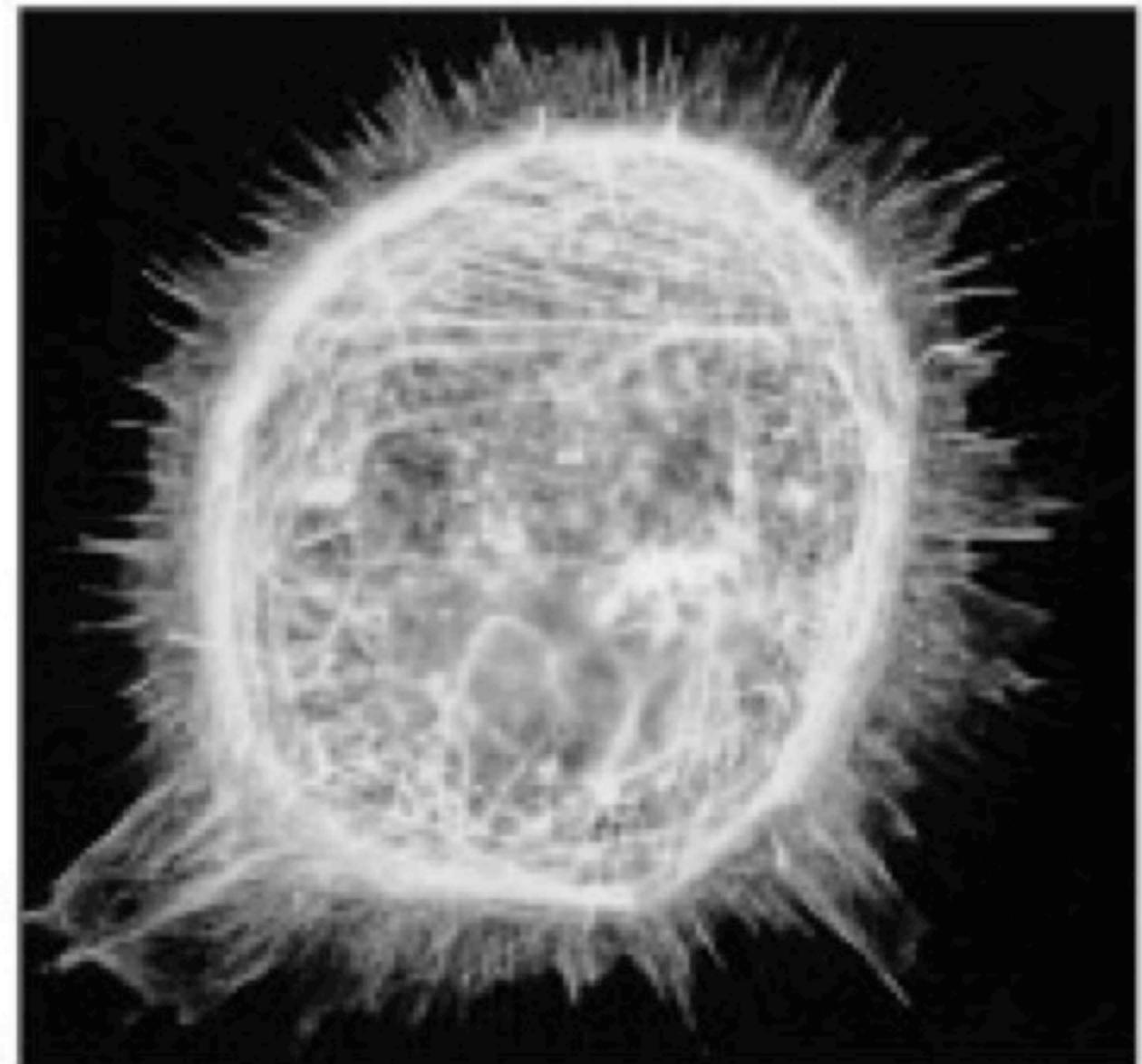
# Regulation de la morphologie cellulaire par les GTPases



(A) quiescent cells



(B) Rac activation



(C) Cdc42 activation

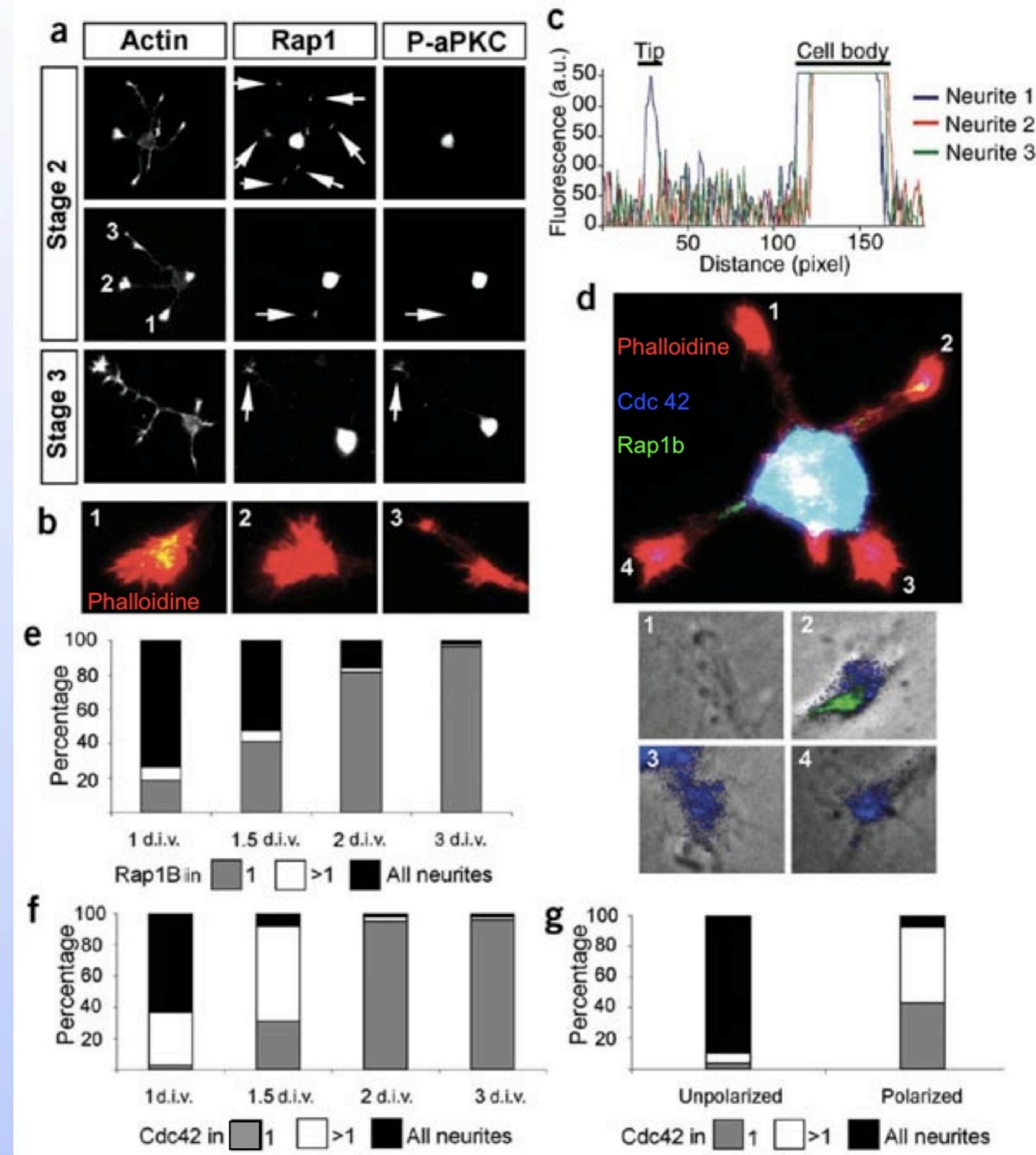
Formation de  
lamellipodes

Formation de  
filopodes

*The sequential activity  
of the GTPases  
*Rap1B* and *Cdc42*  
determines neuronal  
polarity.*

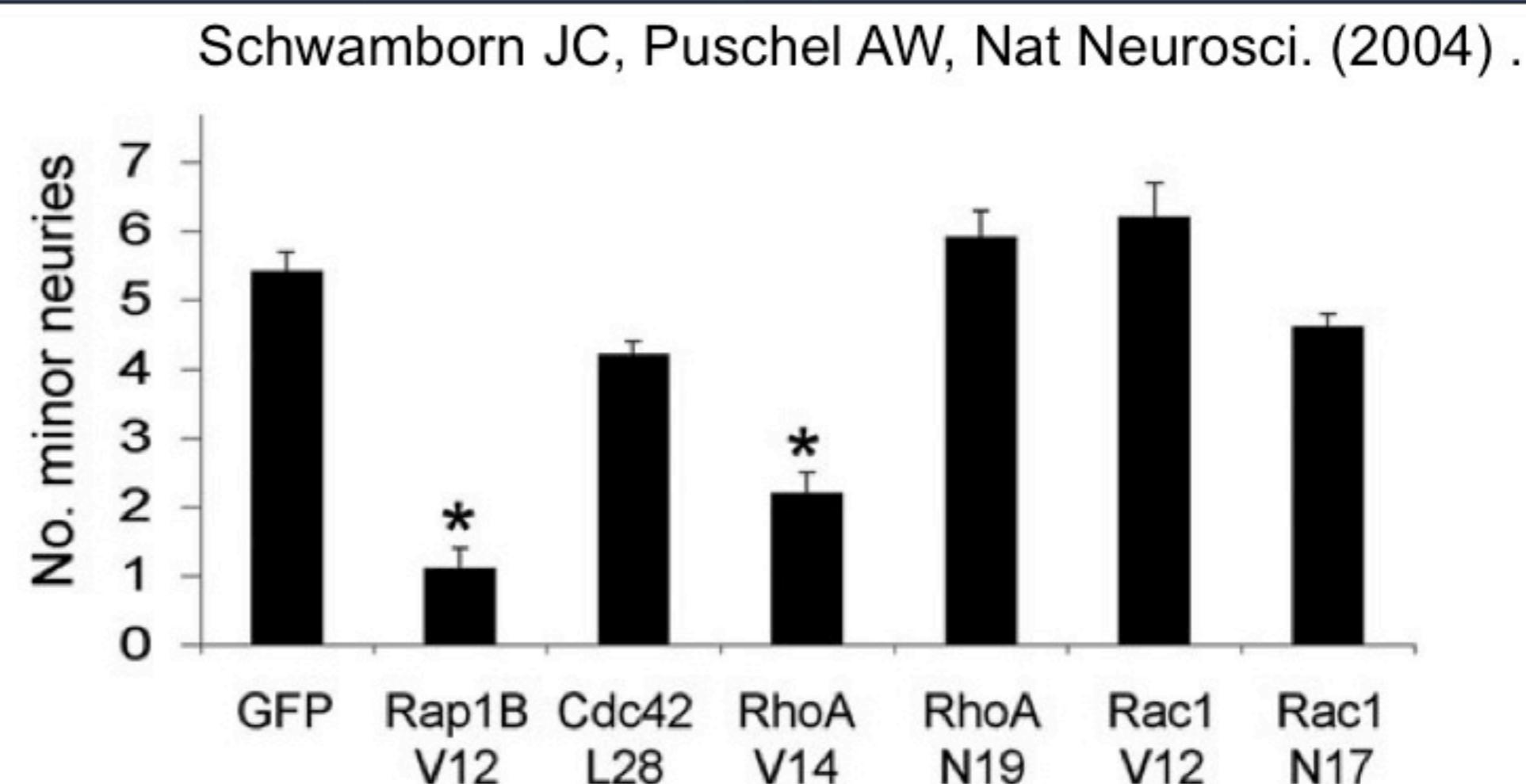
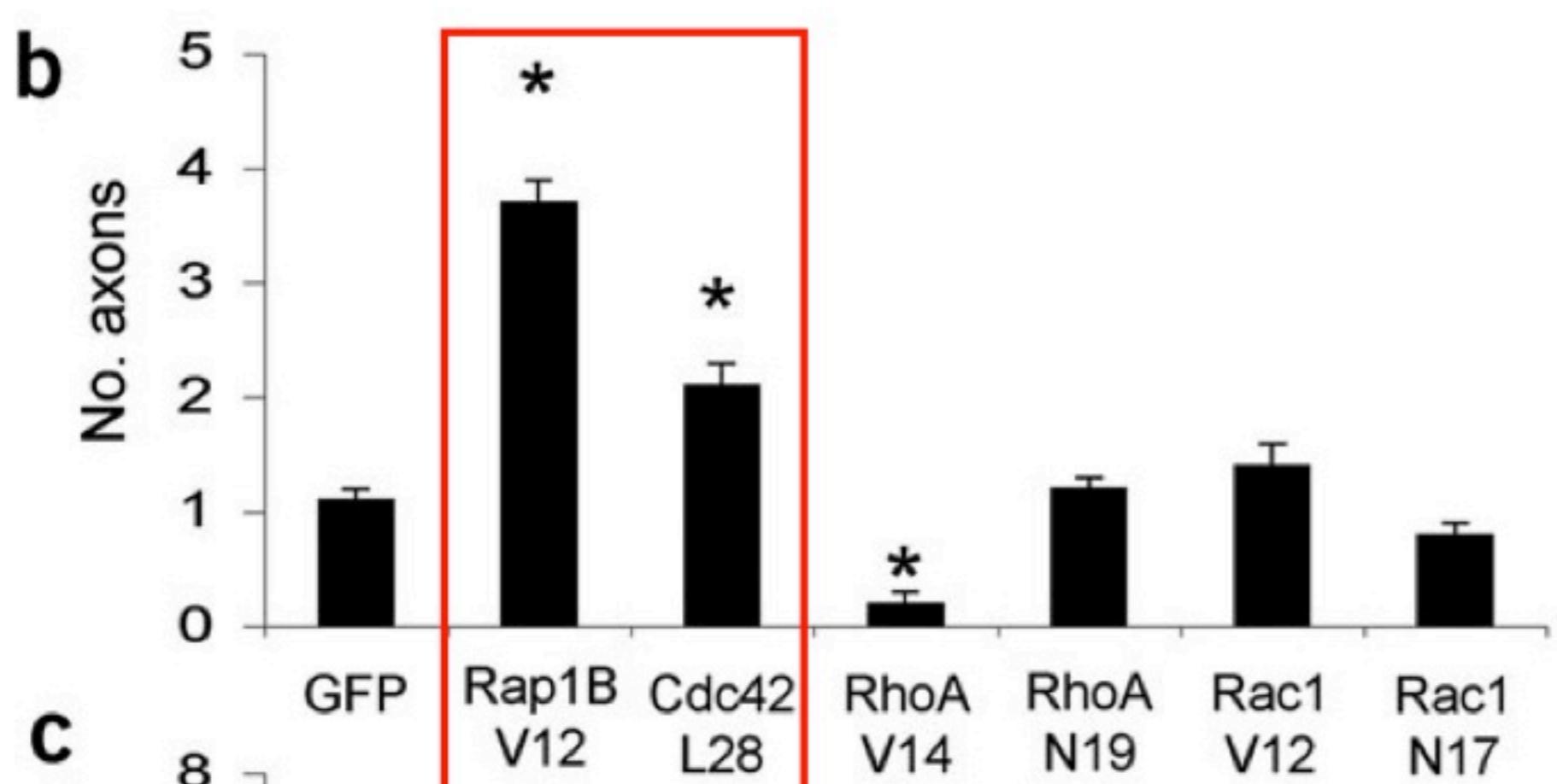
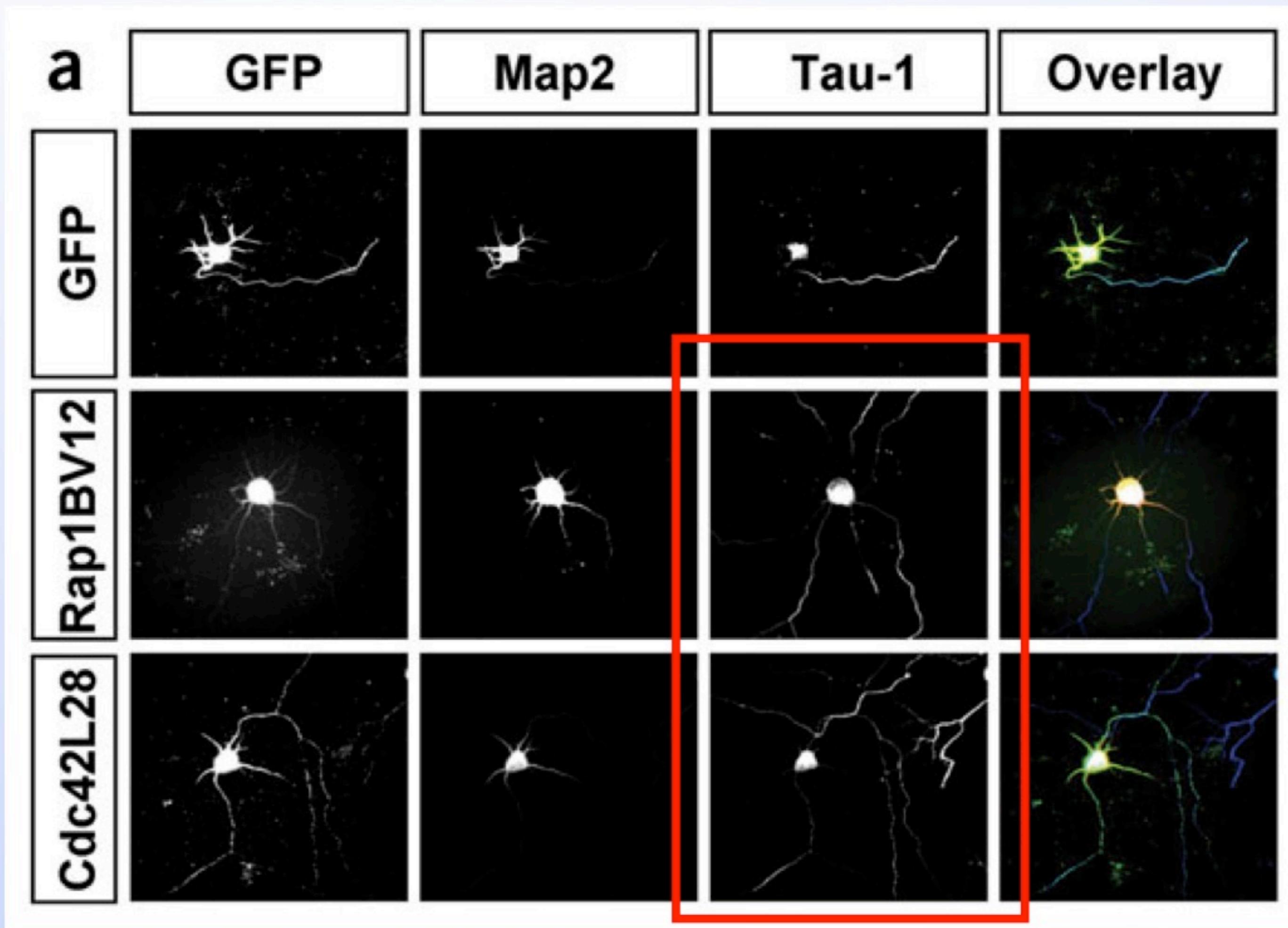
Rap1B est localisé dans tous les neurites

Puis il est restreint ensuite à l' extrémité de l' axone avant cdc42.



The sequential activity  
of the GTPases  
Rap1B and Cdc42  
determines neuronal  
polarity.

L' activation de  
rap1b et cdc42 induit  
la formation  
d' axones multiples.

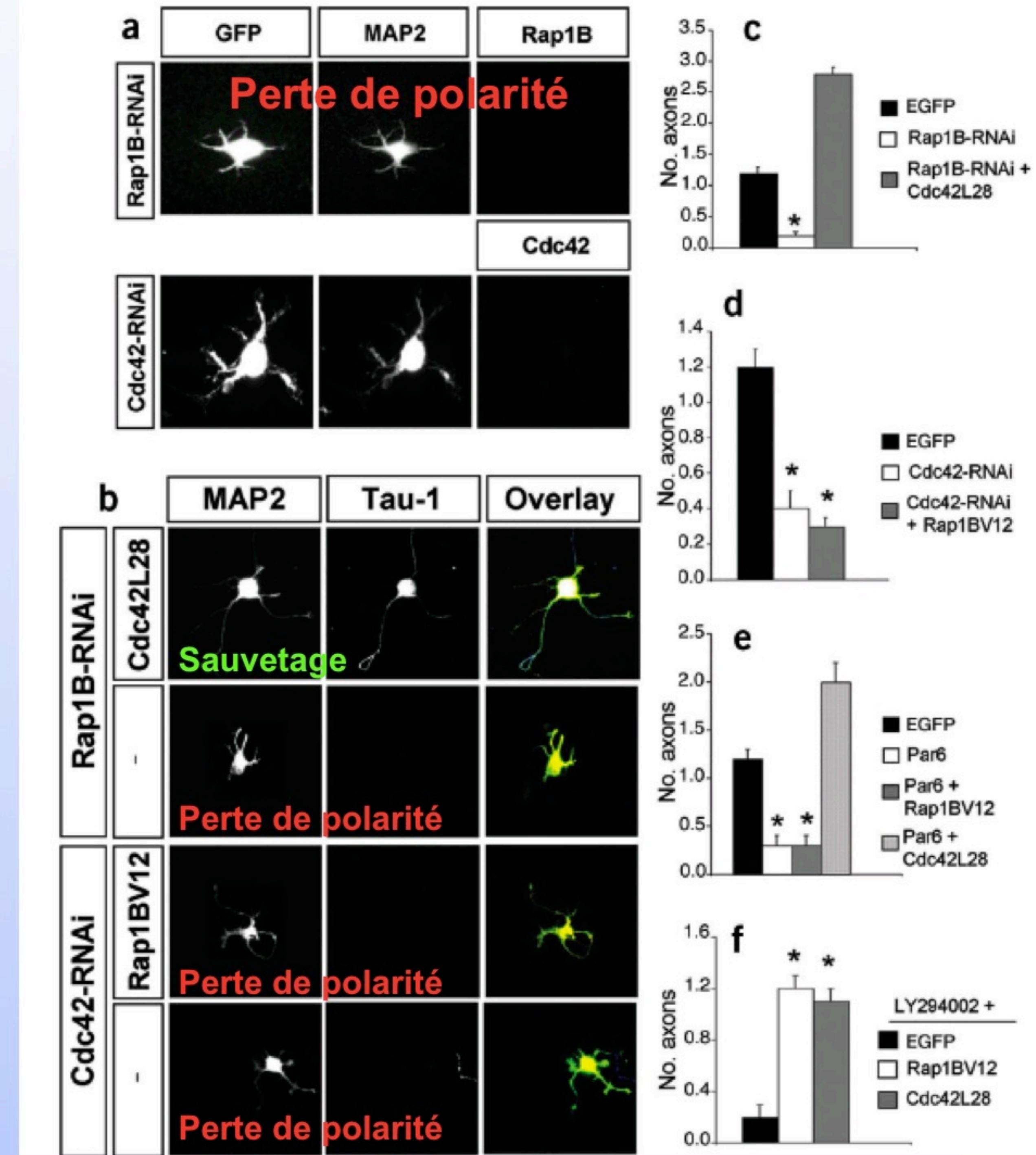


The sequential activity of the GTPases Rap1B and Cdc42 determines neuronal polarity.

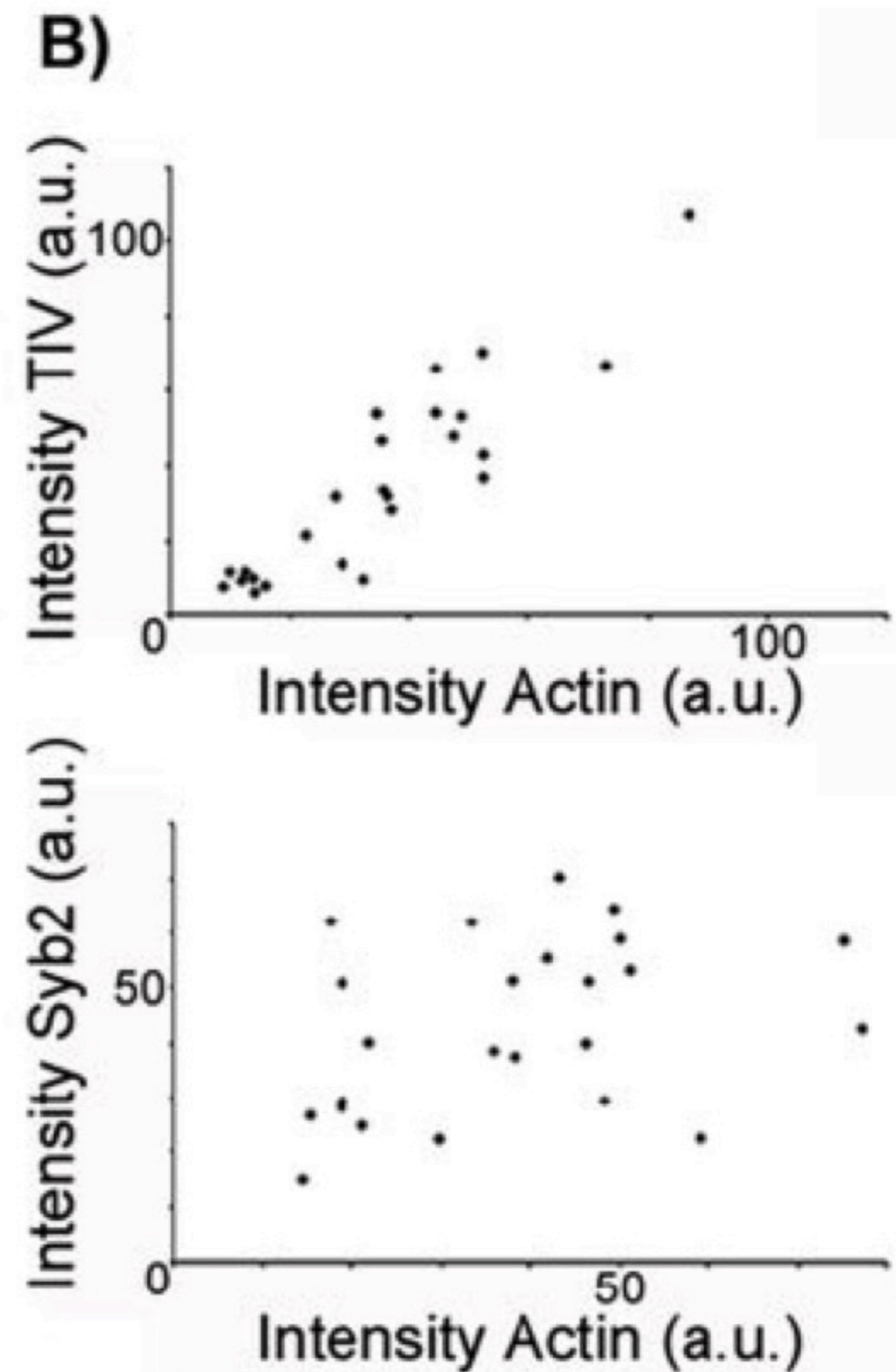
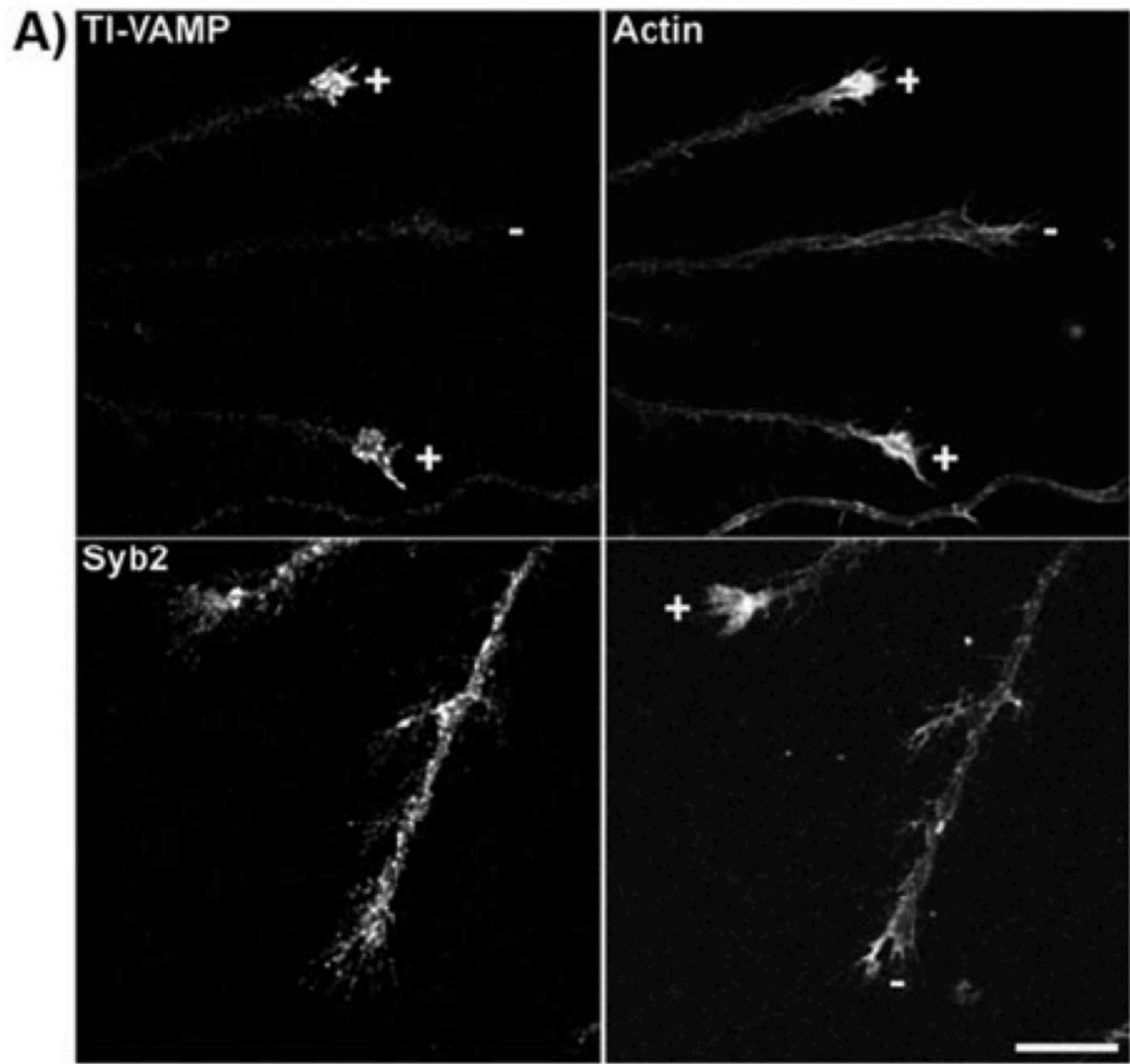
La perte de polarité du au RNAi Rap1B est sauvé par la transfection de cdc42.

La réciproque est fausse.

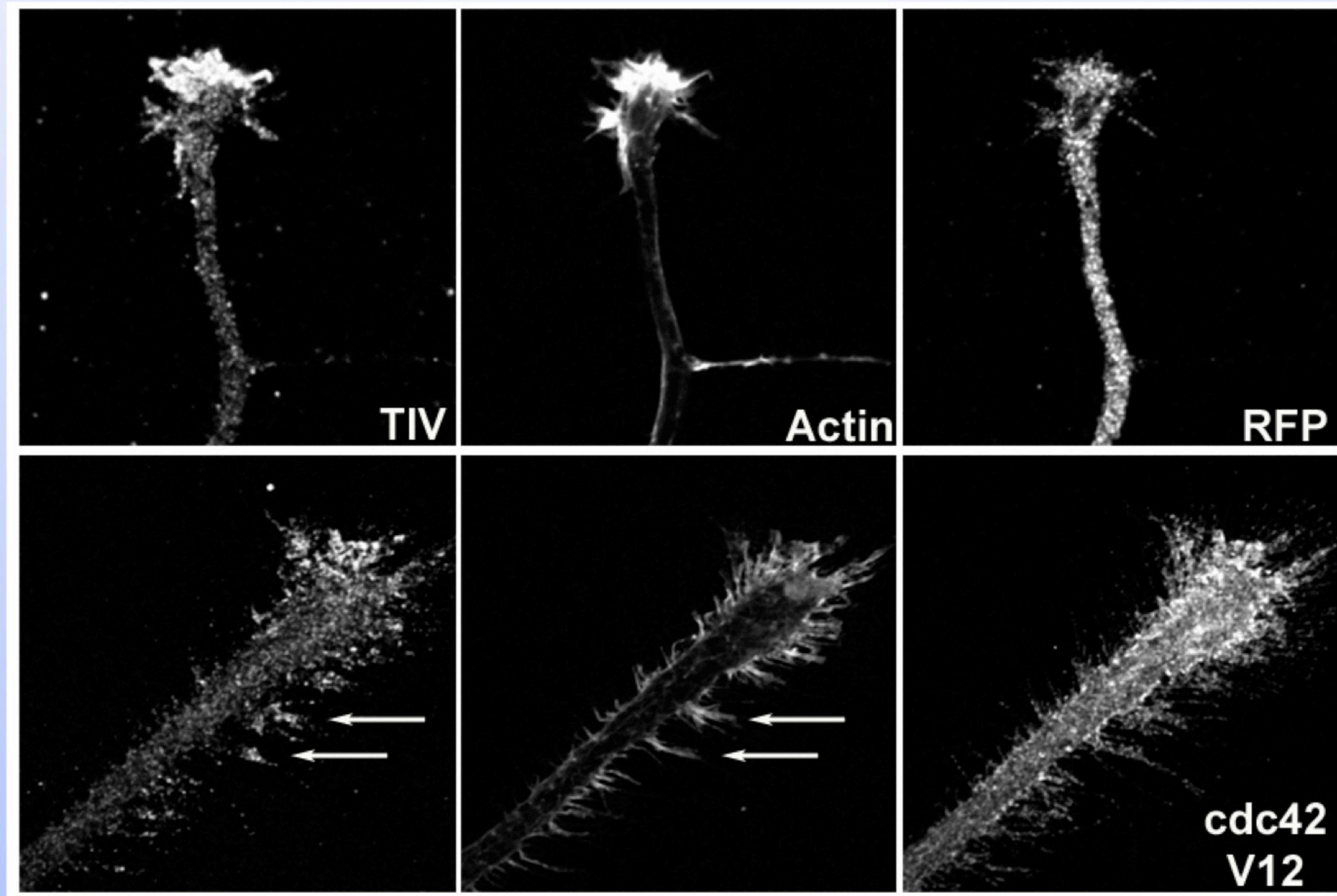
Rap1b agit donc en amont de cdc42.



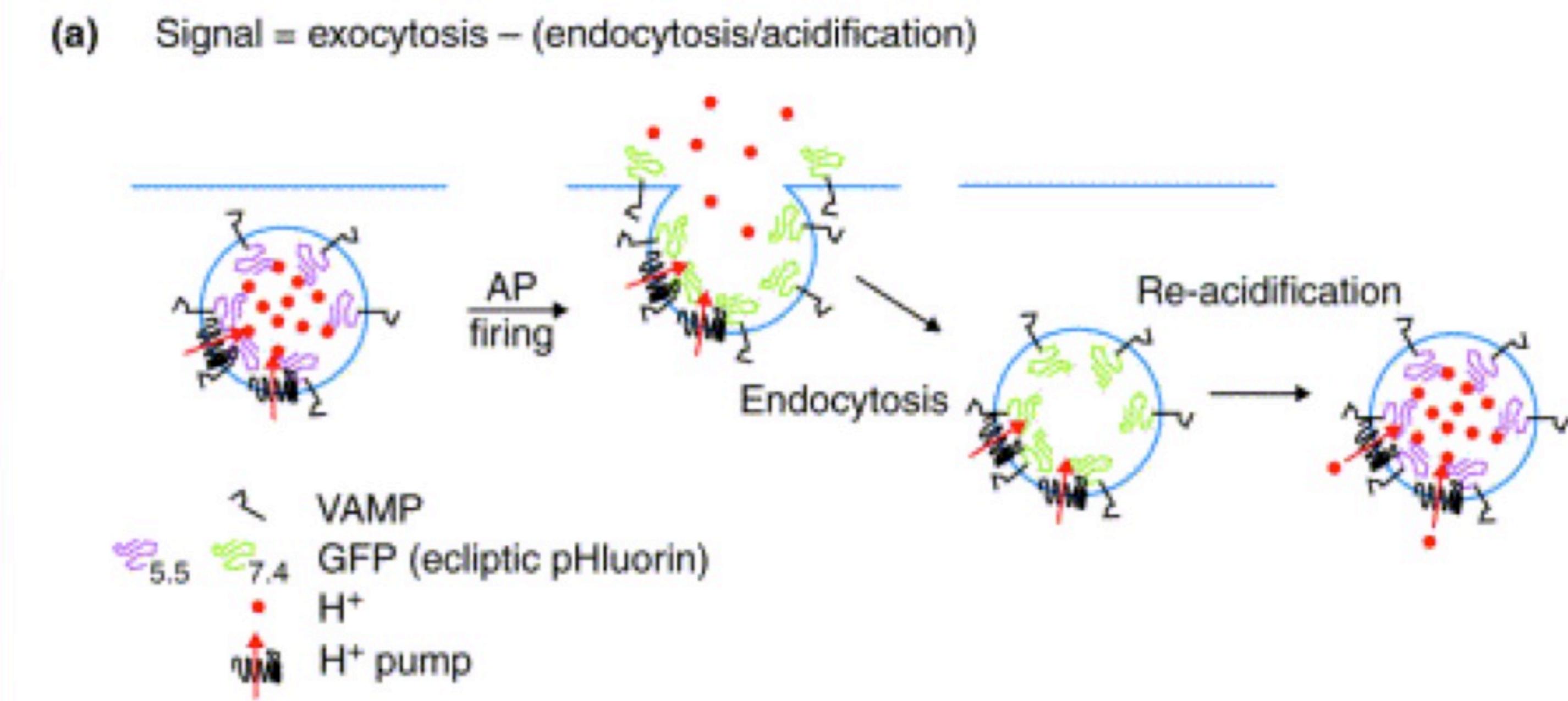
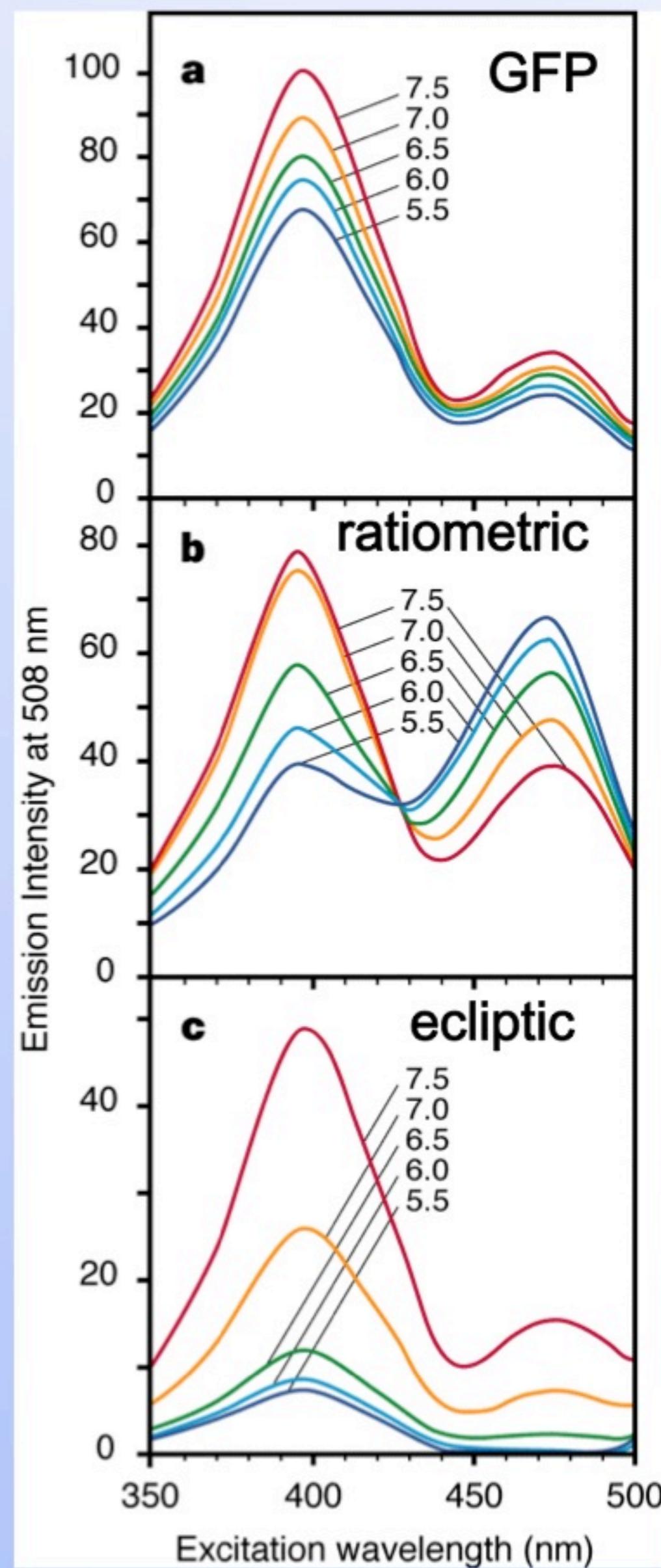
# TI-VAMP est associé au cône de croissance riches en actine



# Expression of *cdc42 V12* induces loss of polarisation of actin and TI-VAMP in growth cones

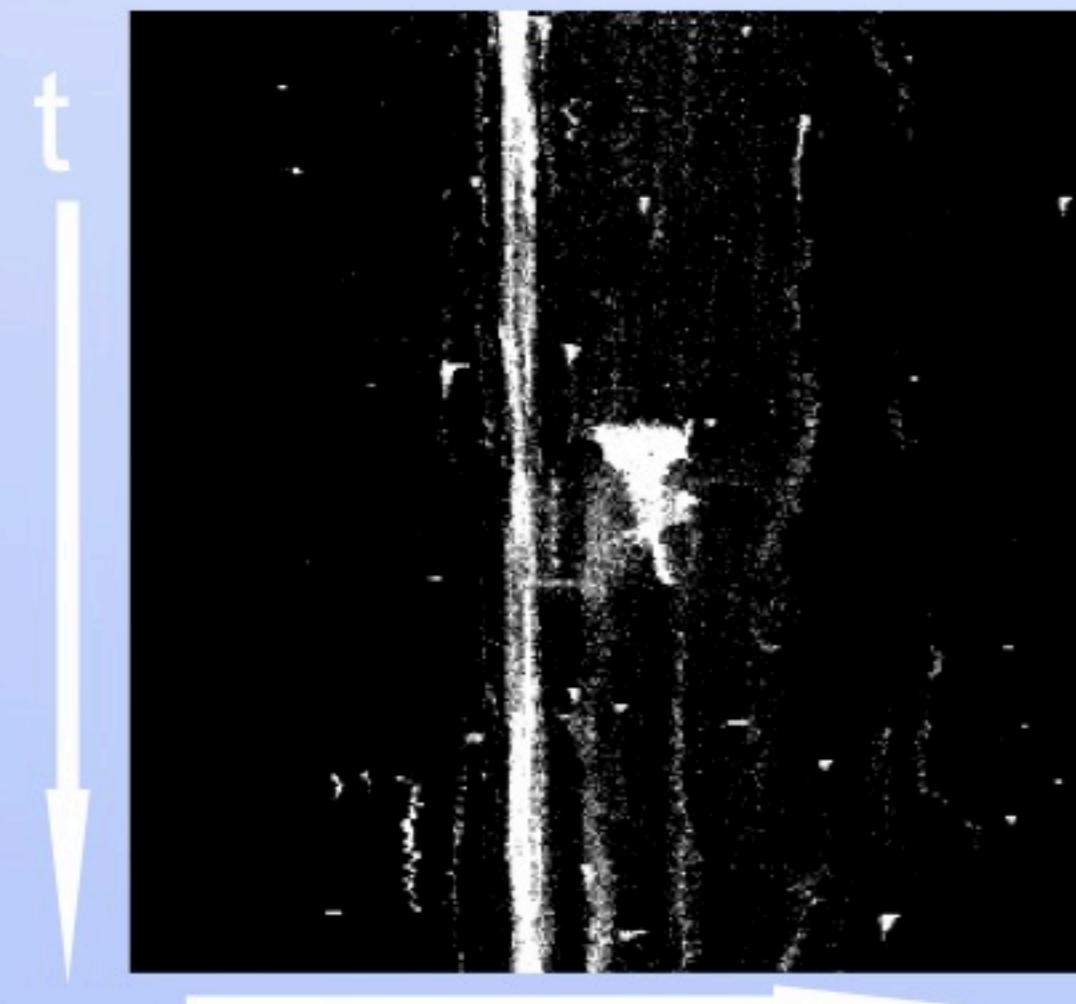
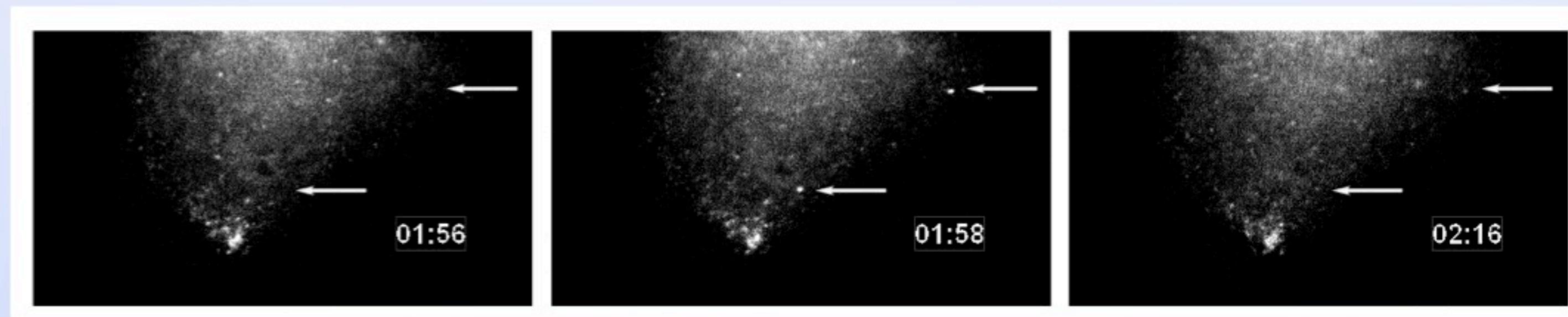


# pHLuorins - pH-sensitive GFP variants: great tools for exocytosis



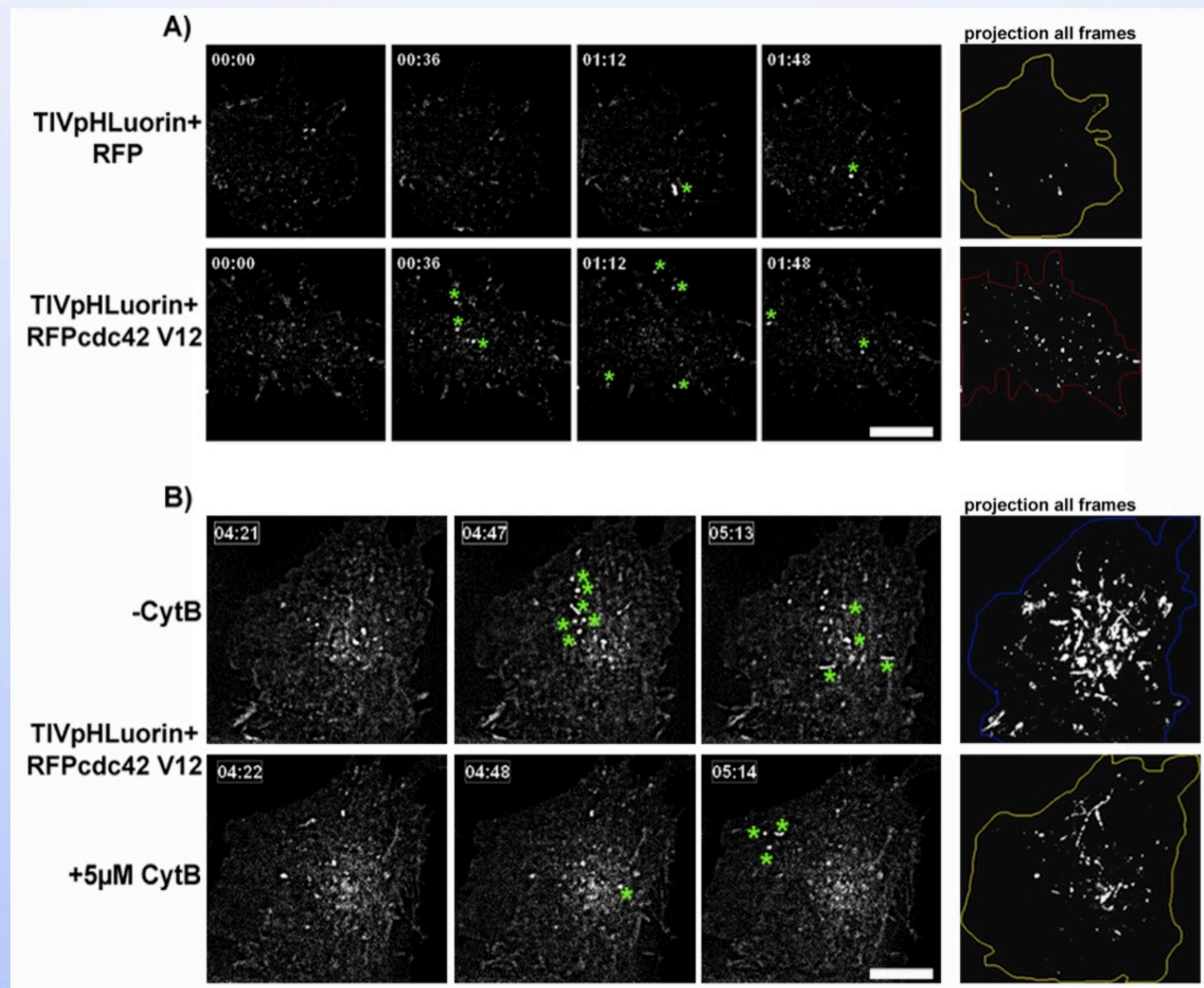
T. Ryan, Current Opinion in Neurobiology 2001

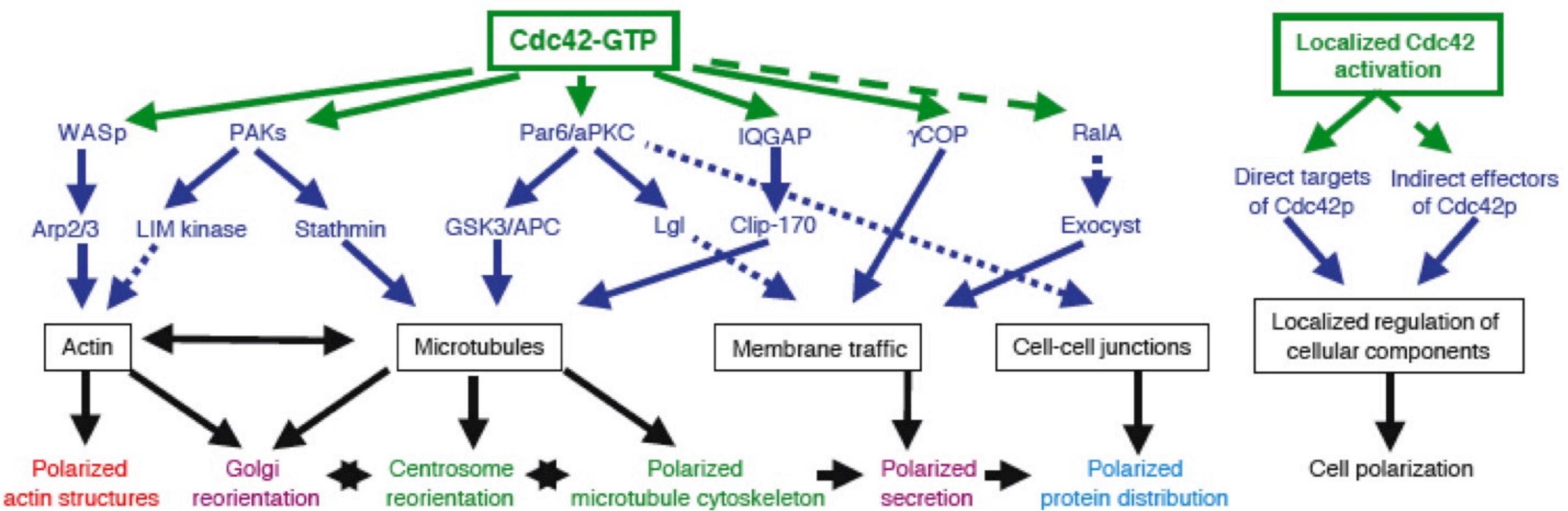
# Exocytosis of TI-VAMP-pHluorin



Region pixel max intensity

# *Actin-dependent Cdc42-mediated activation of TI-VAMP exocytosis*

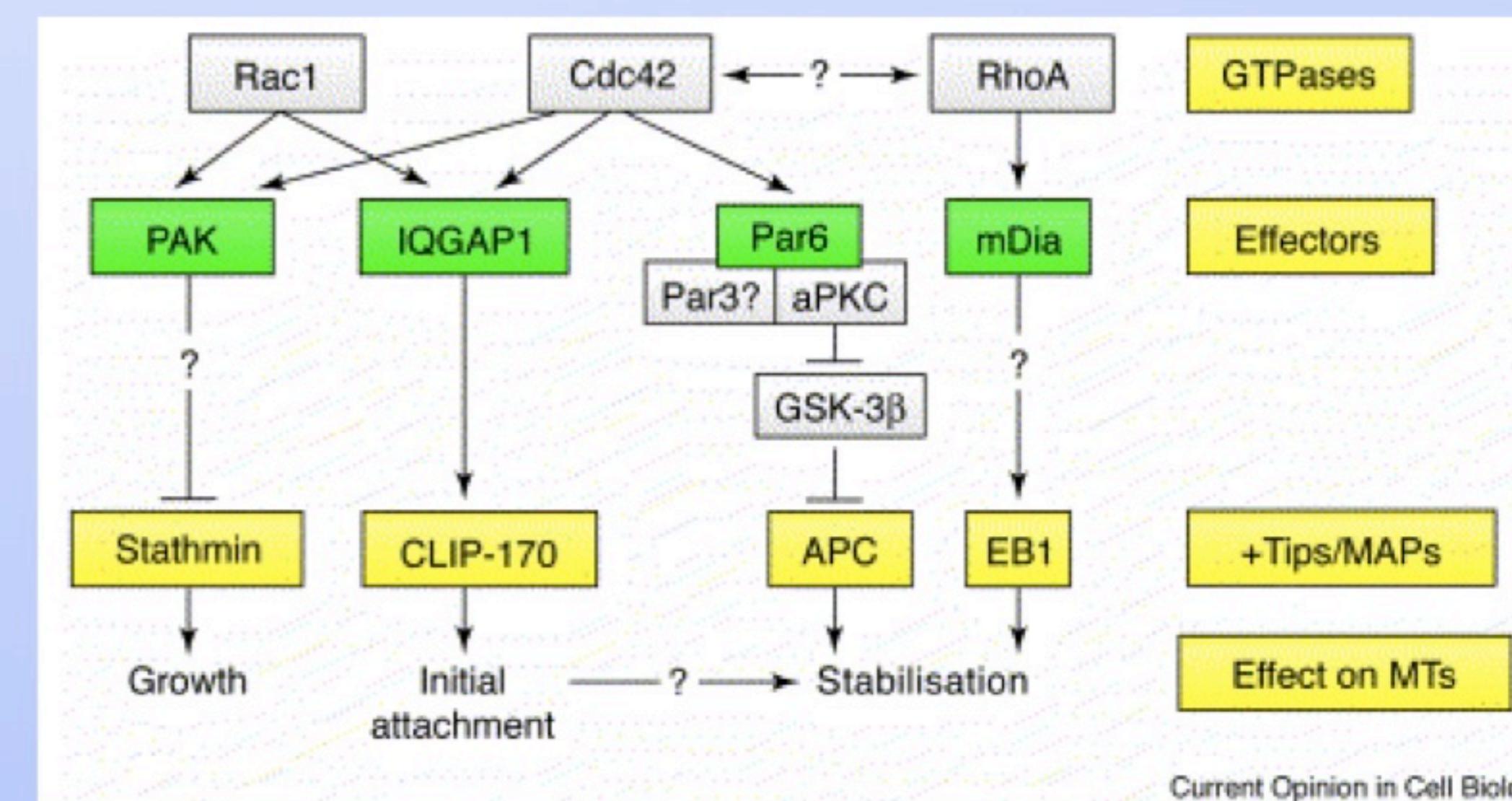
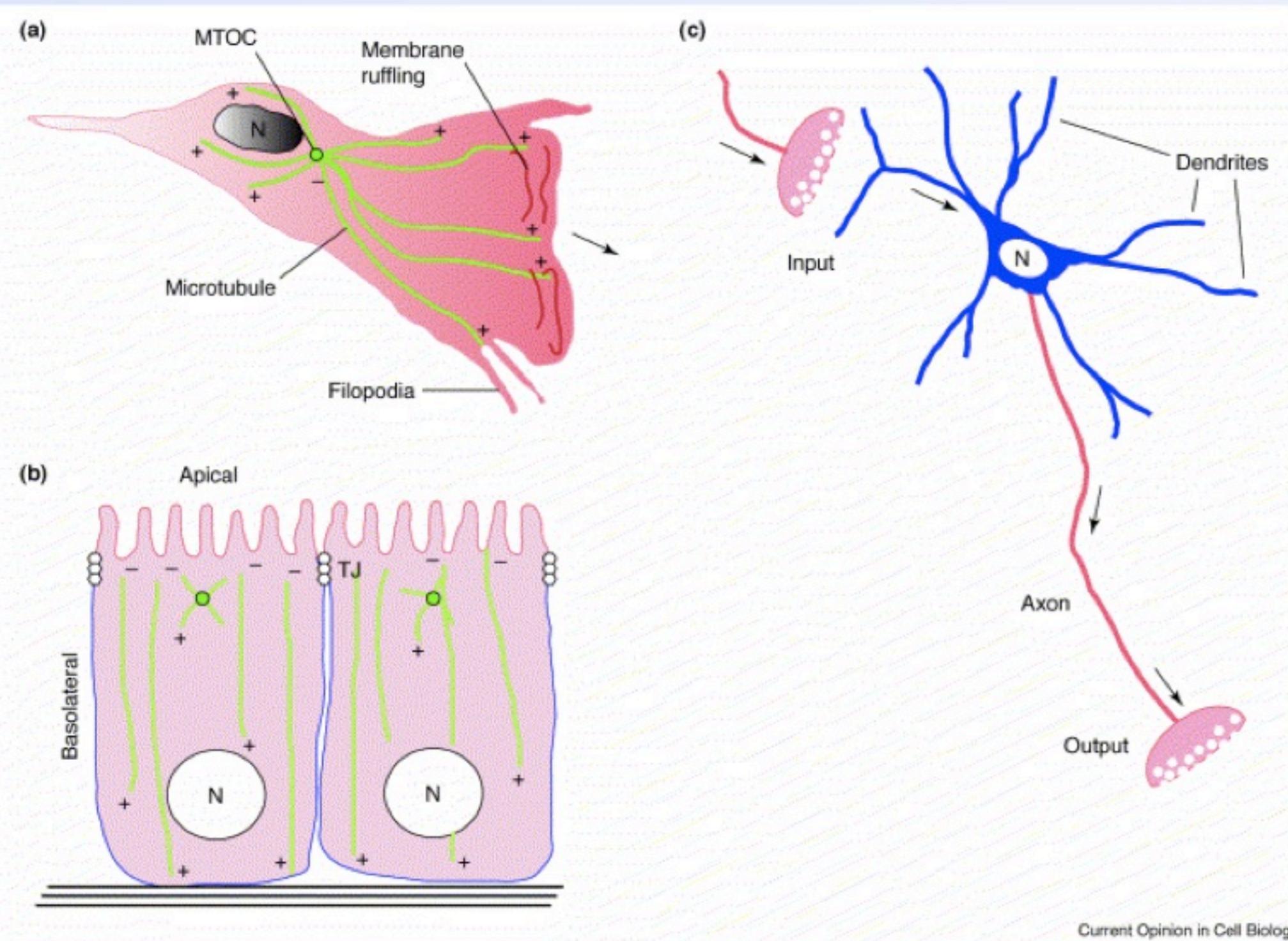


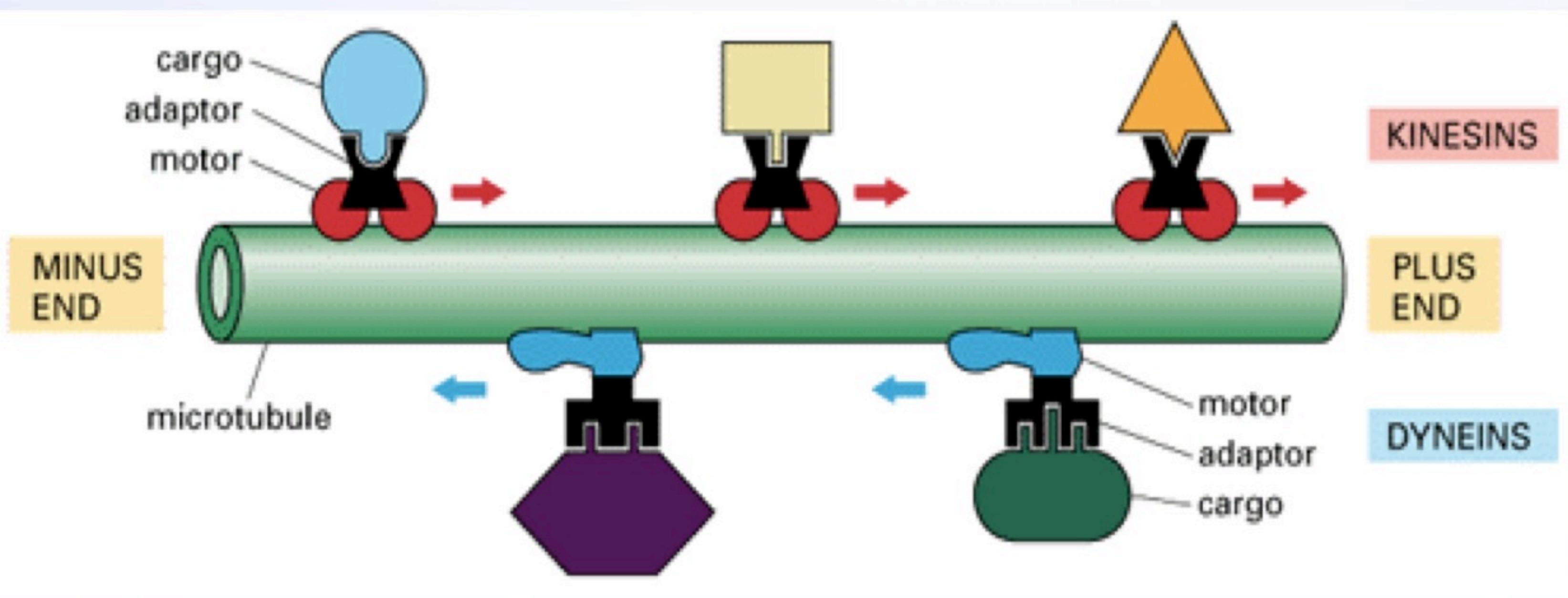


**Fig. 4.** Multiple signalling pathways controlled by Cdc42. Cell polarization requires the spatial and temporal regulation of several cell components. Orientation of the actin and microtubule cytoskeletons, regulation of cell contacts and organization of membrane traffic occur in concert. Multiple signalling pathways downstream of Cdc42 regulate these different cellular components (black box). These signals are transduced by different Cdc42 direct (solid line) or indirect (dotted line) effectors (blue) and involve several intermediates (blue). Cell polarization results from the localized activation of Cdc42, which leads to a localized regulation of cellular components and therefore to their asymmetric organization. The different cellular components cooperate and generate the general characteristics of cell polarization.

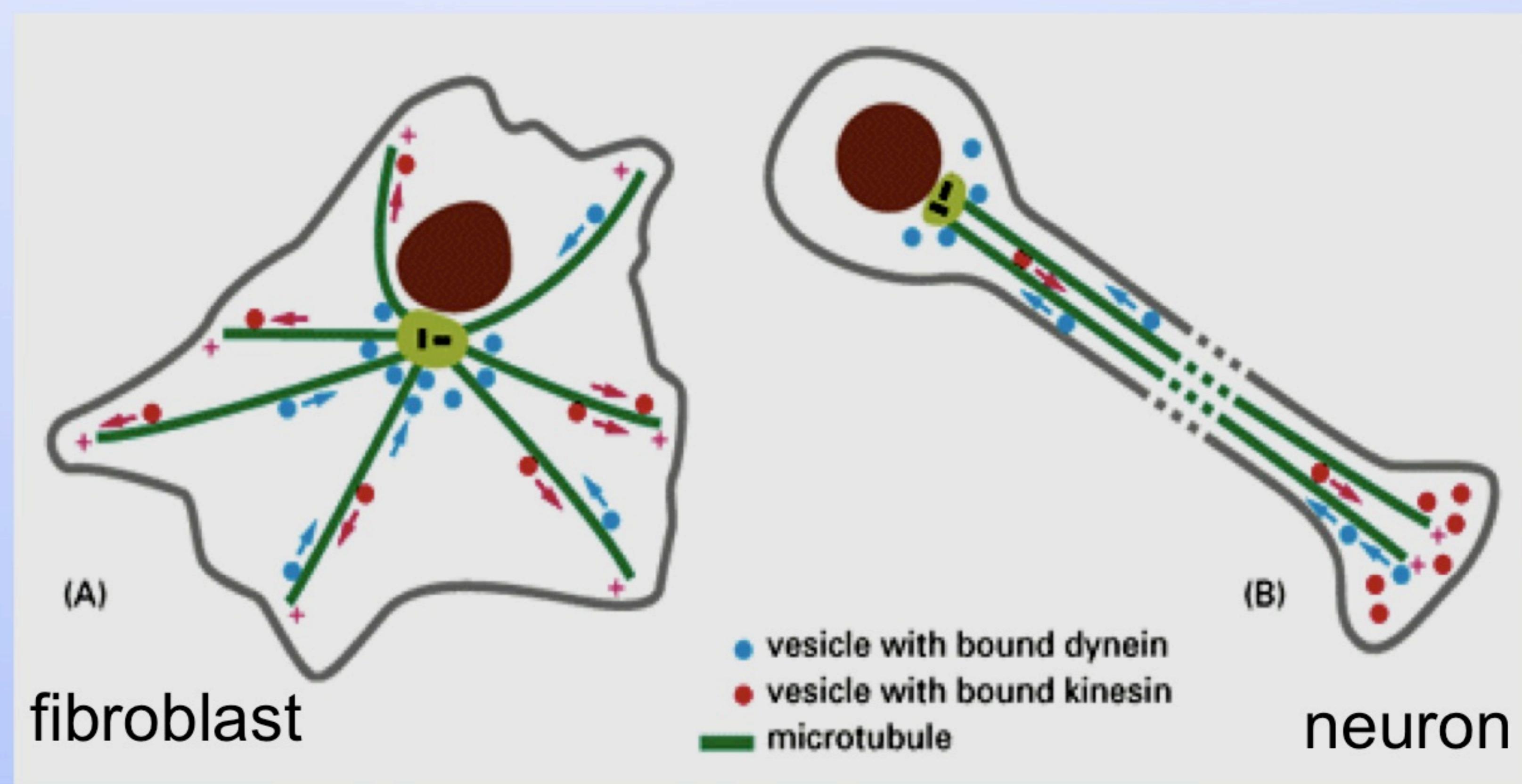
# 5. Cytosquelette et polarité

## b) Le rôle des microtubules et de leur régulateurs

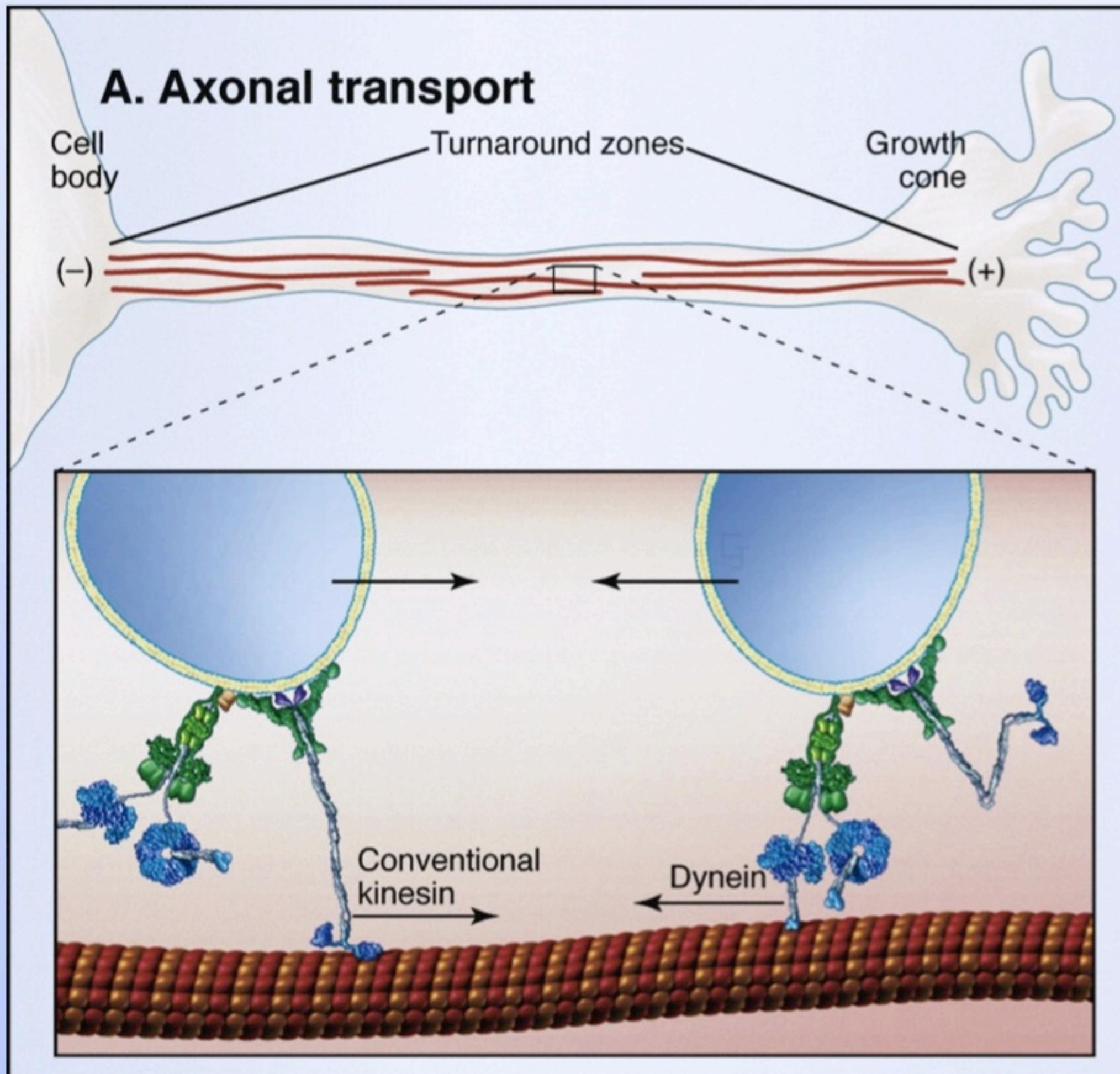




Motors carrying different cargoes in different directions

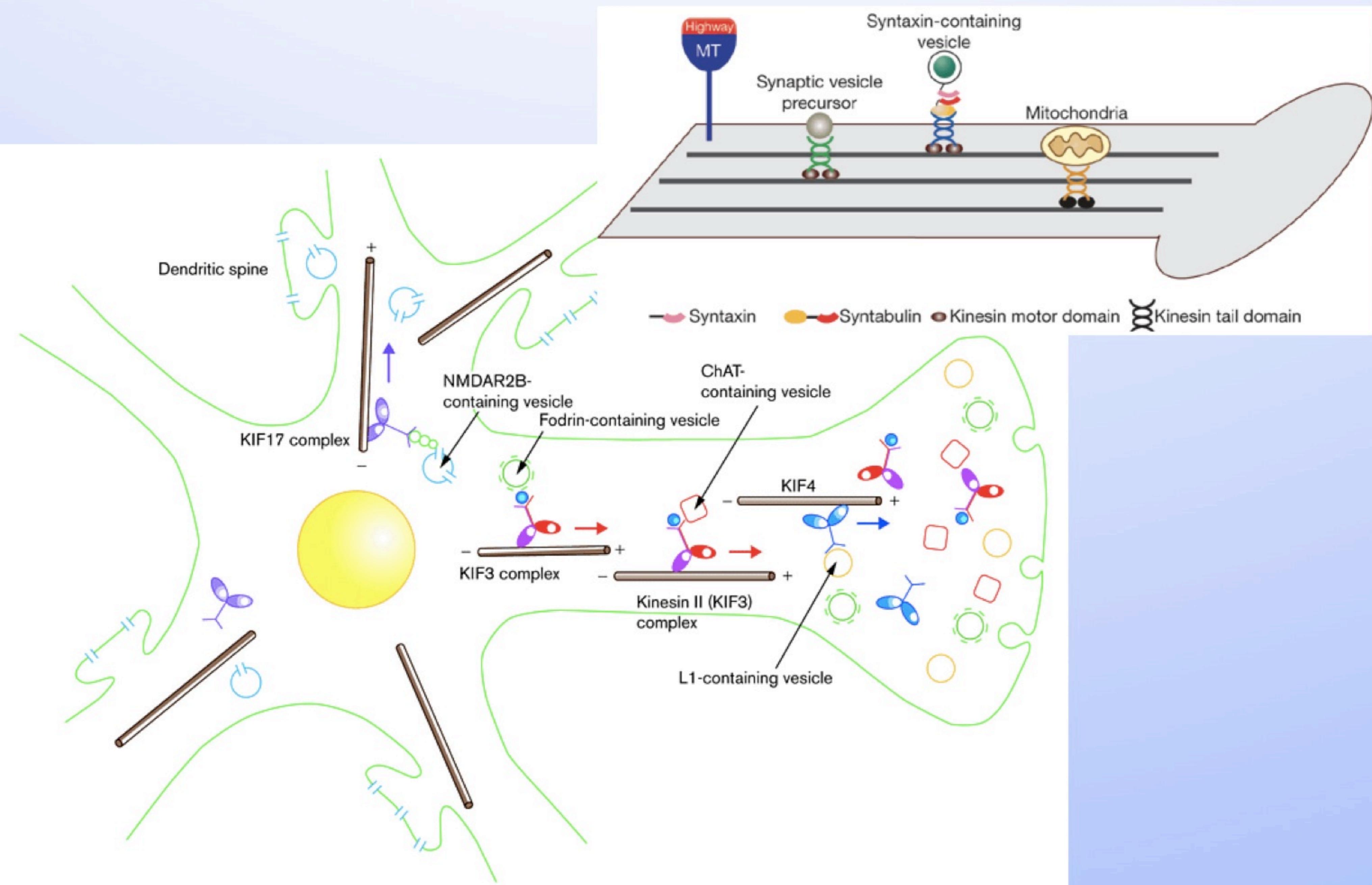


# Transport Anterograde et retrograde



Cargo structures	Overall rate (pulse labeling)	Instantaneous rate (light microscopy)	Directionality	Duty ratio
Golgi-derived vesicles (fast anterograde)	200–400 mm/d <sup>a</sup> (2–5 µm/s)	1–5 µm/s <sup>b</sup>	Anterograde	High
Endocytic vesicles, lysosomes, autophagosomes (fast retrograde)	100–250 mm/d <sup>a</sup> (1–3 µm/s)	1–3 µm/s <sup>b</sup>	Retrograde	High
Mitochondria	<70 mm/d <sup>c</sup> (<0.8 µm/s)	0.3–0.7 µm/s <sup>d</sup>	Bidirectional	Intermediate
Microfilaments, cytosolic protein complexes (slow component b)	2–8 mm/d <sup>e</sup> (0.02–0.09 µm/s)	Unknown	Unknown	Unknown
Microtubules, neurofilaments (slow component a)	0.2–1 mm/d <sup>e</sup> (0.002–0.01 µm/s)	0.3–1 µm/s <sup>f</sup>	Bidirectional	Low

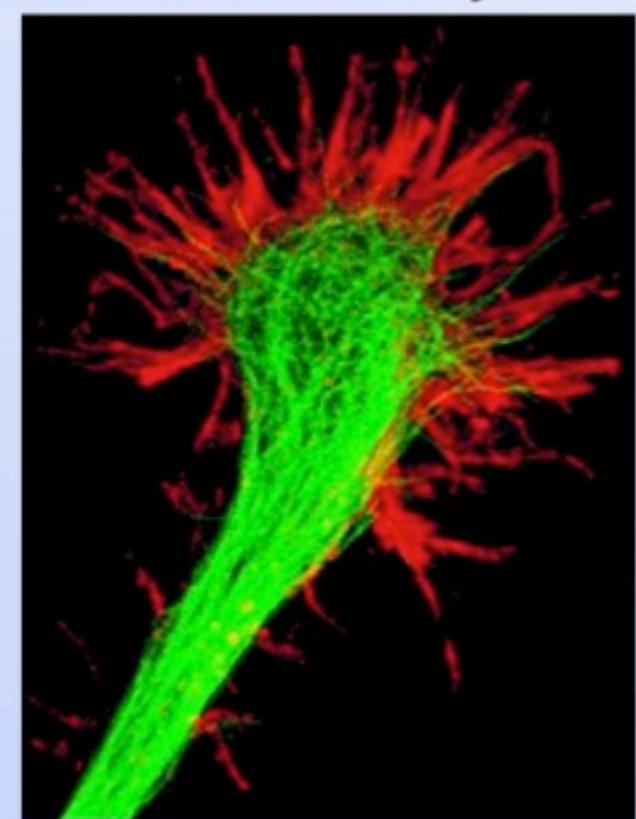
# Role des kinesines



# Microtubule stabilization by CRMP2

(Collapsin Response Mediator Protein 2 /unc33)

In cultured hippocampal neurons, one axon and several dendrites differentiate from a common immature process. Here we found that CRMP-2/TOAD-64/Ulip2/DRP-2 (refs. 2-4) level was higher in growing axons of cultured hippocampal neurons, that **overexpression of CRMP-2** in the cells led to the formation of **supernumerary axons** and that expression of truncated CRMP-2 mutants suppressed the formation of primary axon in a dominant-negative manner. Thus, CRMP-2 seems to be critical in axon induction in hippocampal neurons, thereby establishing and maintaining neuronal polarity.

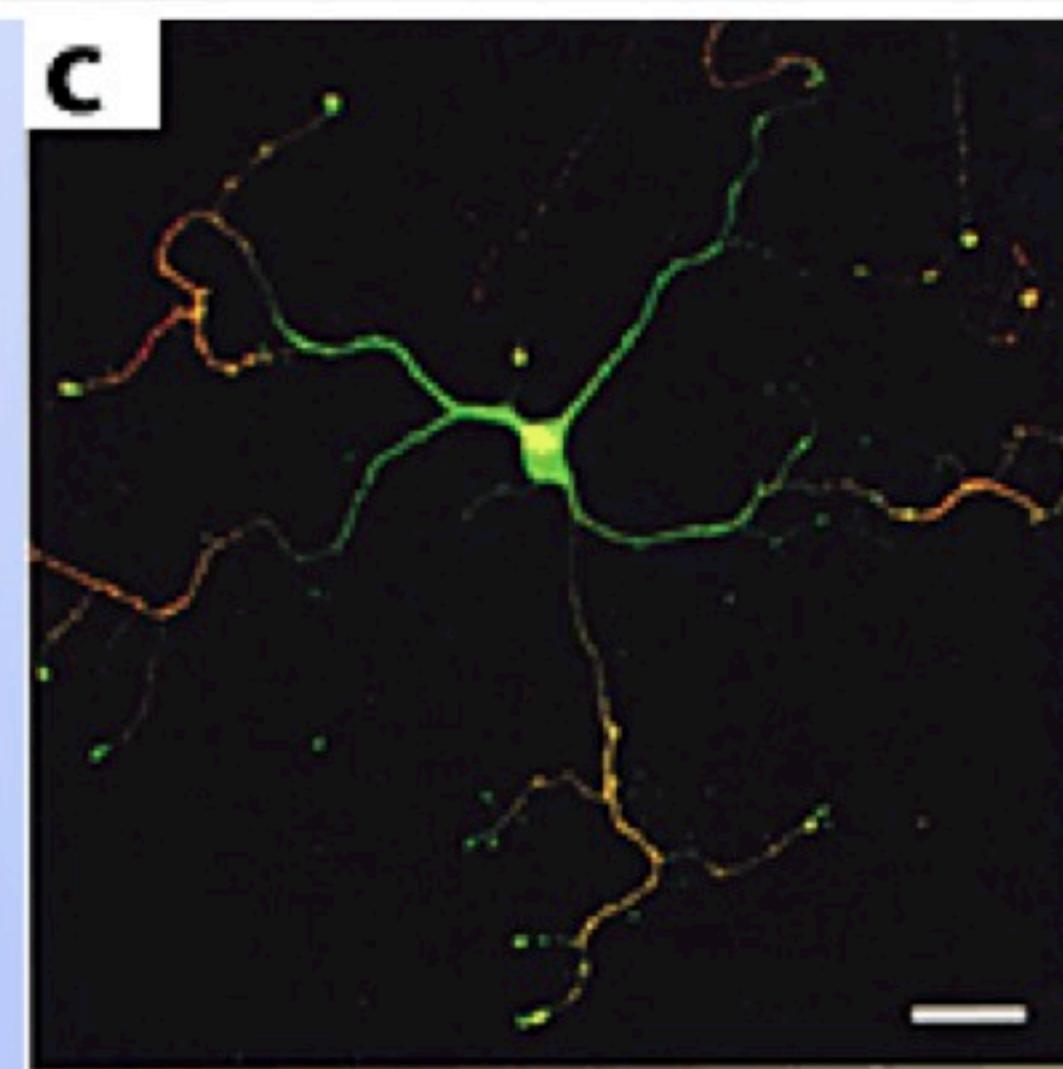
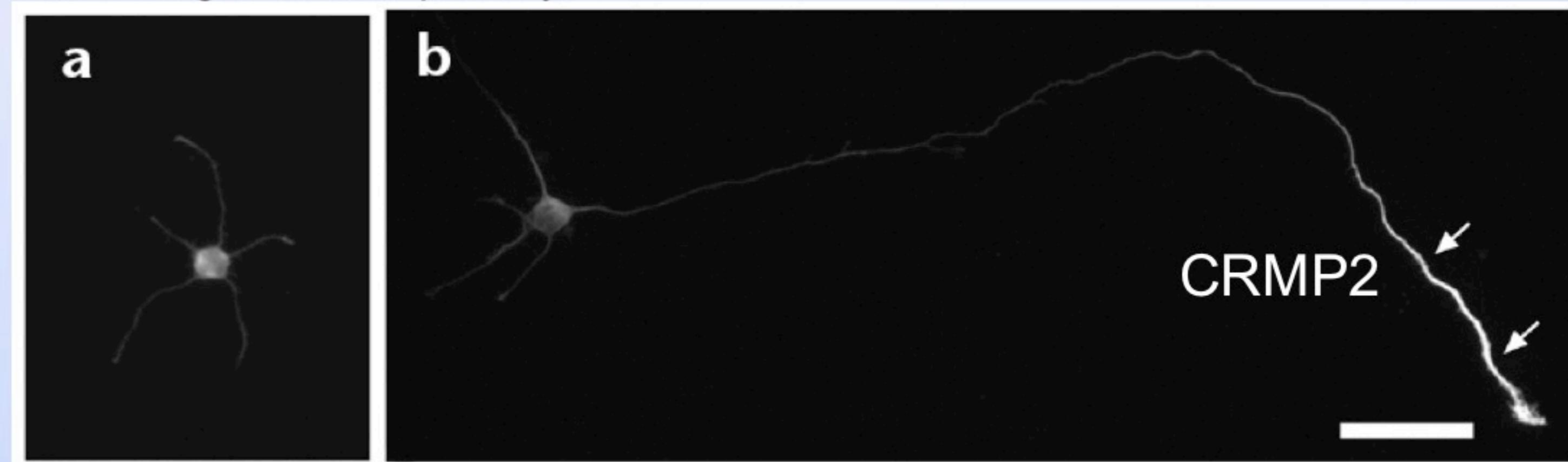


Red: actin

**Green: microtubule**

CRMP2 interagit avec les hétérodimères de tubuline et favorise l'assemblage des microtubules in vitro.

Surexpression de CRMP2: conversion de dendrites en axone surnuméraires.



Inagaki et al., Nat Neurosci. 2001 Aug;4(8):781-2.

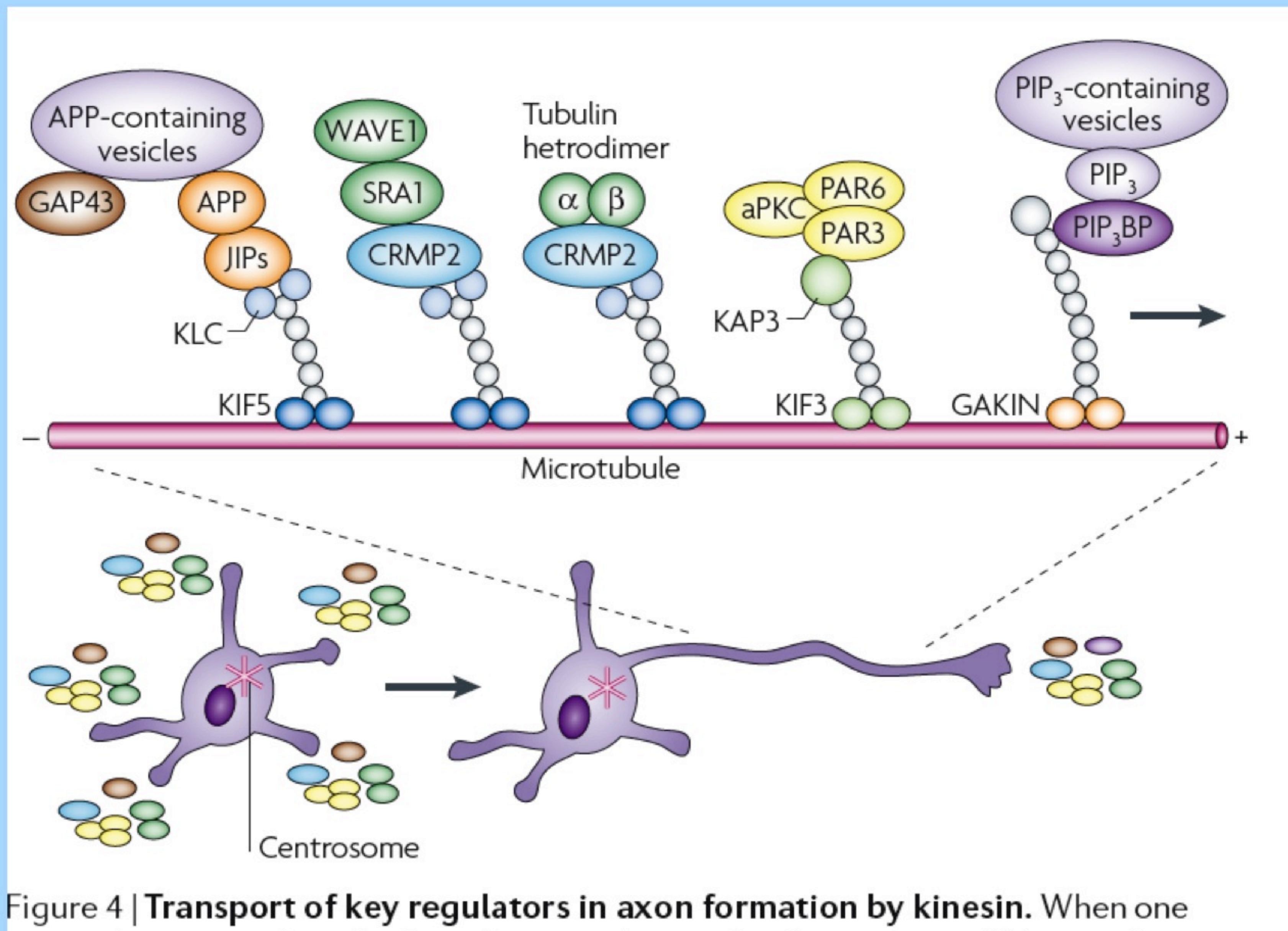
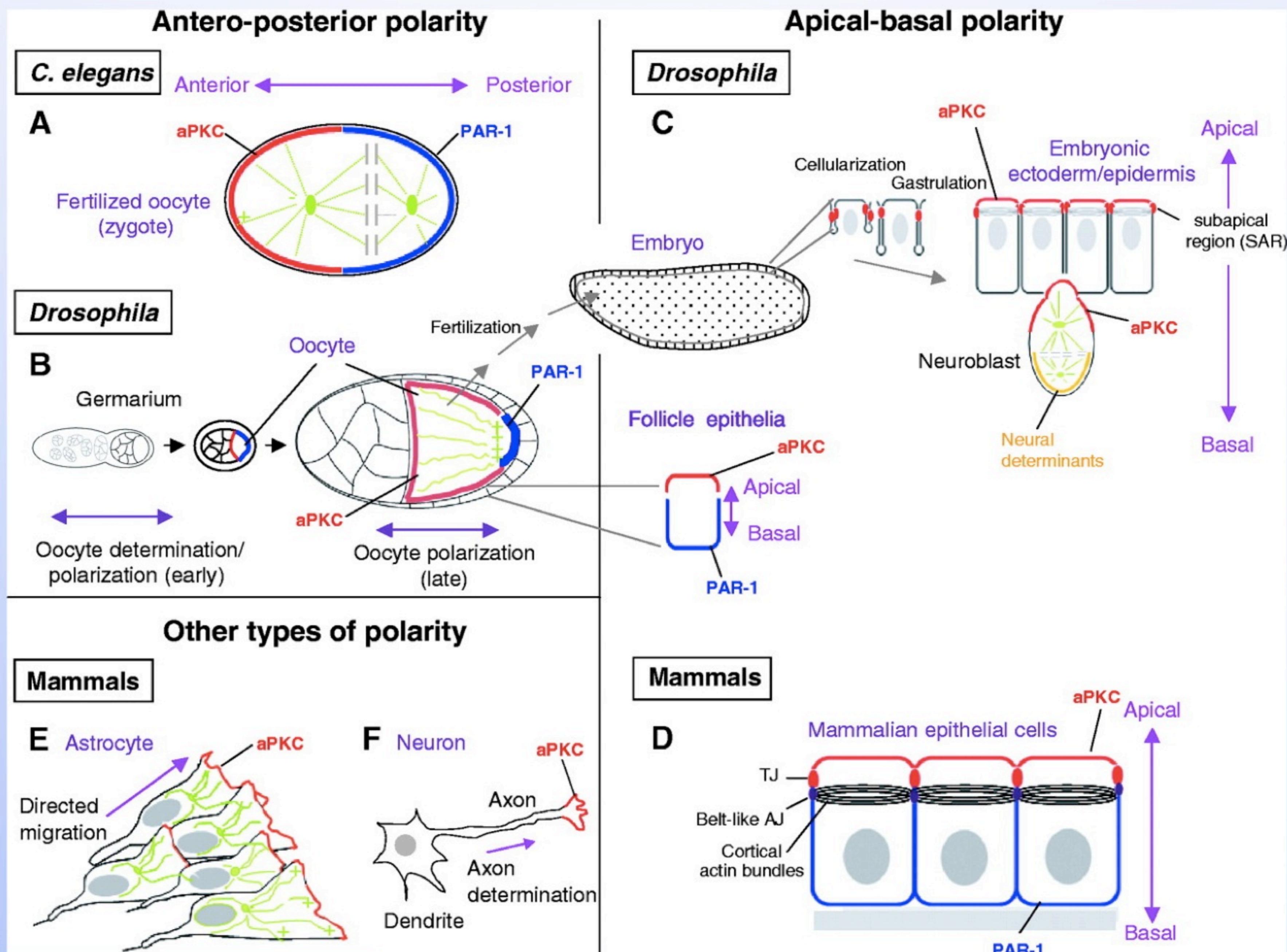


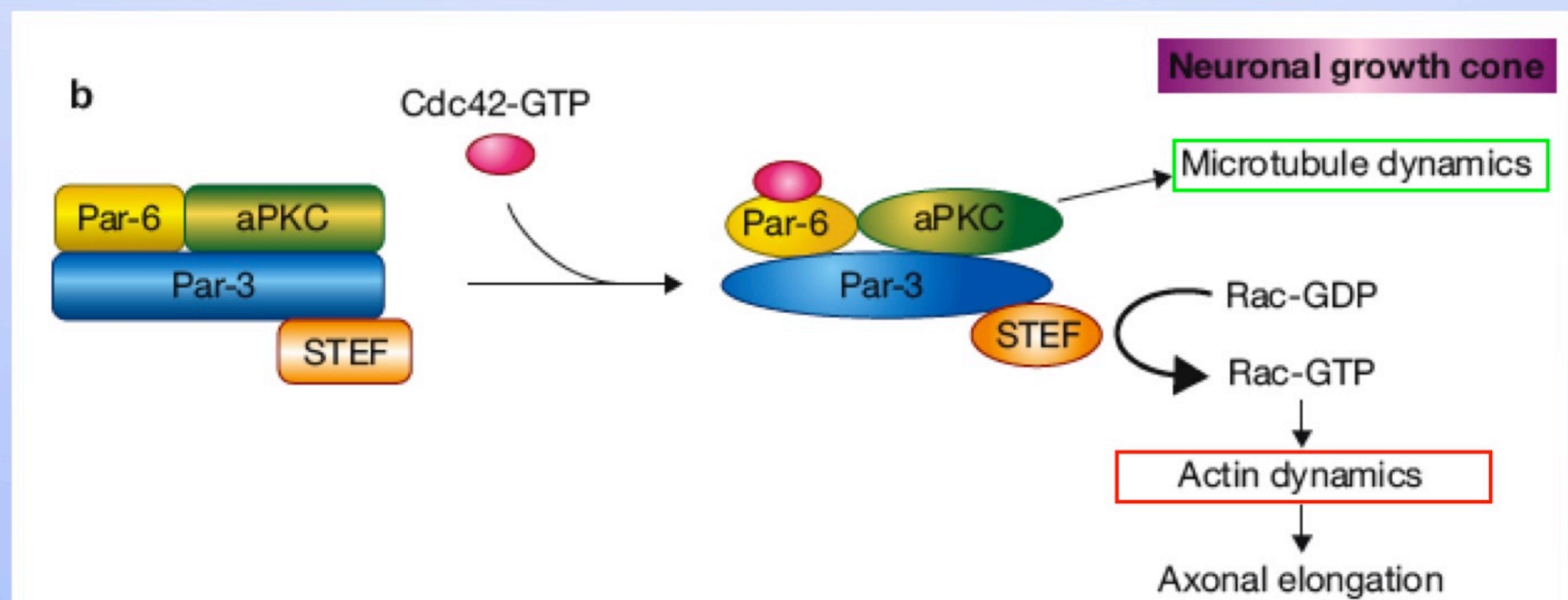
Figure 4 | Transport of key regulators in axon formation by kinesin. When one

## Various types of cell polarity in which the PAR-aPKC system is involved



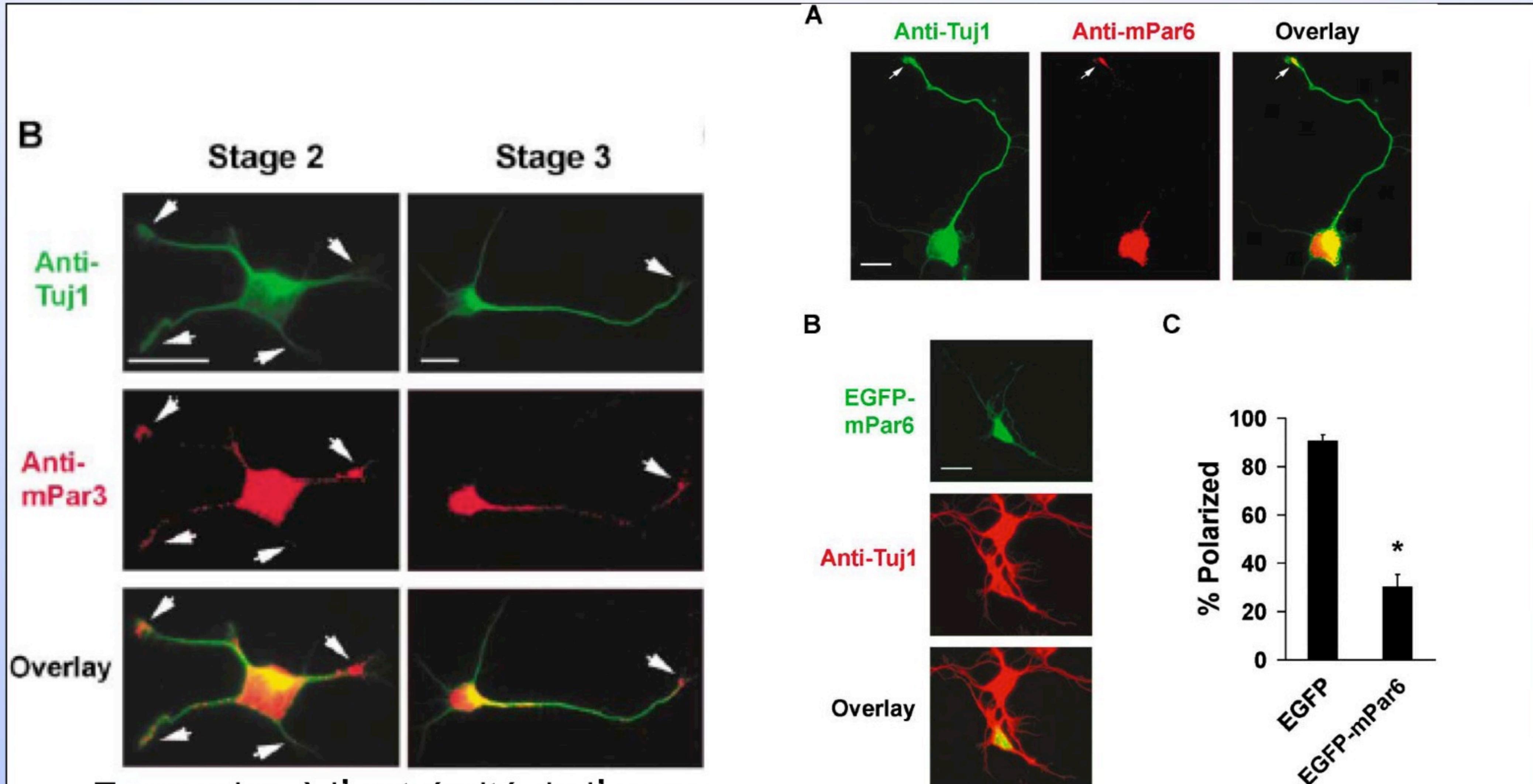
**Table 1. Localization of proteins of the PAR-aPKC complex in different species**

<i>S. pombe</i>	<i>C. elegans</i>	<i>Drosophila</i>	Mammal	Localization				Mammal
				<i>C. elegans</i>	late oocyte	<i>Drosophila</i>	neuroblast	
–	PAR-3	Bazooka	PAR-3/ASIP*	zygote	Anterior cortex	SAR/apical membrane	Apical cortex	TJ <sup>†</sup> /apical membrane
–	PAR-6	PAR-6	PAR-6 $\alpha, \beta, \gamma$					
–	PKC-3	aPKC	aPKC $\lambda, \zeta$					



The evolutionary conserved polarity protein:

# Par3 and Par 6 complex and PI3K



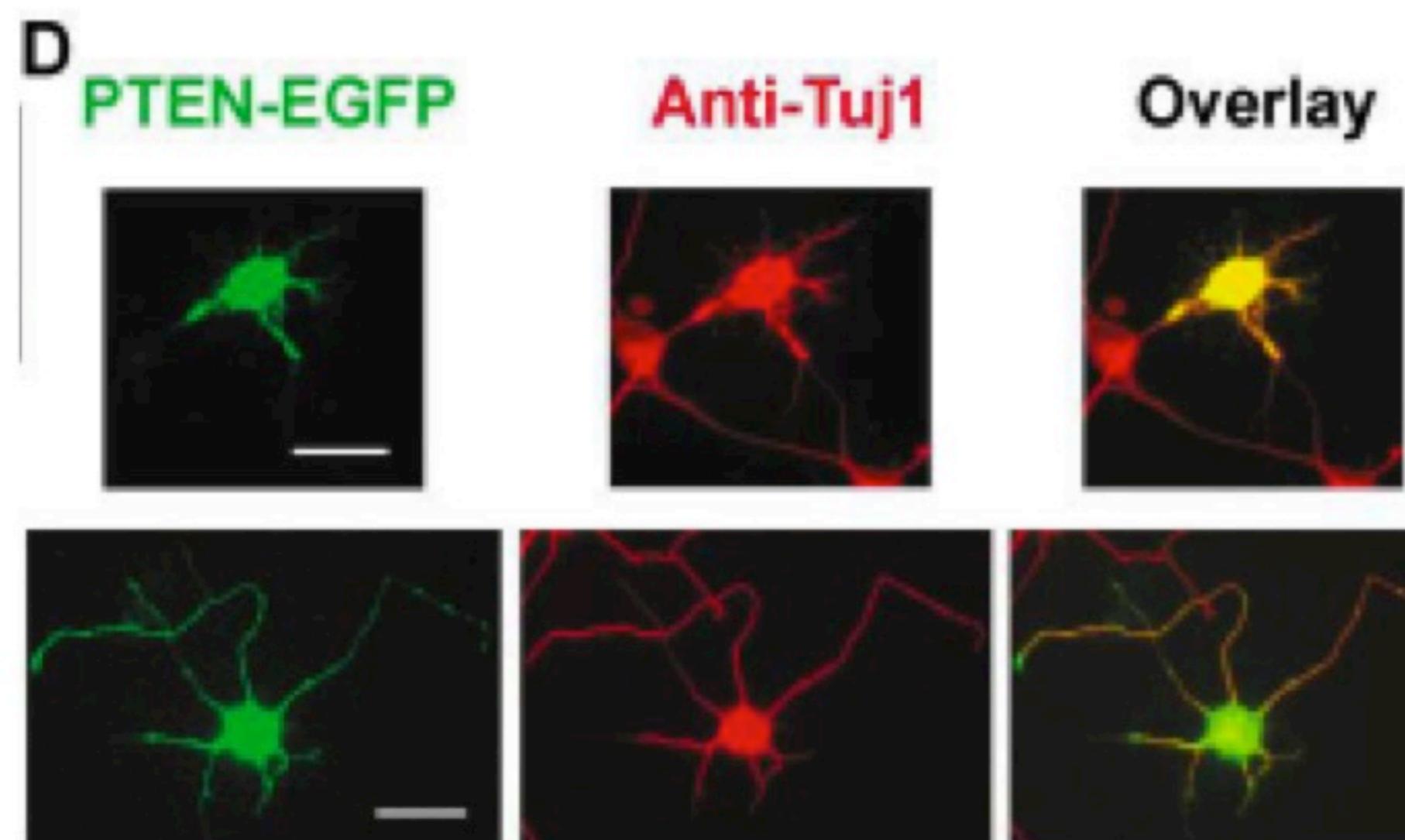
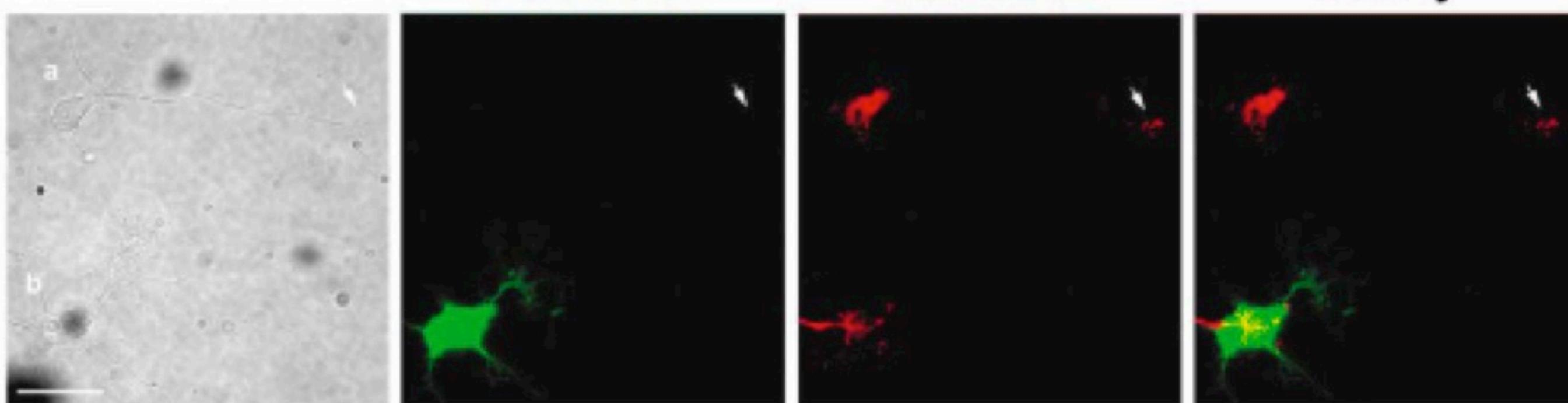
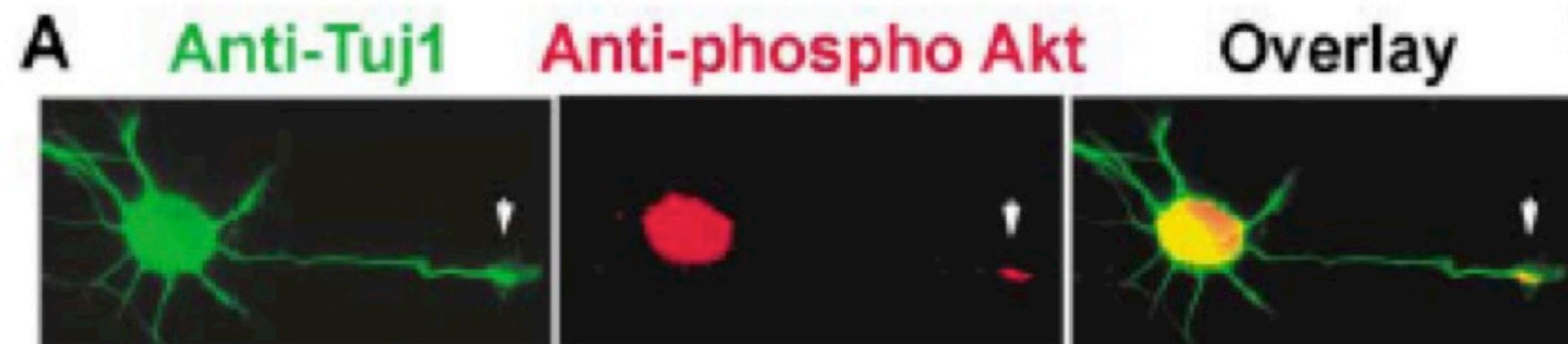
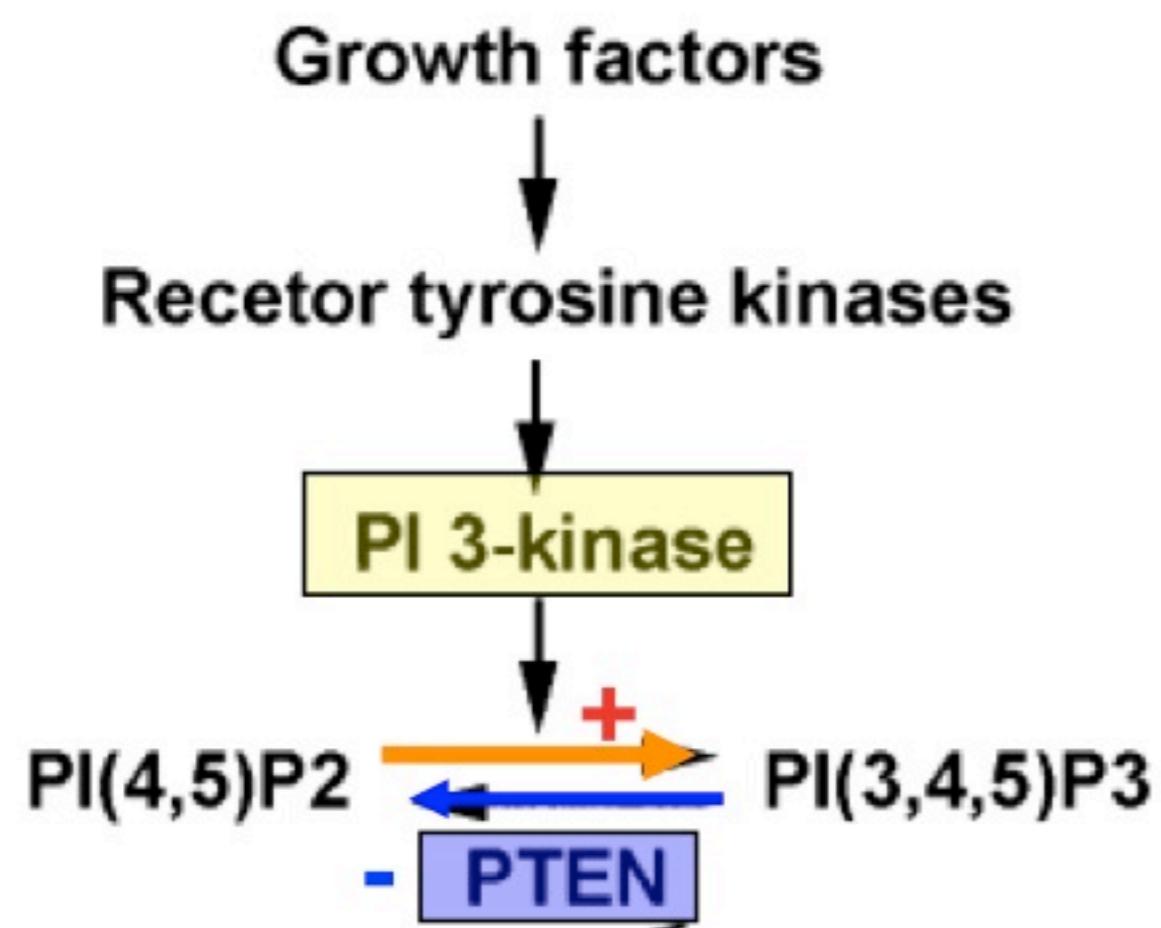
Expression à l' extrémité de l' axone

Surexpression Par6: perte de l' axone,  
neurites multiples: perte de polarité

The evolutionary conserved polarity protein:

# Par3 and Par 6 complex and PI3K

PI3K phosphoryle AKT au bout de l' axone



Les neurones exprimant PTEN, expriment par3 dans le soma et pas au bout de l' axone.

Les neurones exprimant PTEN, ne possède pas d' axone unique mais de multiples prolongements.

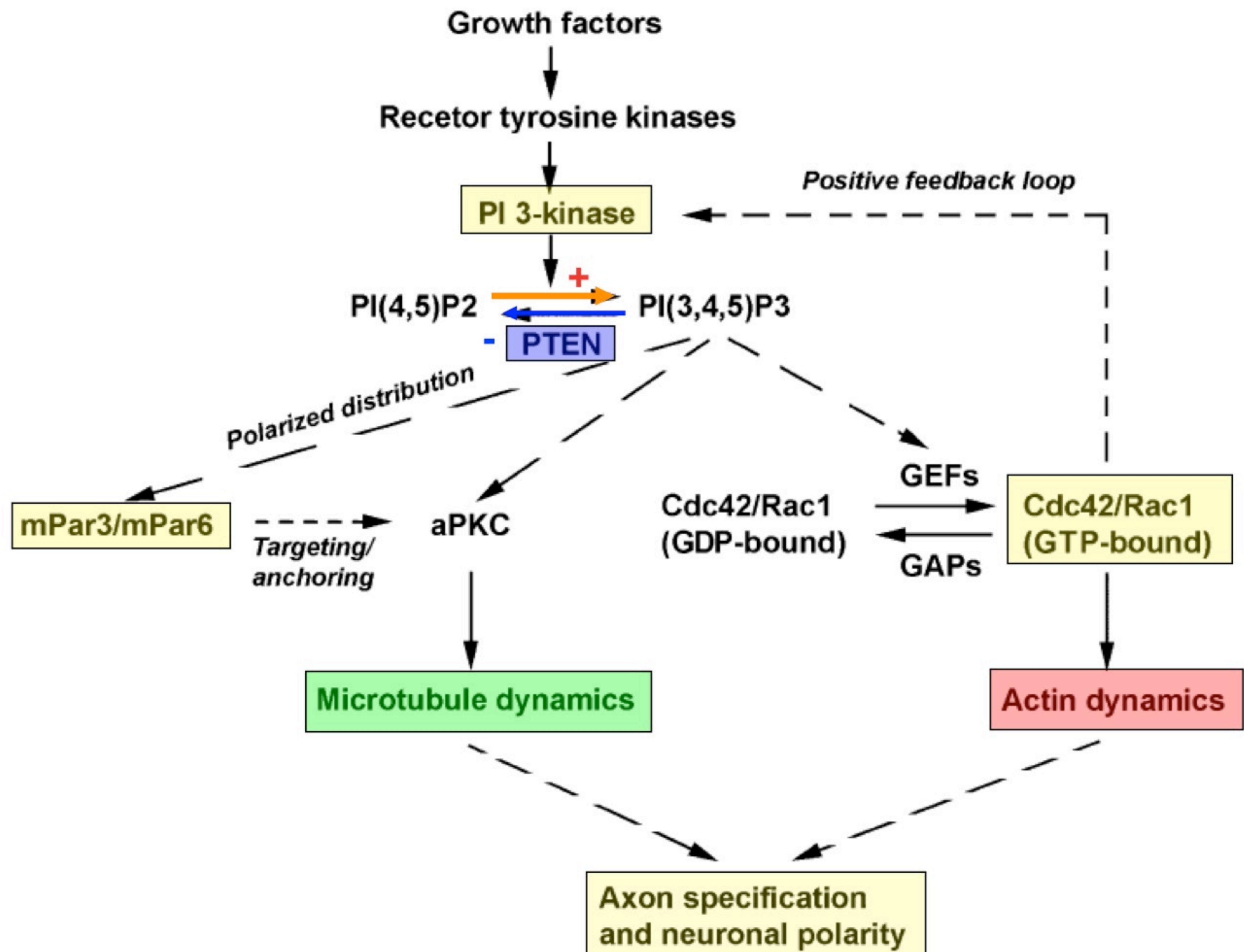
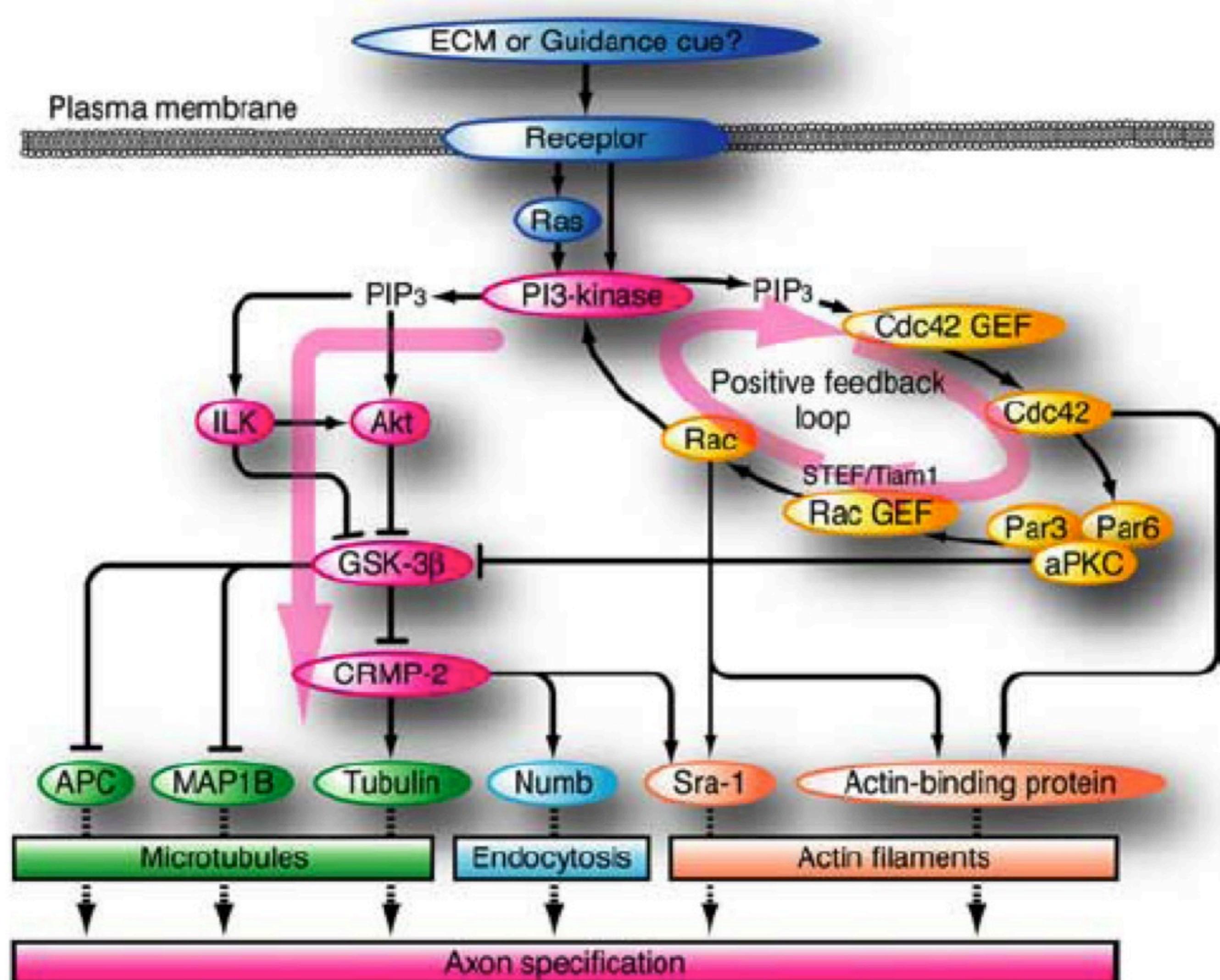


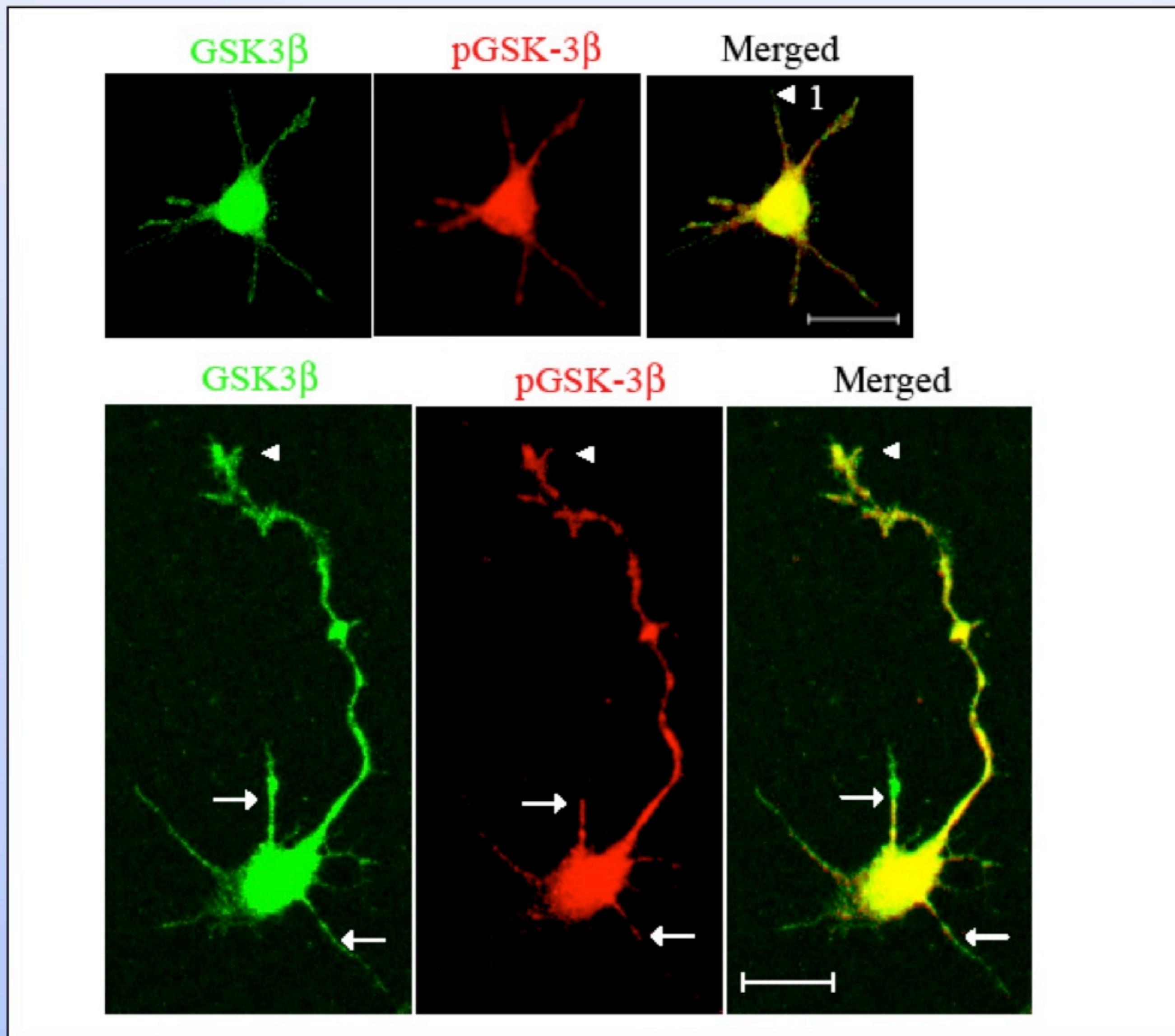
Figure 8. A Model for the Signaling Processes that Specify Neuronal Polarity



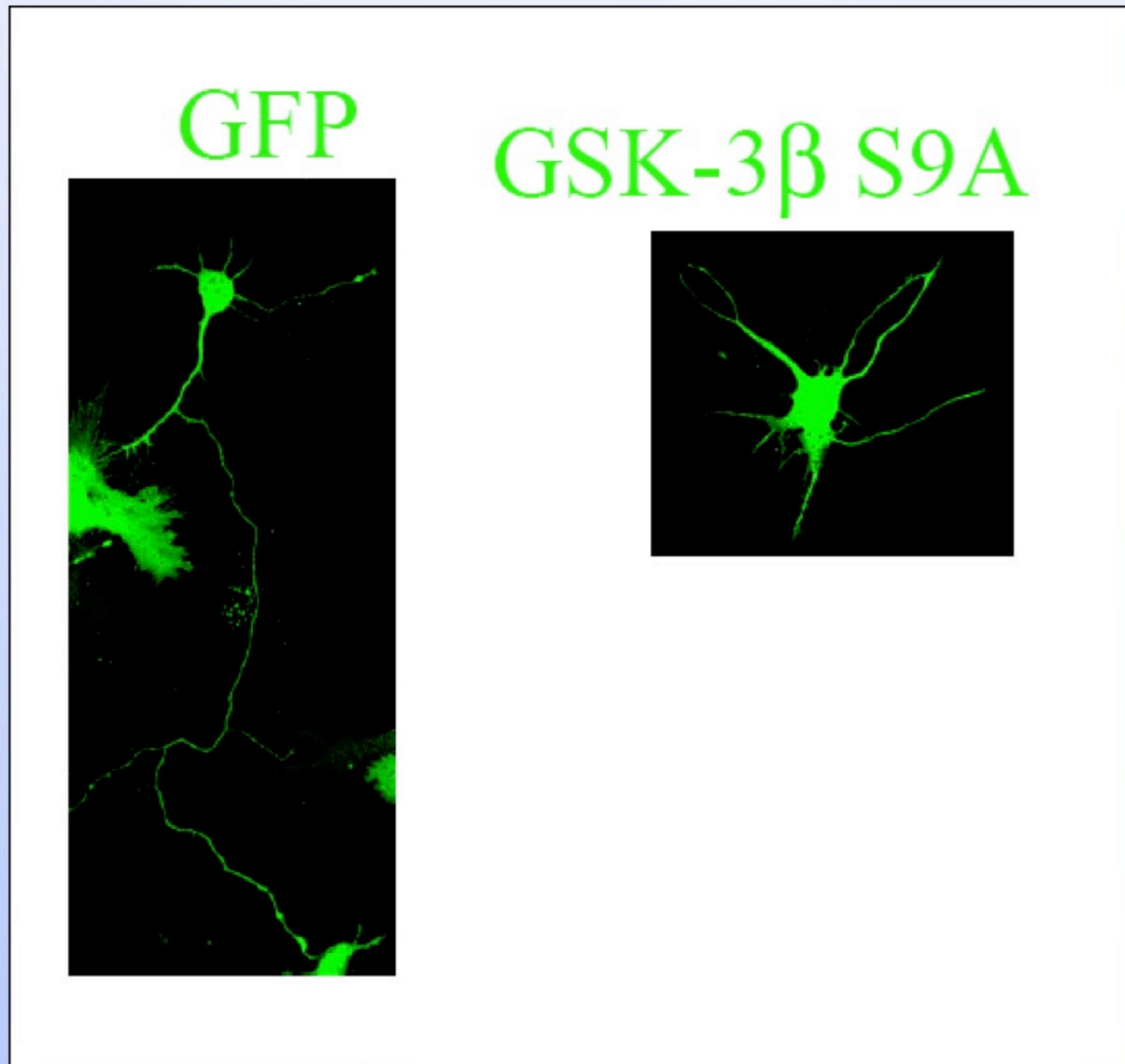
**Figure 2.** Signaling cascades in axon specification. In one immature neurite (the future axon), the extracellular matrix (ECM) activates PI3-kinase through interaction with adhesion molecules or receptors, thereby producing PIP<sub>3</sub>. Accumulated PIP<sub>3</sub> drives two major signaling cascades: the Akt/GSK-3 $\beta$ /CRMP-2 pathway and the positive feedback loop composed of Cdc42, the Par complex, and Rac1. These signaling cascades regulate cytoskeletons, endocytosis, protein trafficking, and transcriptions to promote neurite elongation and to determine axon or dendrite fate.

# GSK-3 $\beta$ in neuronal polarity

Polarized inactivation of GSK-3 $\beta$  (phosphorylation) in axon



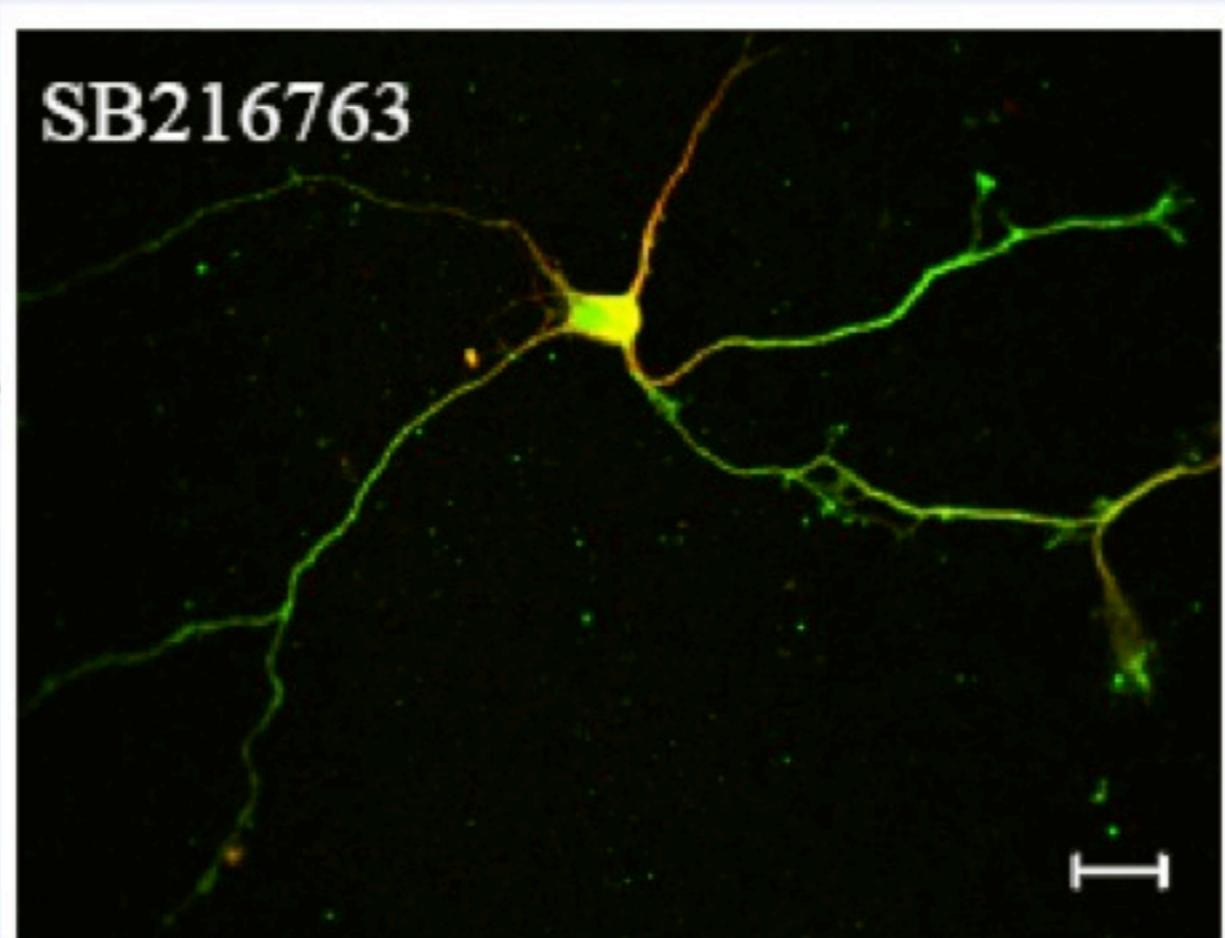
# Inhibition de l'induction de l'axone par un mutant de GSK-3 beta constitutivement actif: GSK-3 beta S9A



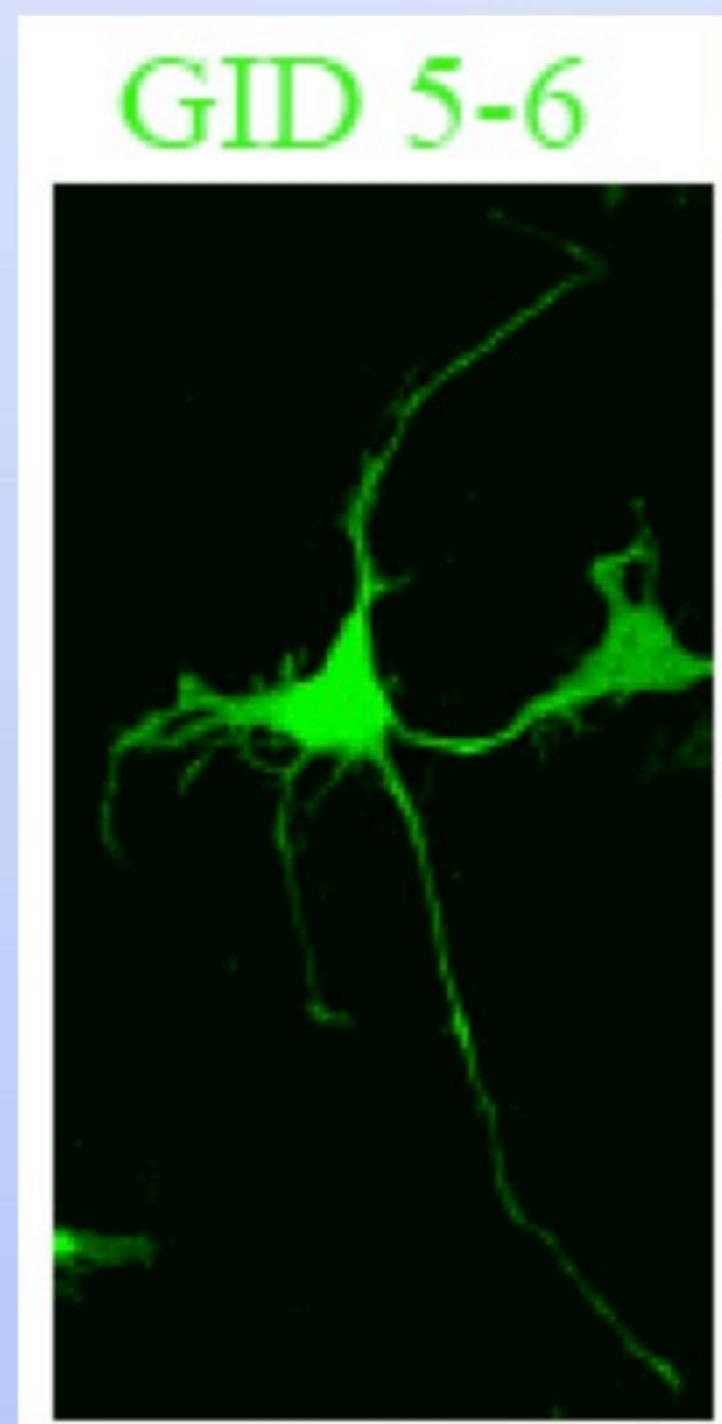
L'activation prolongée de la GSK3b inhibe la formation de l' axone.

# L'inhibition de l'activité de la GSK-3 beta induit la formation de multiples axones.

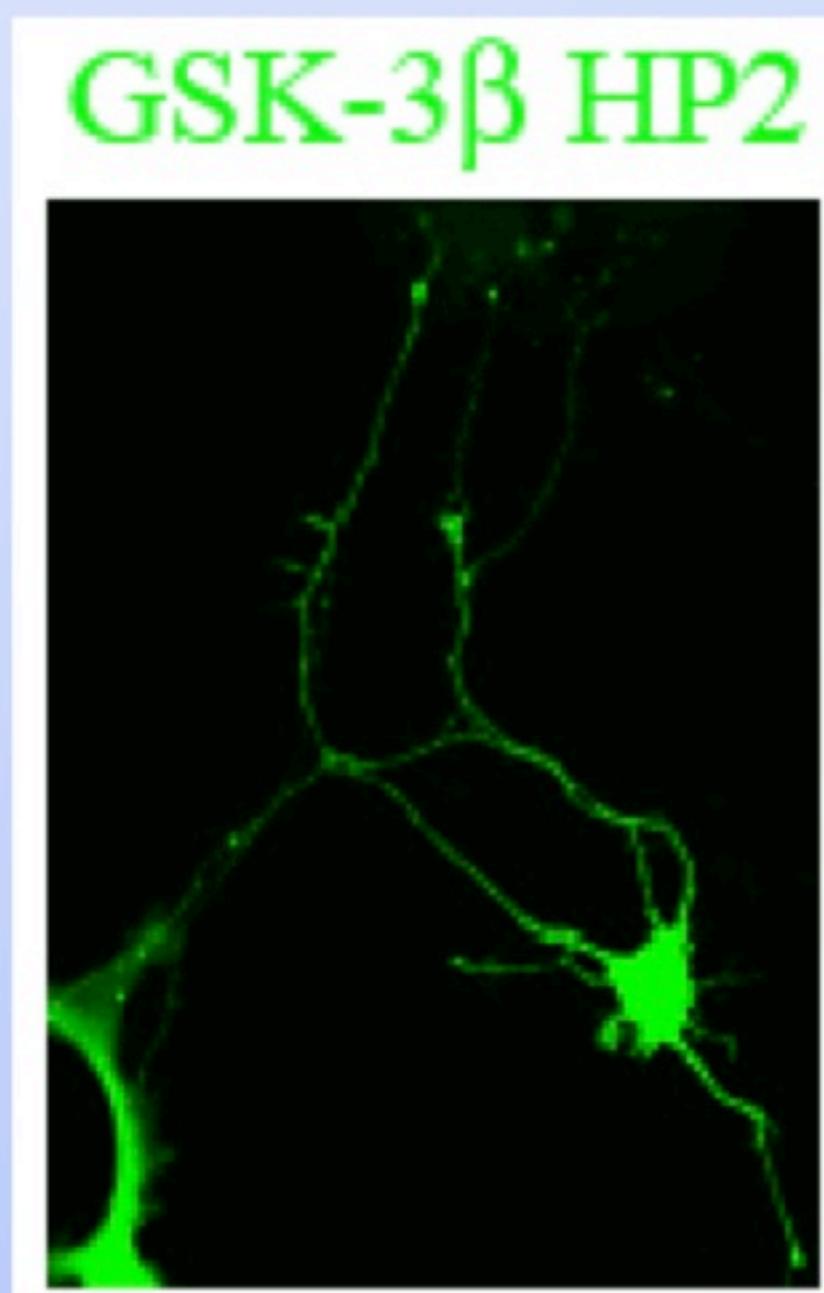
Inhibiteurs chimiques  
ou transfection de la forme  
active d'Akt.



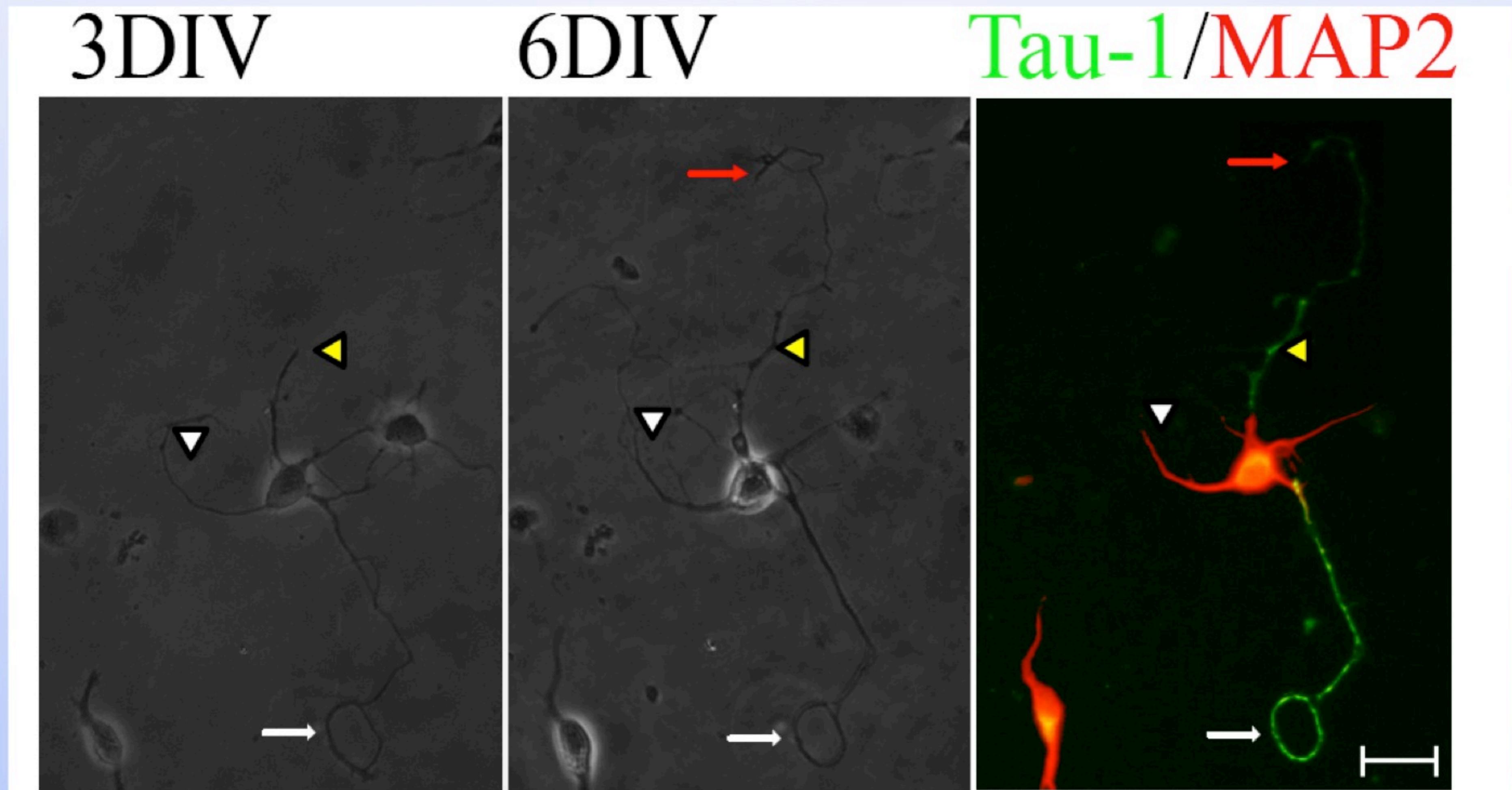
Peptide inhibitor



RNAi



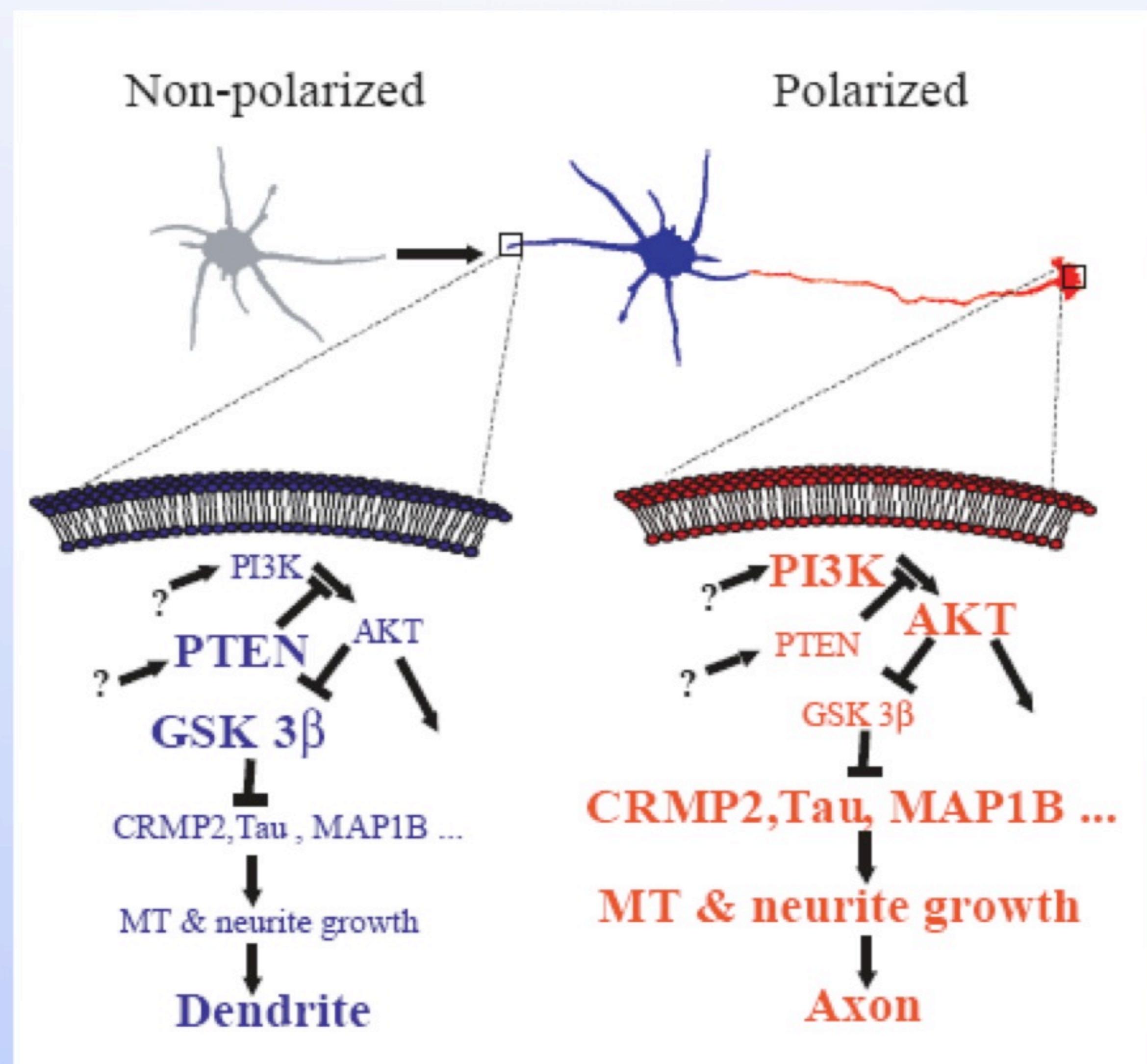
# L'inhibition de GSK3 au stade 3 convertit des dendrites en axone.



---

SB415286 addition time

# Modele



Lors de la croissance neuritique du futur axone, la PI3K phosphoryle le PIP3 et active Akt qui va inhiber GSK3beta. Cela résulte en la phosphorylation de CRMP2 et qui promeut la formation de microtubules et la croissance axonale.

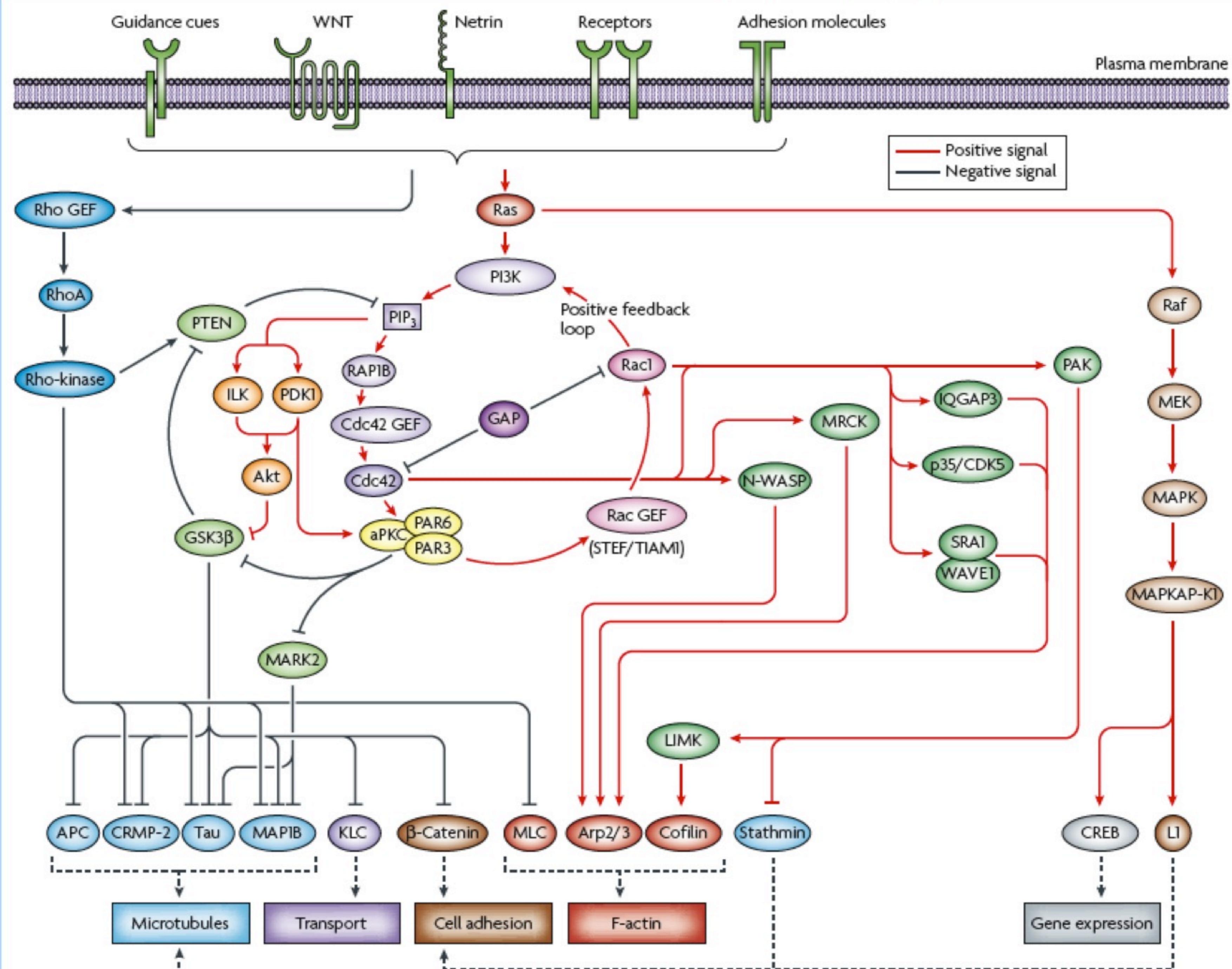
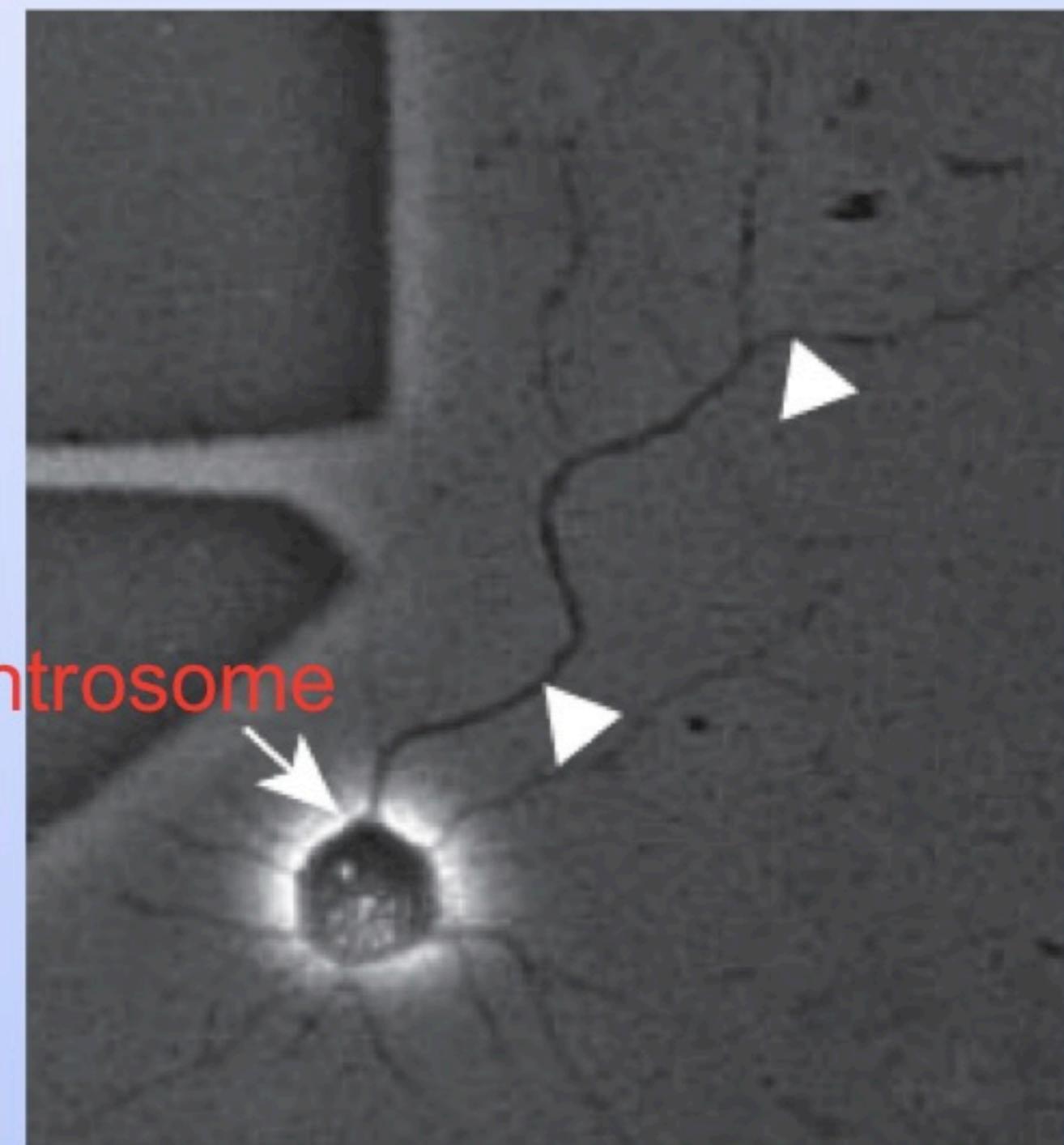
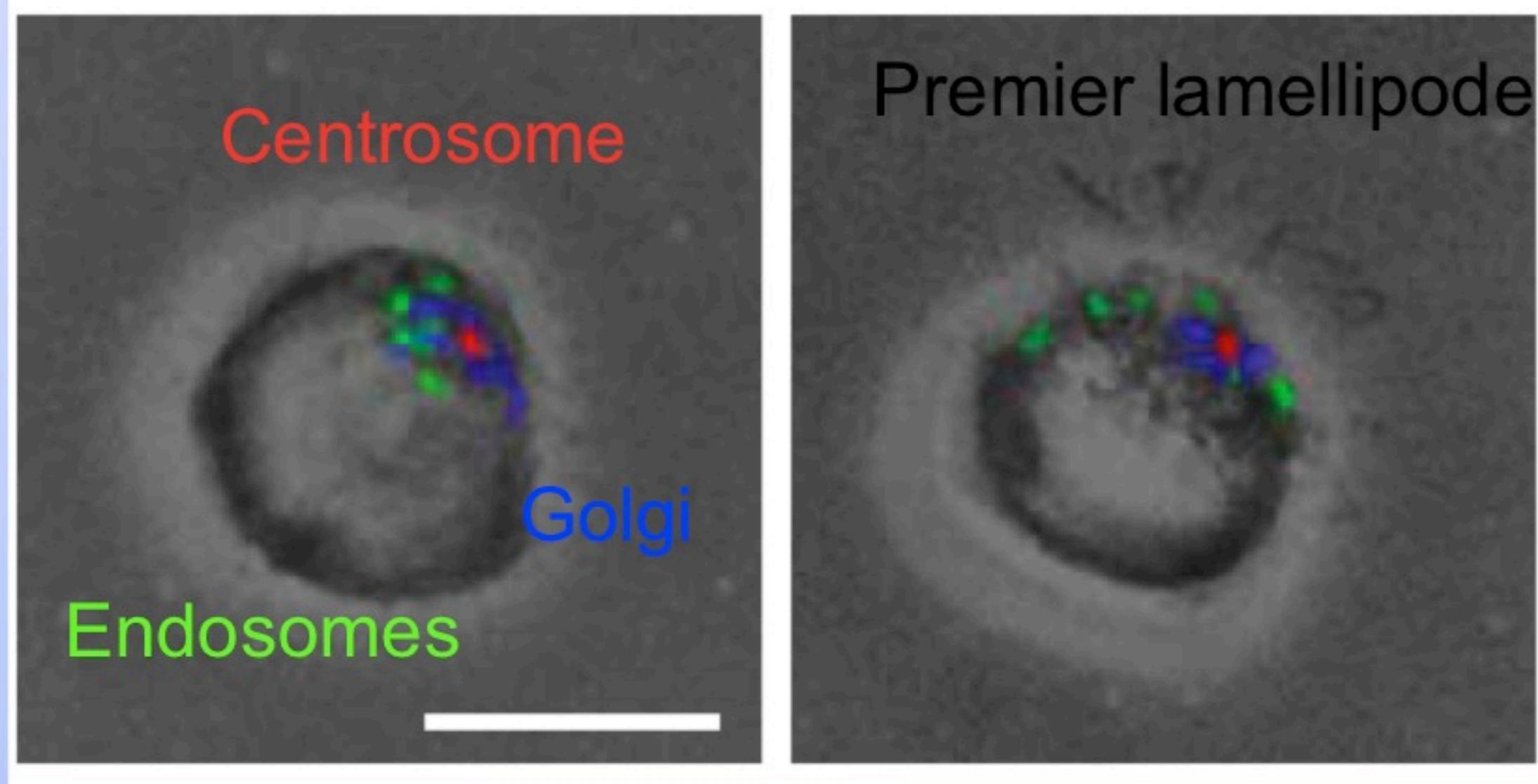
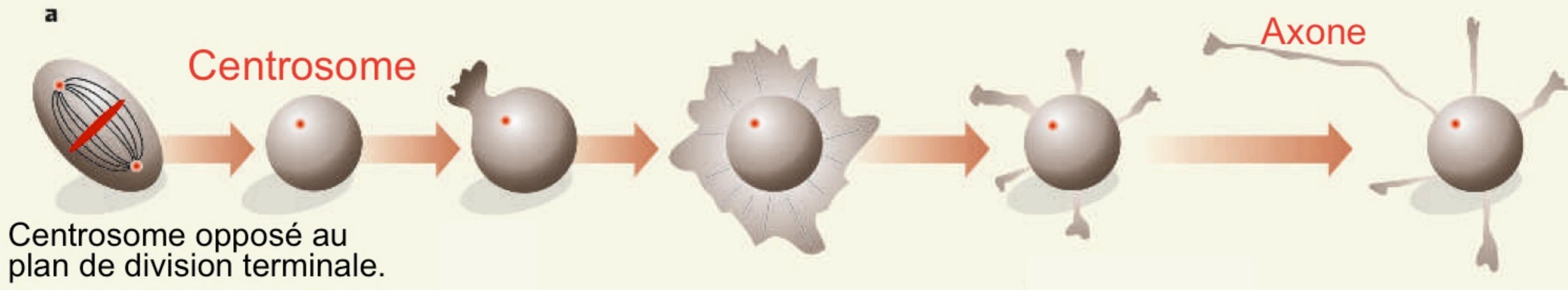
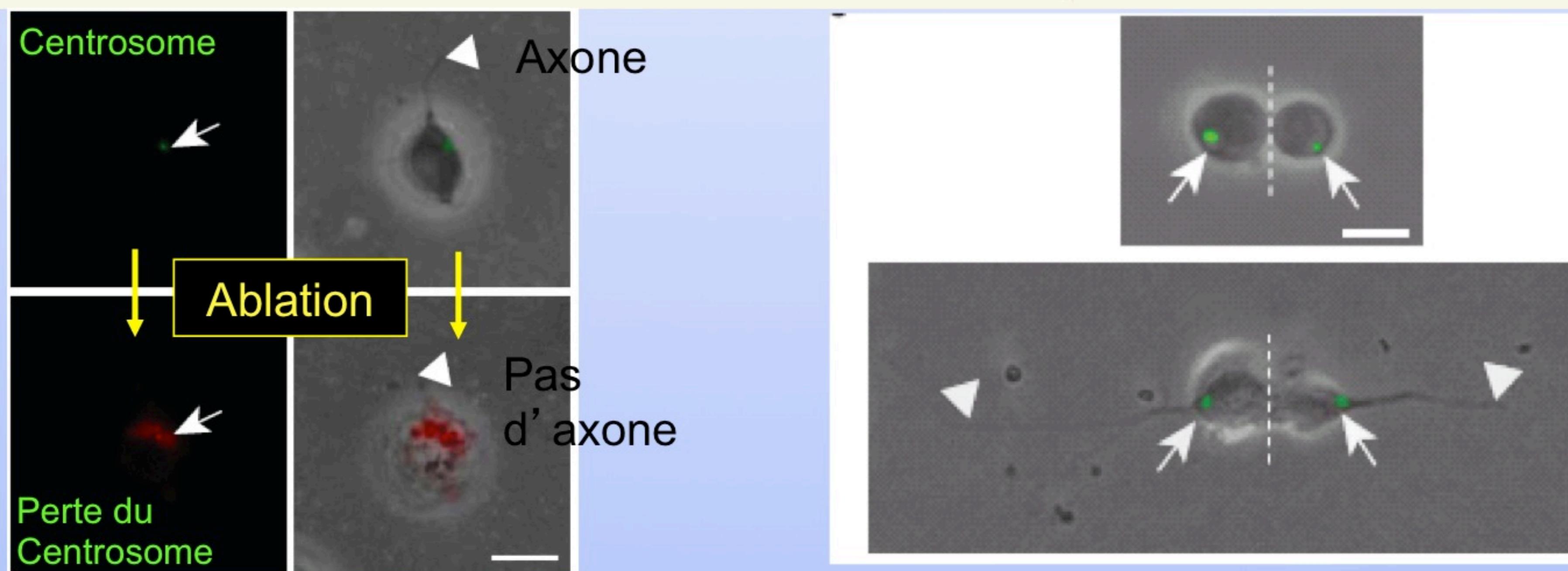
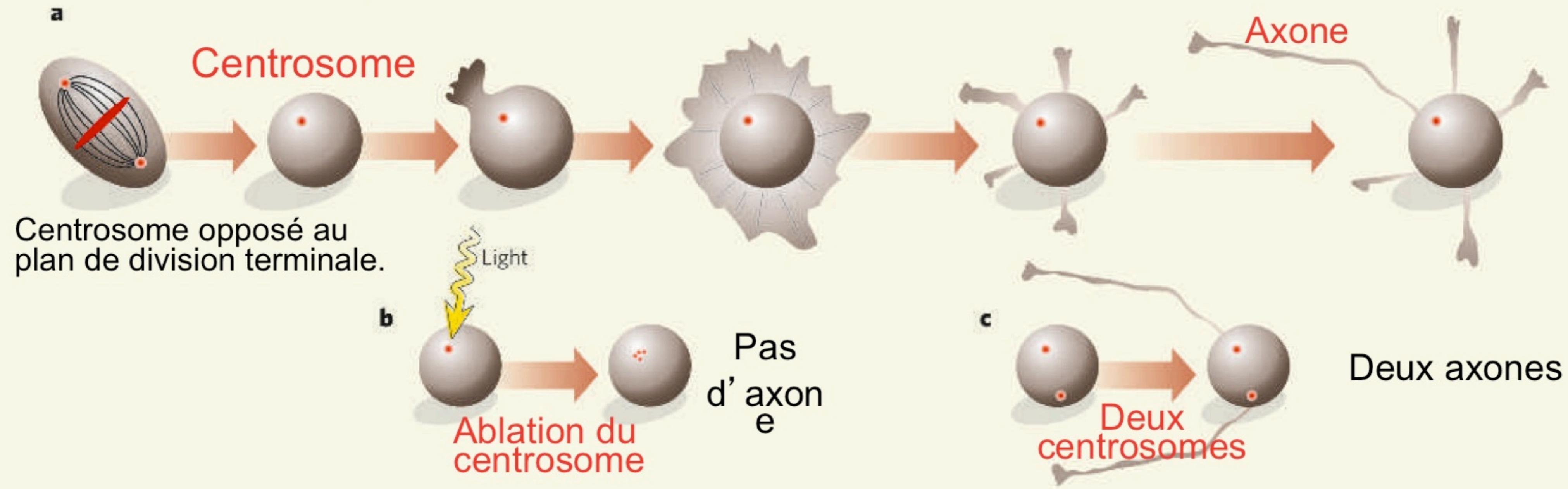


Figure 3 | Simplified model of positive and negative signals. In response to the activation of cell surface receptors by

# La localisation du centrosome détermine la polarisation d'un neurite en axone.



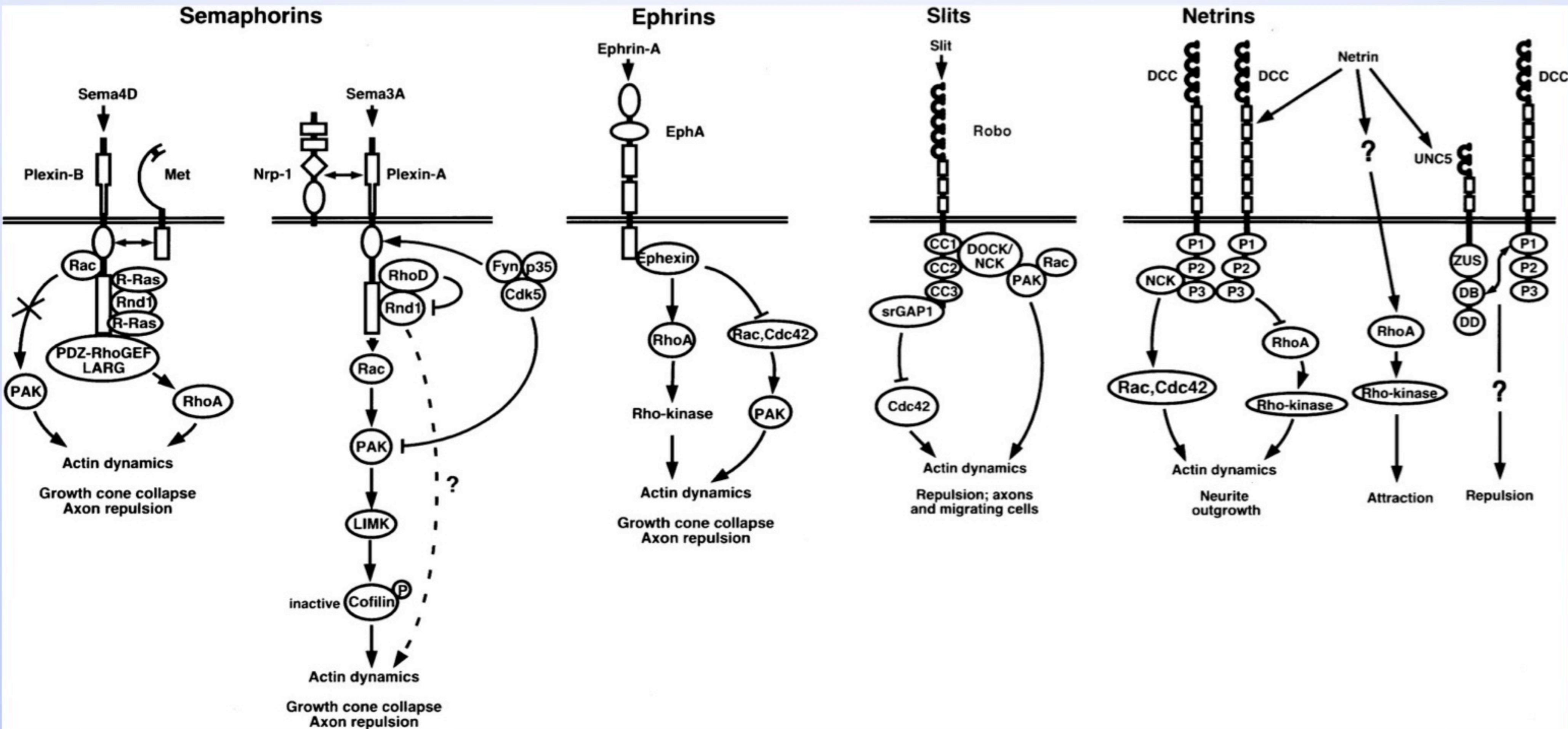
# La localisation du centrosome détermine la polarisation d'un neurite en axone.



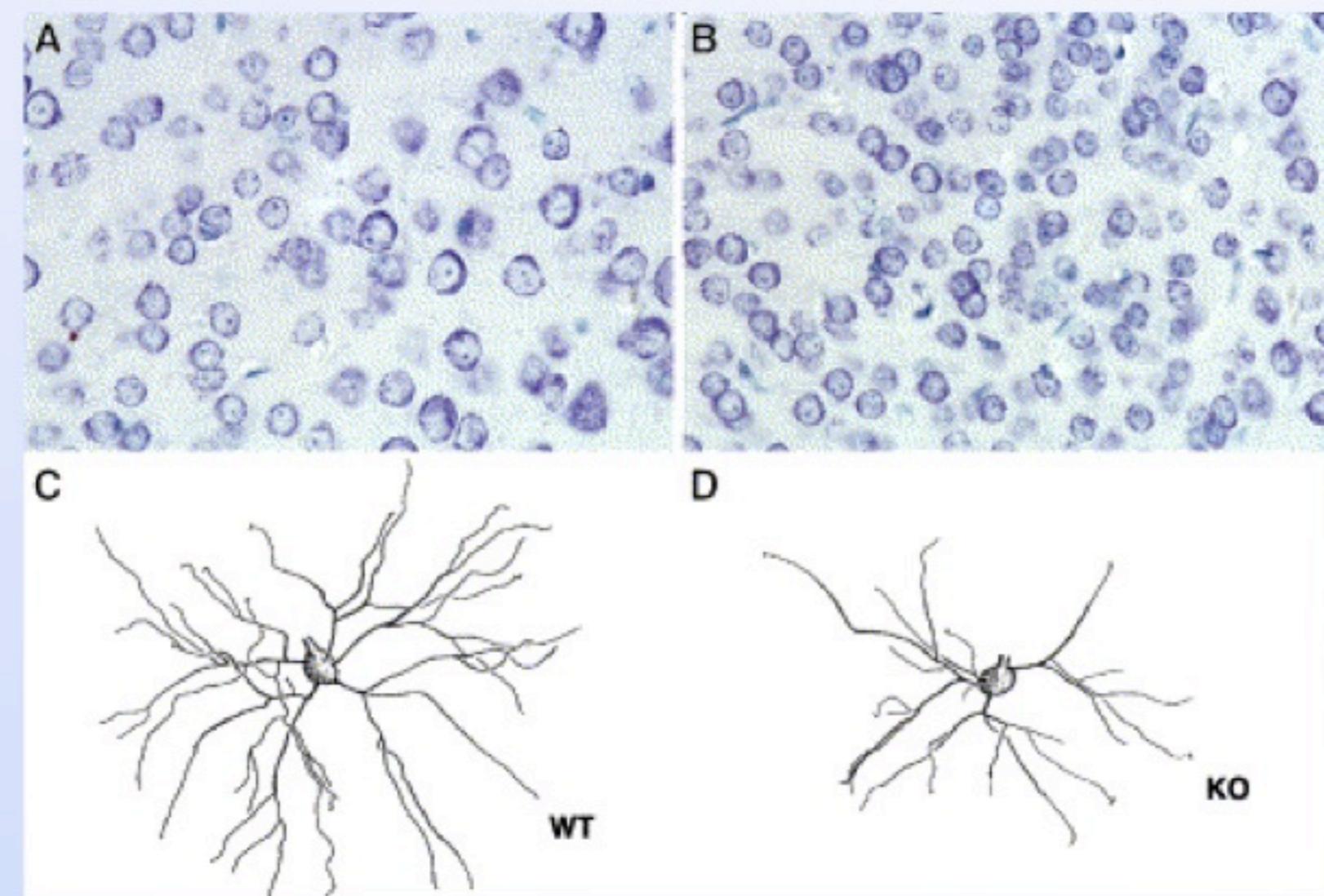
*La polarité cellulaire peut être régulée par des molécules extracellulaire agissant sur des récepteurs présents sur les cônes de croissance.*

1. Récepteurs membranaires (GPCRs, RTKs) détectant un signal asymétrique extérieur.
2. Les récepteurs activent les GTPase Rho.
3. Les protéines Rho induisent des changement de cytosquelette induce cytoskeletal changes at the leading

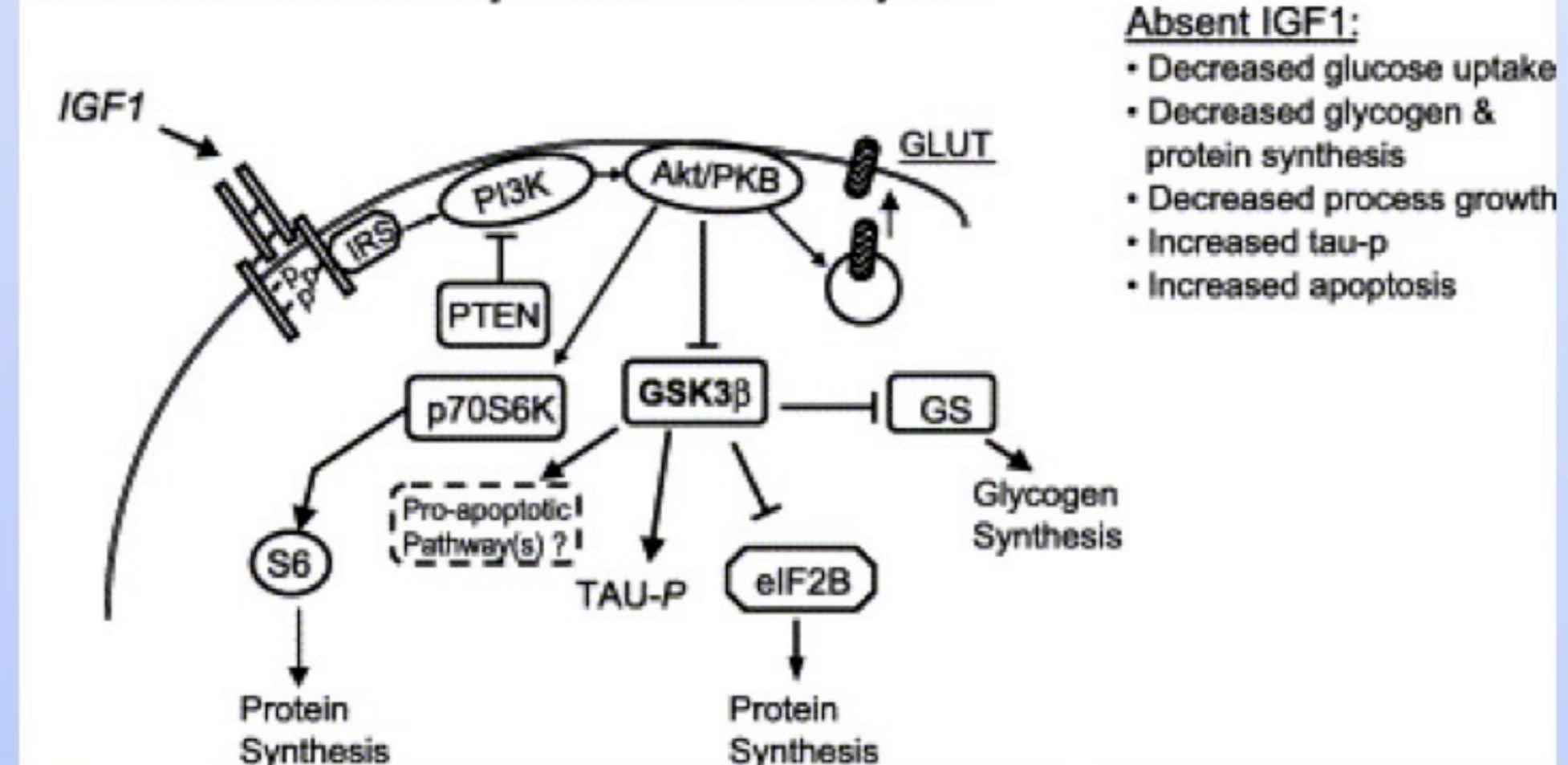
# Signalisation et croissance



# IGF1R?

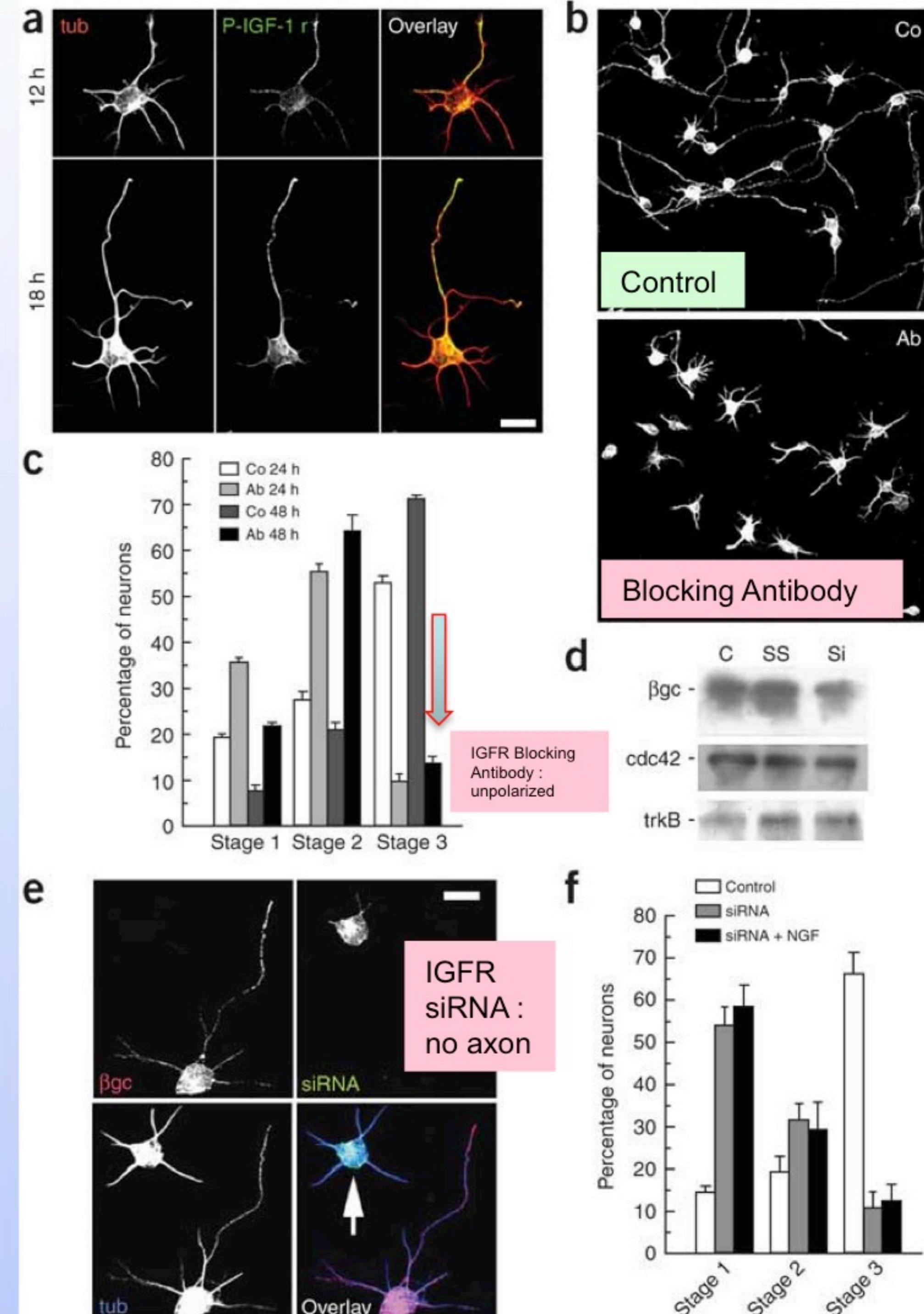


IGF1's Anabolic Pathways in the Nervous System



**IGF-1 receptor is essential for the establishment of hippocampal neuronal polarity**

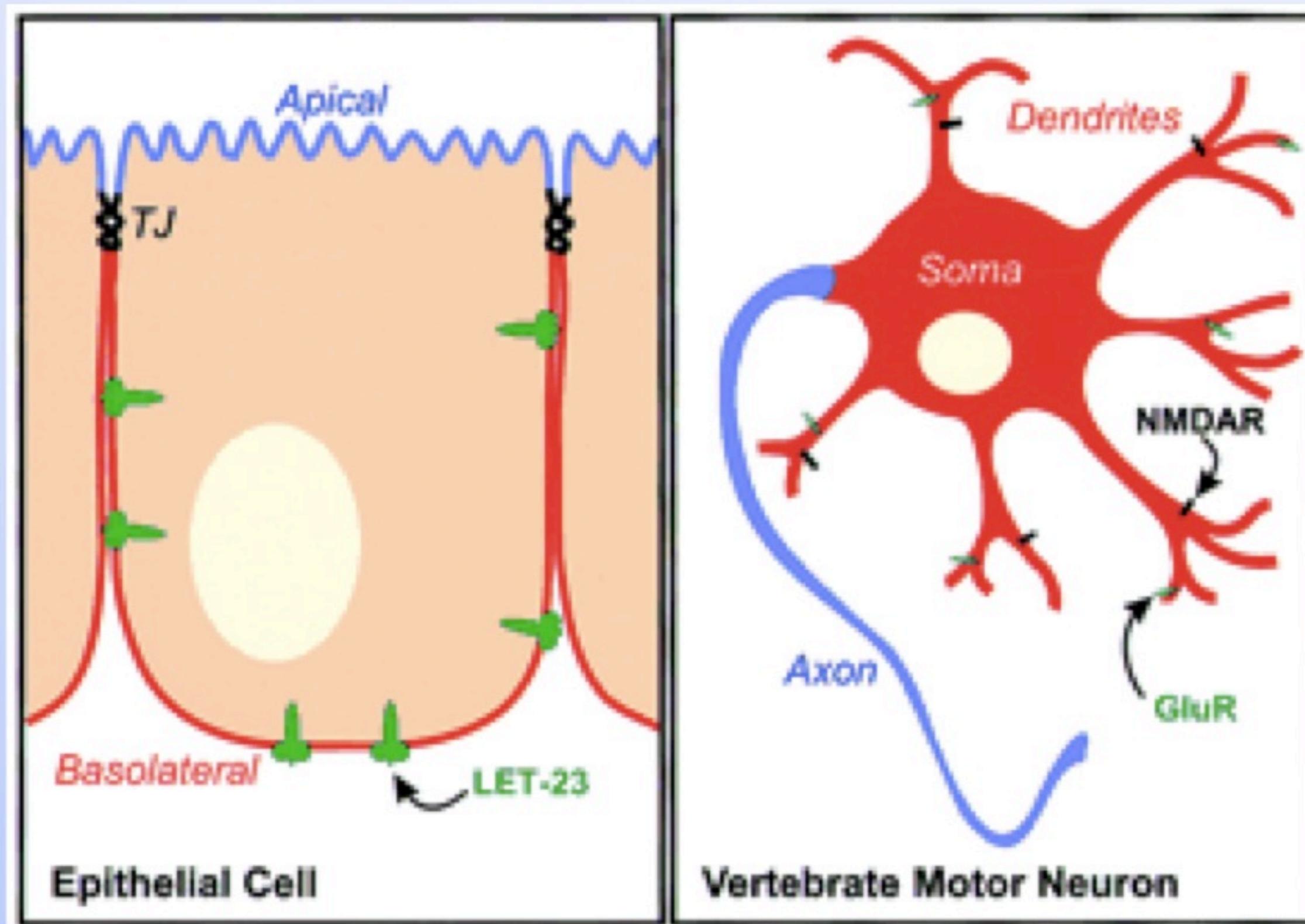
Lucas Sosa, Sebastian Dupraz, Lisandro Laurino, Flavia Bollati, Mariano Bisbal, Alfredo Cáceres, Karl H Pfenninger & Santiago Quiroga Nature Neuroscience 2006



Reducing IGF-1R function by incubation with blocking antibody or by siRNA silencing prevents axon formation in rat hippocampal neurons

# 6. La métaphore épithéliale imparfaite

APICAL~AXON



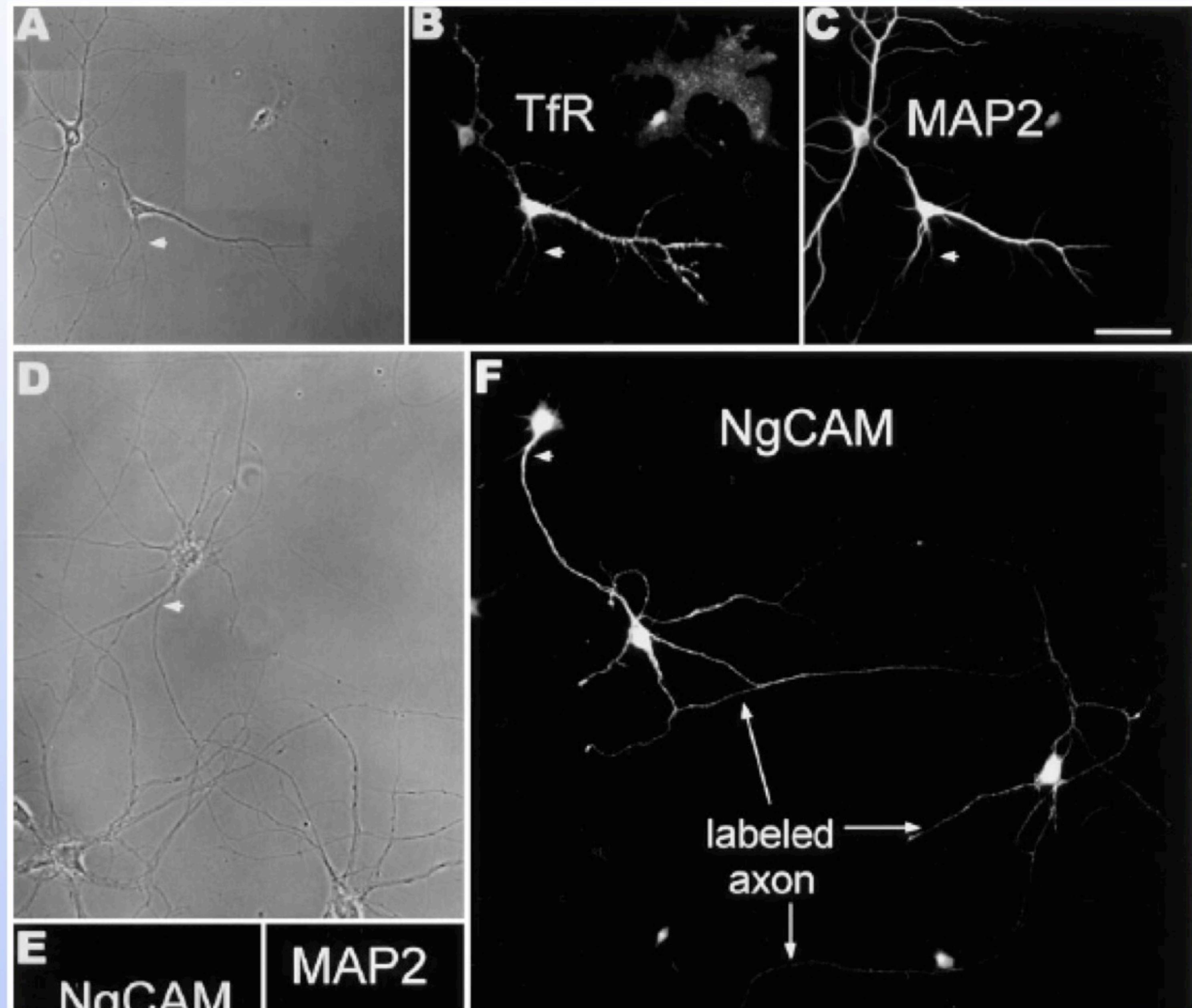
BASOLATERAL~DENDRITE

**Polarized Sorting of Viral Glycoproteins to the Axon and Dendrites of Hippocampal Neurons In Culture**  
C. G. Dotti and K. Simons Cell, Vol. 62, 63–72, July, 1990

TfR:

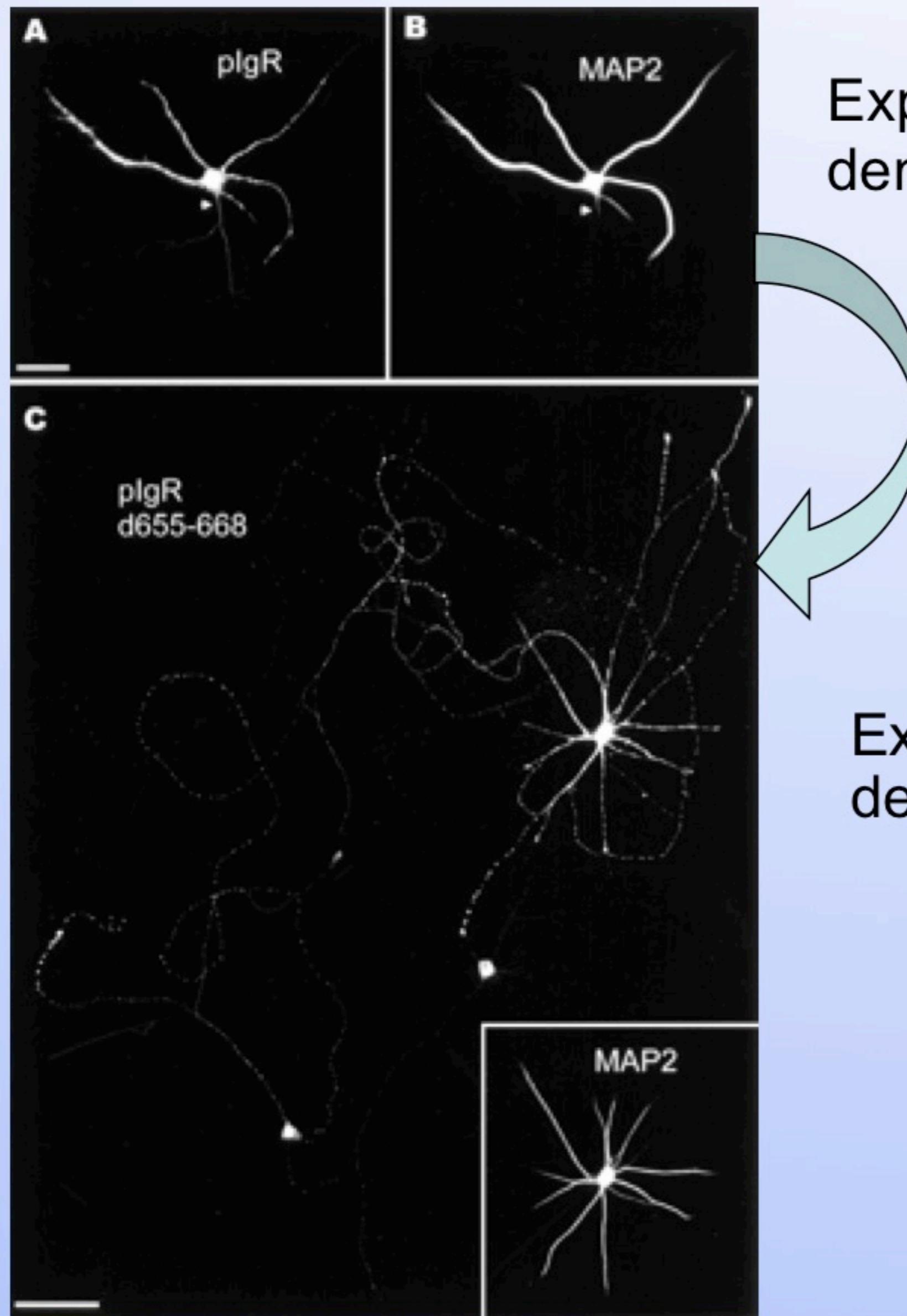
Epithelial: basolateral

Neurone: dendrite



Jareb, Mark and Bunker, Gary (1998).

The Polarized Sorting of Membrane Proteins Expressed in Cultured Hippocampal Neurons Using Viral Vectors. *Neuron* 20:855



Expression  
dendritique

Mutation du  
motif  
basolatéral

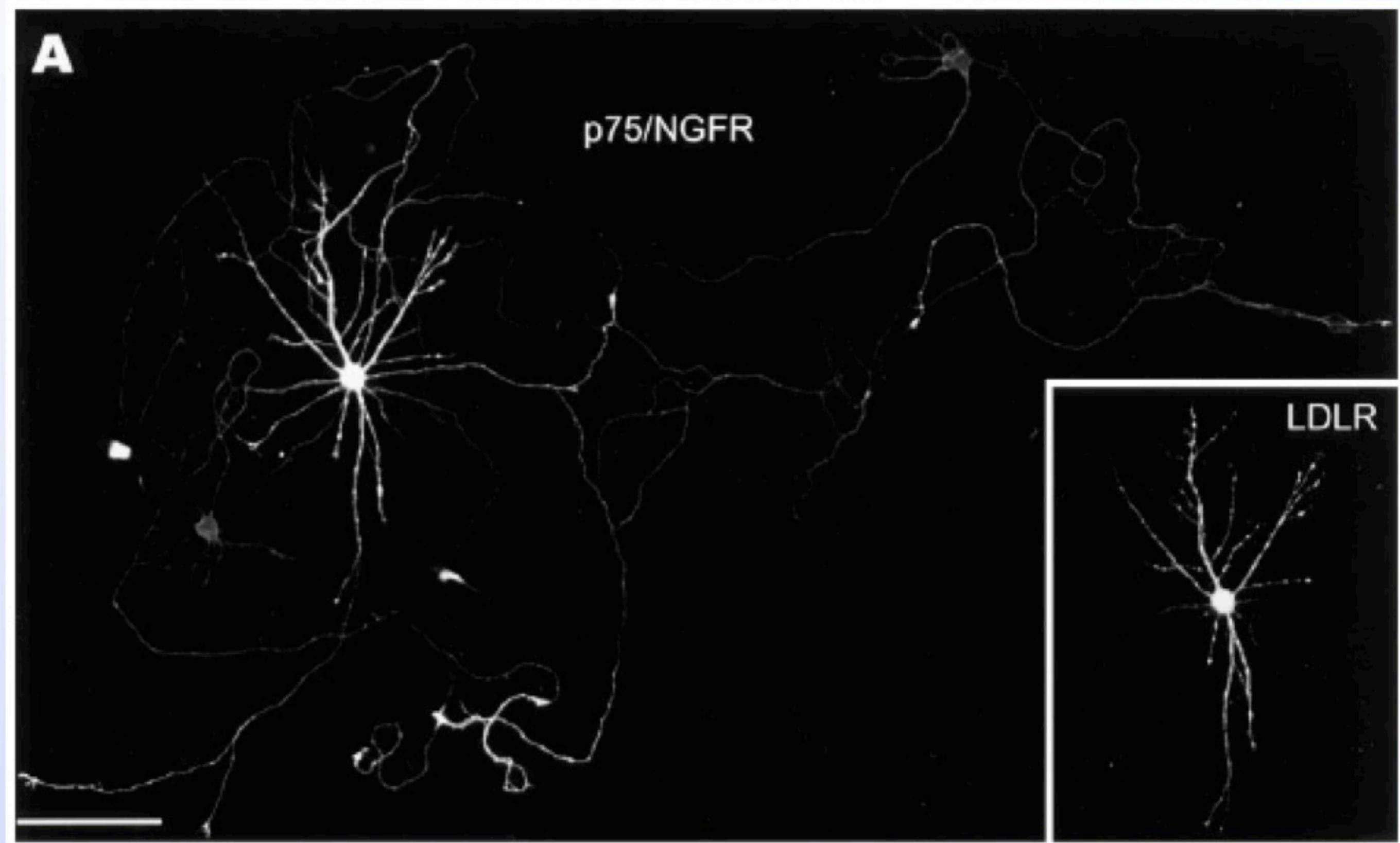
Expression  
dendritique  
+  
axonale

La mutation du motif basolatéral  
épithéliale induit une perte de  
localisation polarisée.

**Jareb, Mark and Bunker, Gary (1998).  
The Polarized Sorting of Membrane Proteins  
Expressed in Cultured Hippocampal  
Neurons Using Viral Vectors. Neuron 20:855**

Contre-exemple:

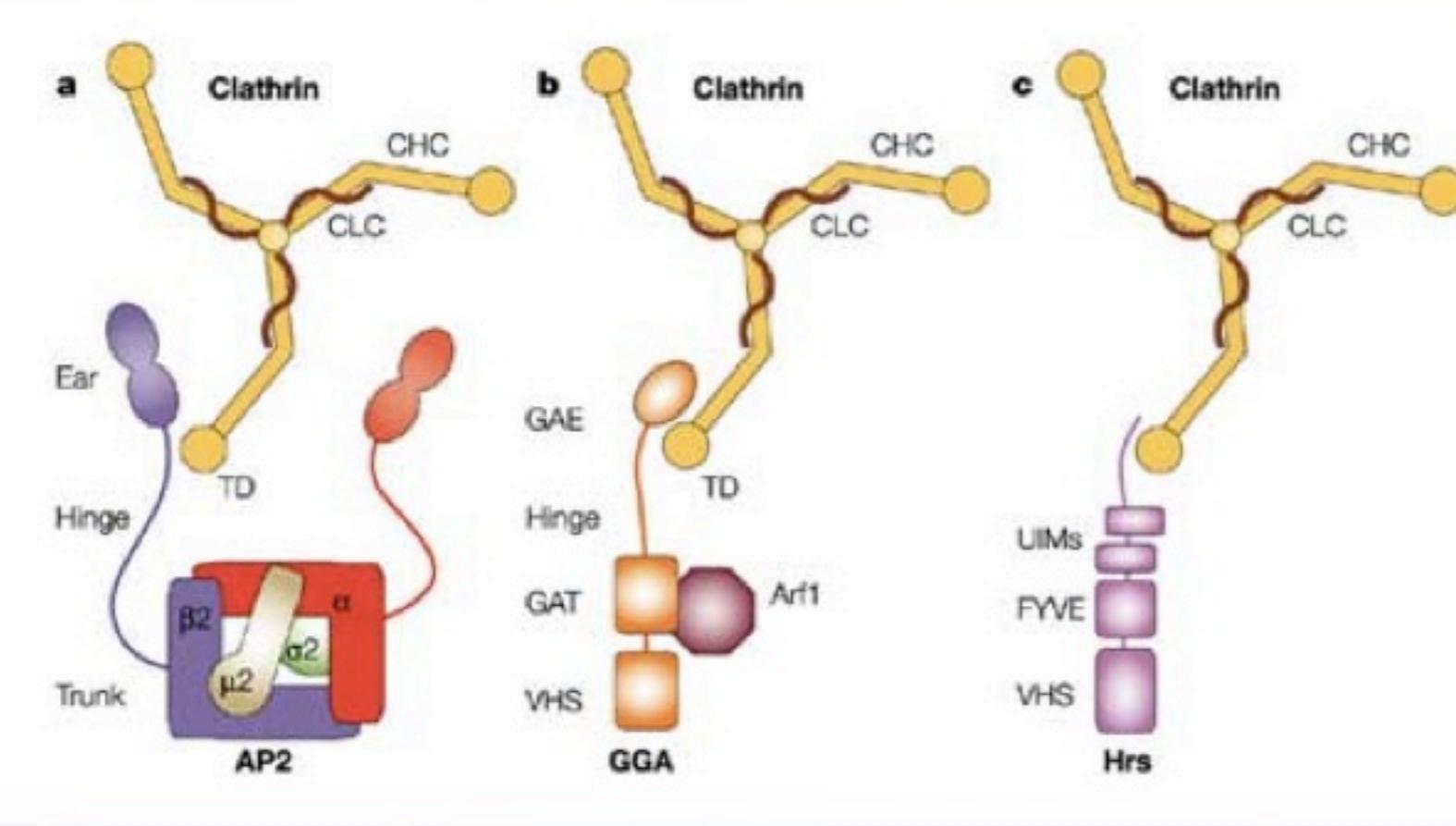
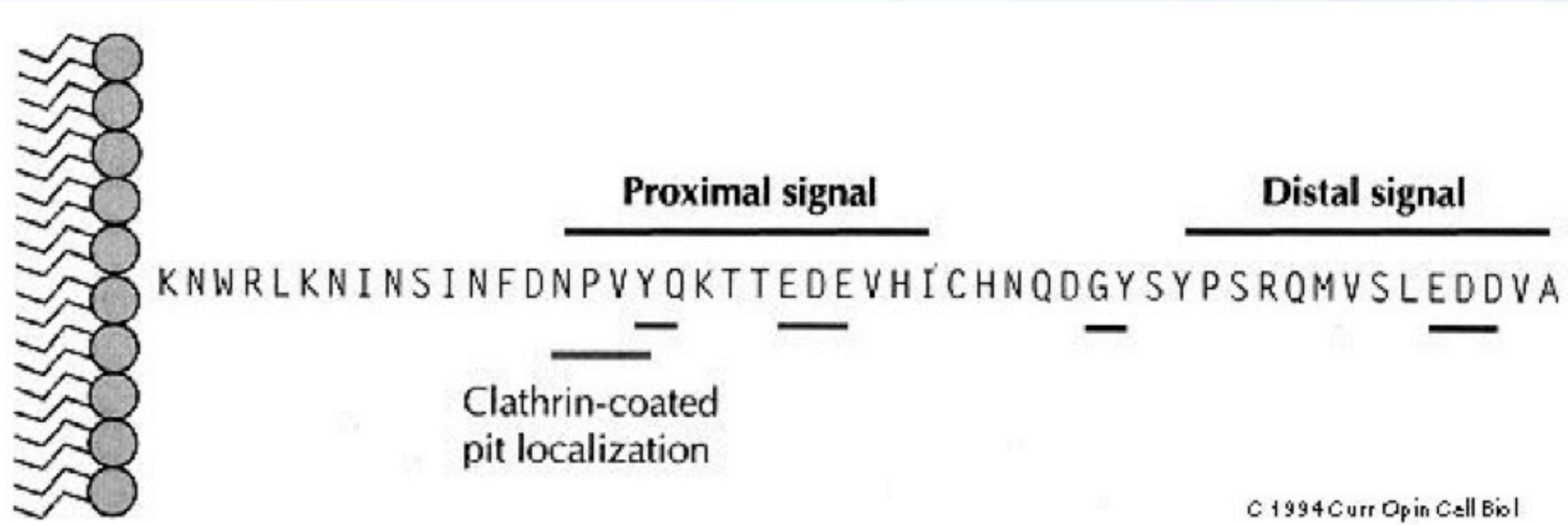
Des protéines exprimées dans le pole apical des cellules épithéliales se retrouve dans les dendrites des neurones.



**Les signaux basolatéraux semblent conservés des cellules épithéliales aux neurones,  
mais les neurones possèdent des signaux axonaux propres.**

**Jareb, Mark and Bunker, Gary (1998).  
The Polarized Sorting of Membrane Proteins  
Expressed in Cultured Hippocampal  
Neurons Using Viral Vectors. *Neuron* 20:855**

# motifs de ciblages



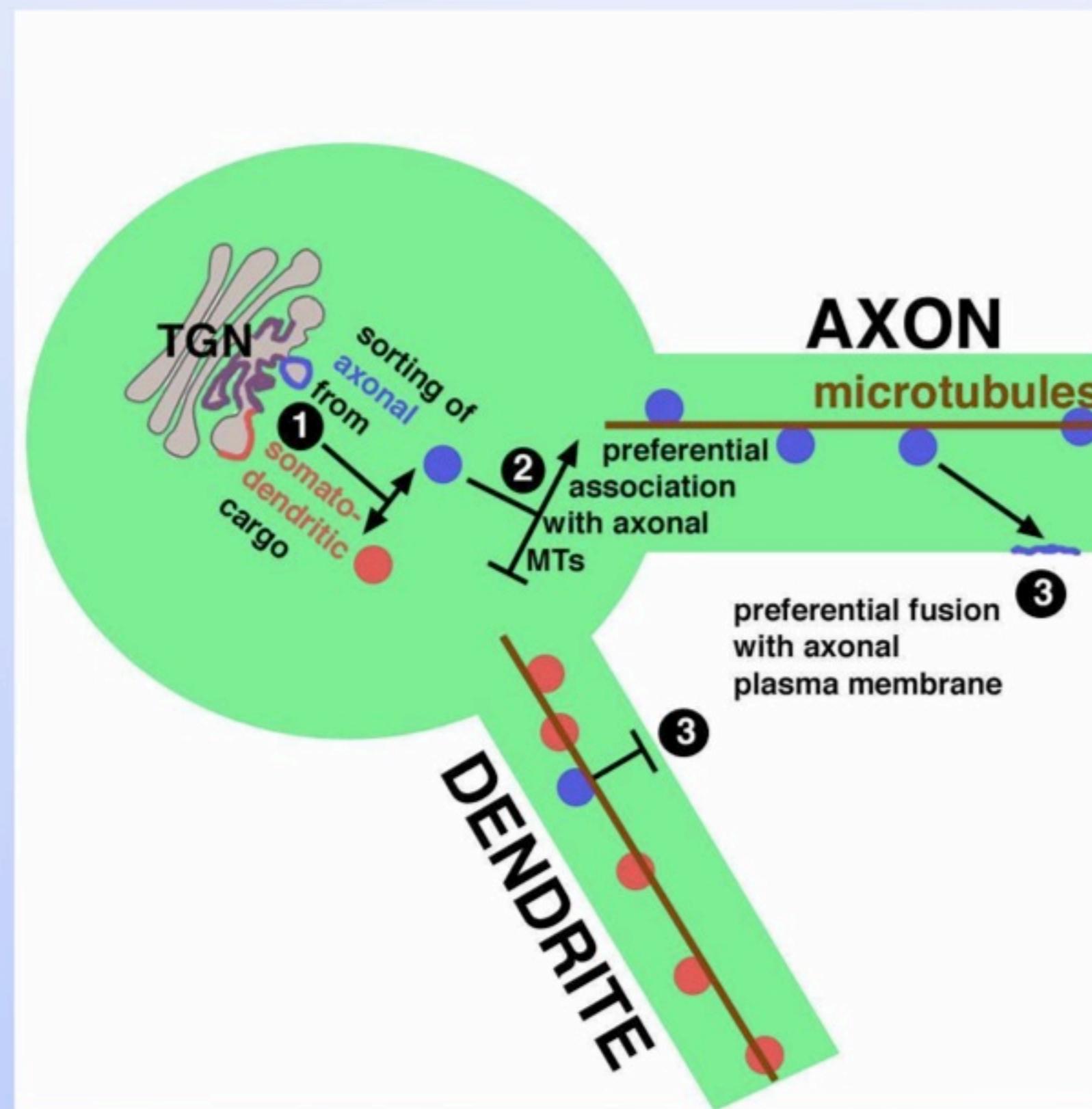
Major sorting signal and adaptor interactions	
SORTING SIGNAL	BINDS TO:
Tyr-X-X-hydrophobic (YXXØ)	$\mu$ subunits of adaptor complexes
Asn-Pro-X-Tyr (NPXY)	Other accessory proteins (ARH, Dab2)
[Asp/Glu]-X-X-X-Leu-[Leu/Ile] (Dileucine)	GGA, Others?
Monoubiquitin	Other accessory proteins (Eps 15)

www.ergito.com

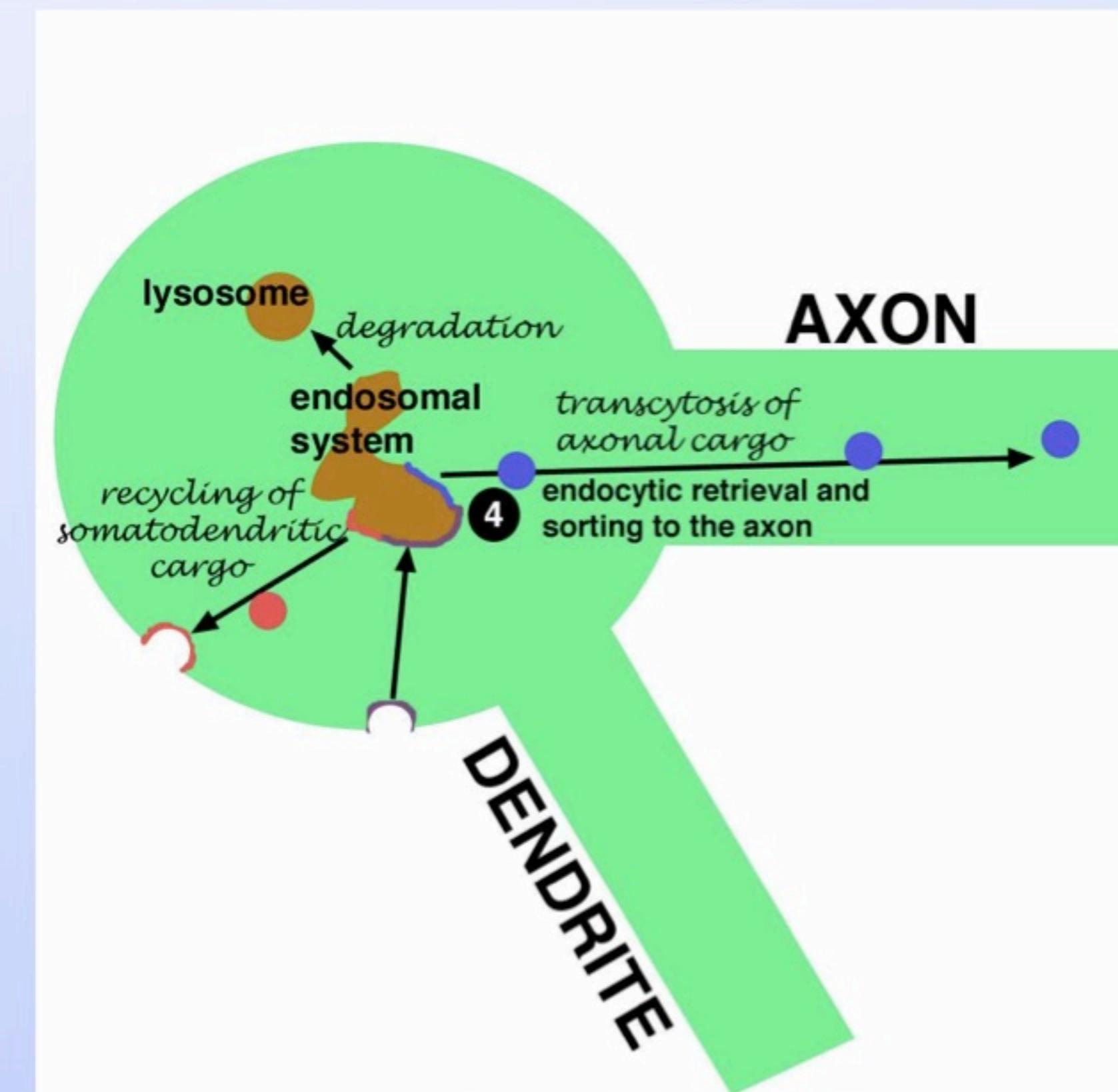
Type	Location	Representative examples
<b>Basolateral sorting signals</b> YXX <sup>-</sup>	Cytoplasmic	Influenza virus hemagglutinin (Tyr mutant) Vesicular stomatitis virus glycoprotein
	Cytoplasmic	FcRII-B2 receptor
	Cytoplasmic	Transferrin receptor LDL receptor distal domain Polymeric immunoglobulin receptor
<b>Apical sorting signals</b> GPI-anchor	Membrane, luminal leaflet	Placental alkaline phosphatase Thy-1
	Luminal	gp-80 Erythropoietin
	Membrane	Influenza virus neuraminidase Influenza virus hemagglutinin
<b>Late endosomes/lysosomes</b> Mannose-6-phosphate	Luminal	Cathepsin B Cathepsin D
	Cytoplasmic	Lamp-1, Lamp-2 CD63
	Cytoplasmic	Invariant chain (Ii)
<b>MHC class II compartments</b> LI, ML	Cytoplasmic	Chromogranin B Pro-opiomelanocortin
	Loop stabilized by one disulfide bridge	
<b>Regulated secretory vesicles</b> Conformation-dependent motif	Luminal	
	Selective aggregation	

Sequences are given in the single letter amino acid code; <sup>-</sup> is a bulky hydrophobic amino acid.

# Transport sélectif ou rétention ?

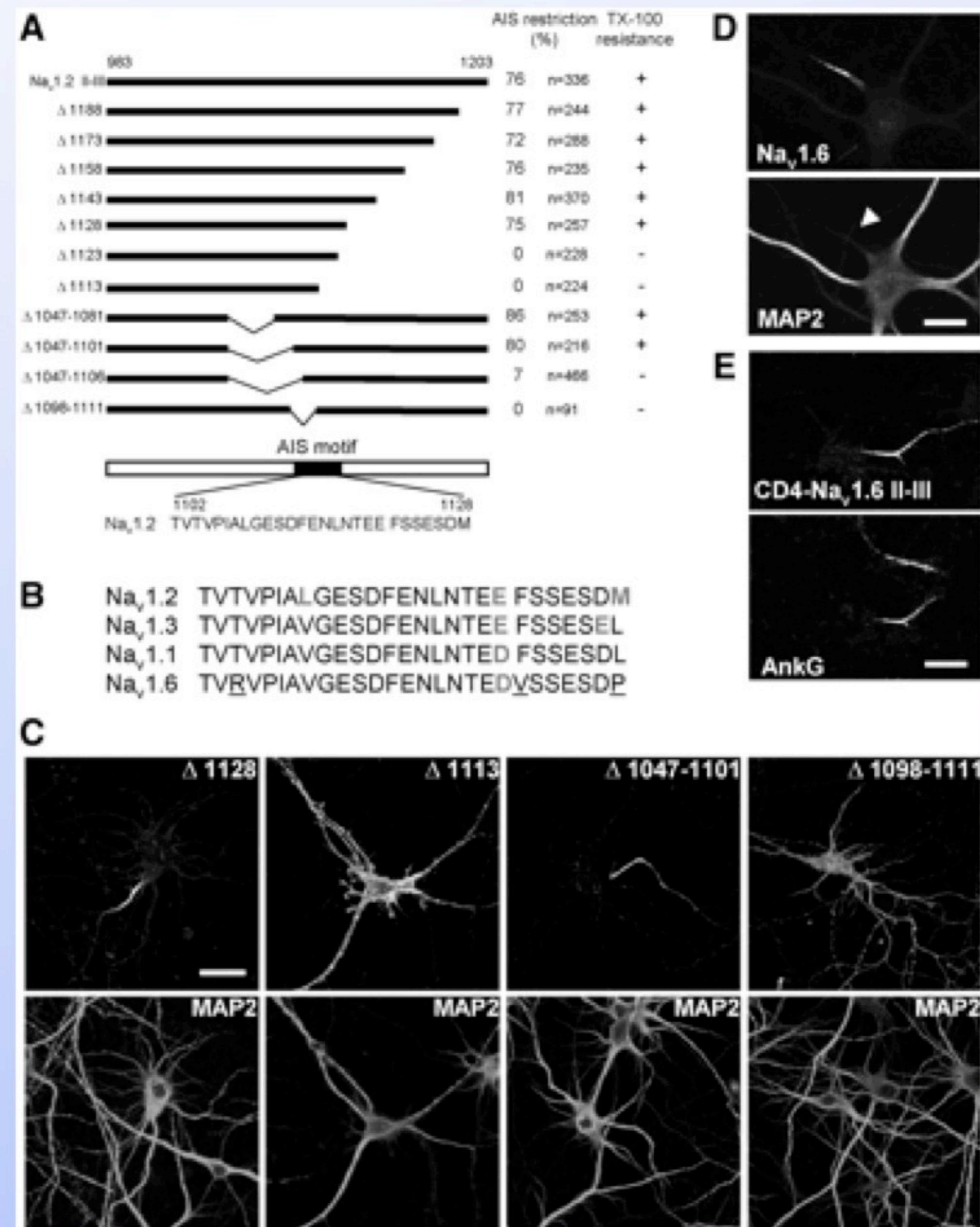
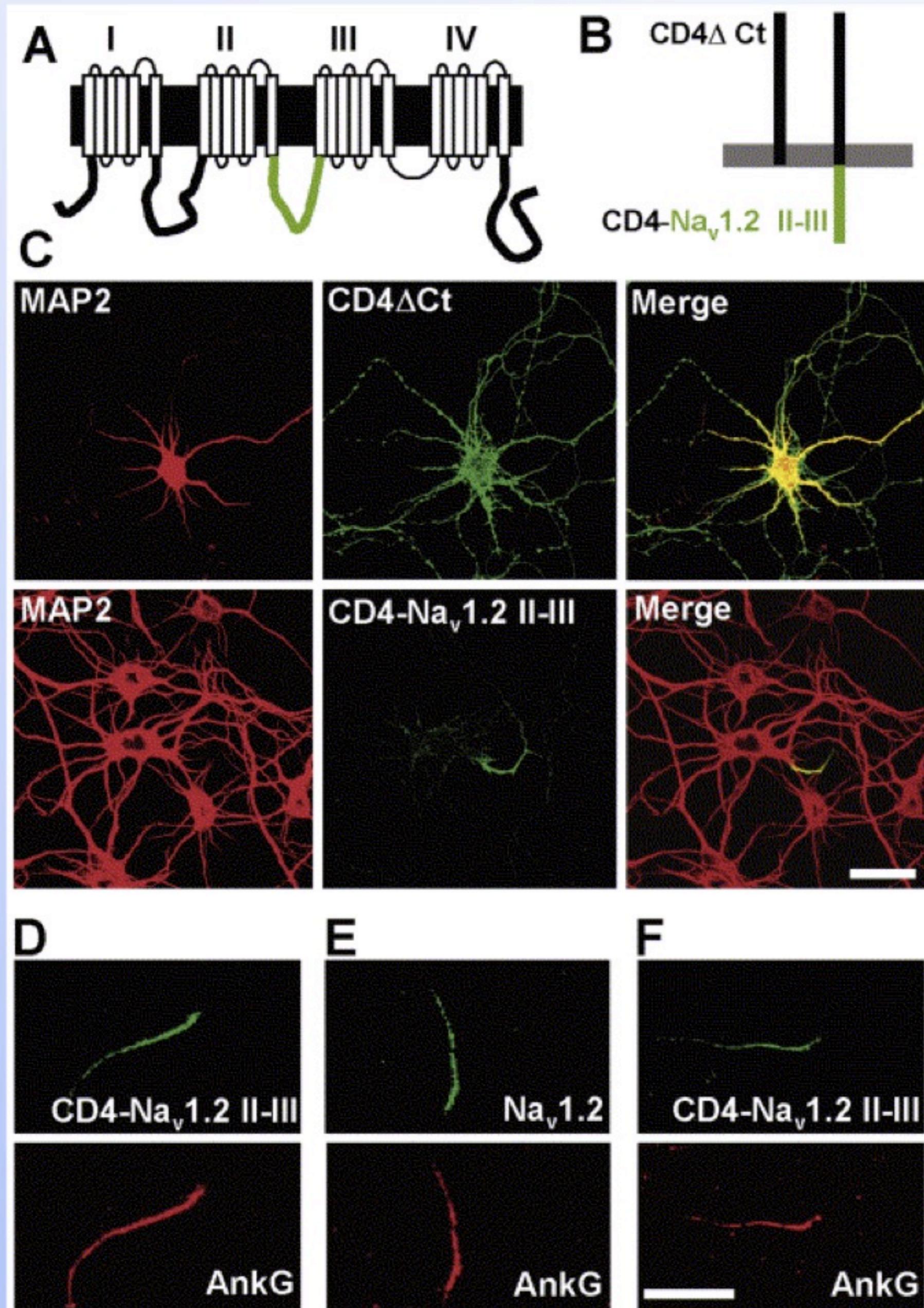


RTf

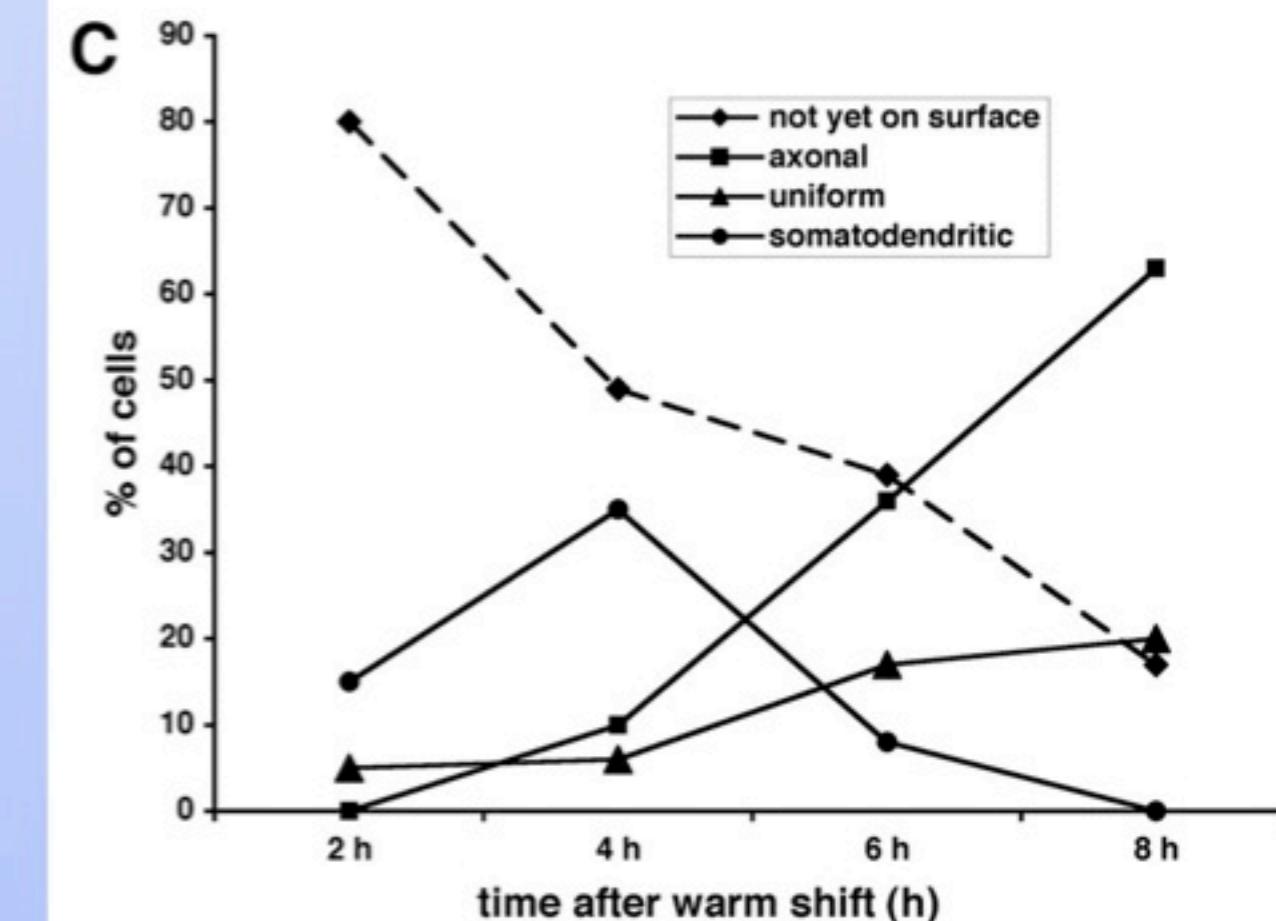
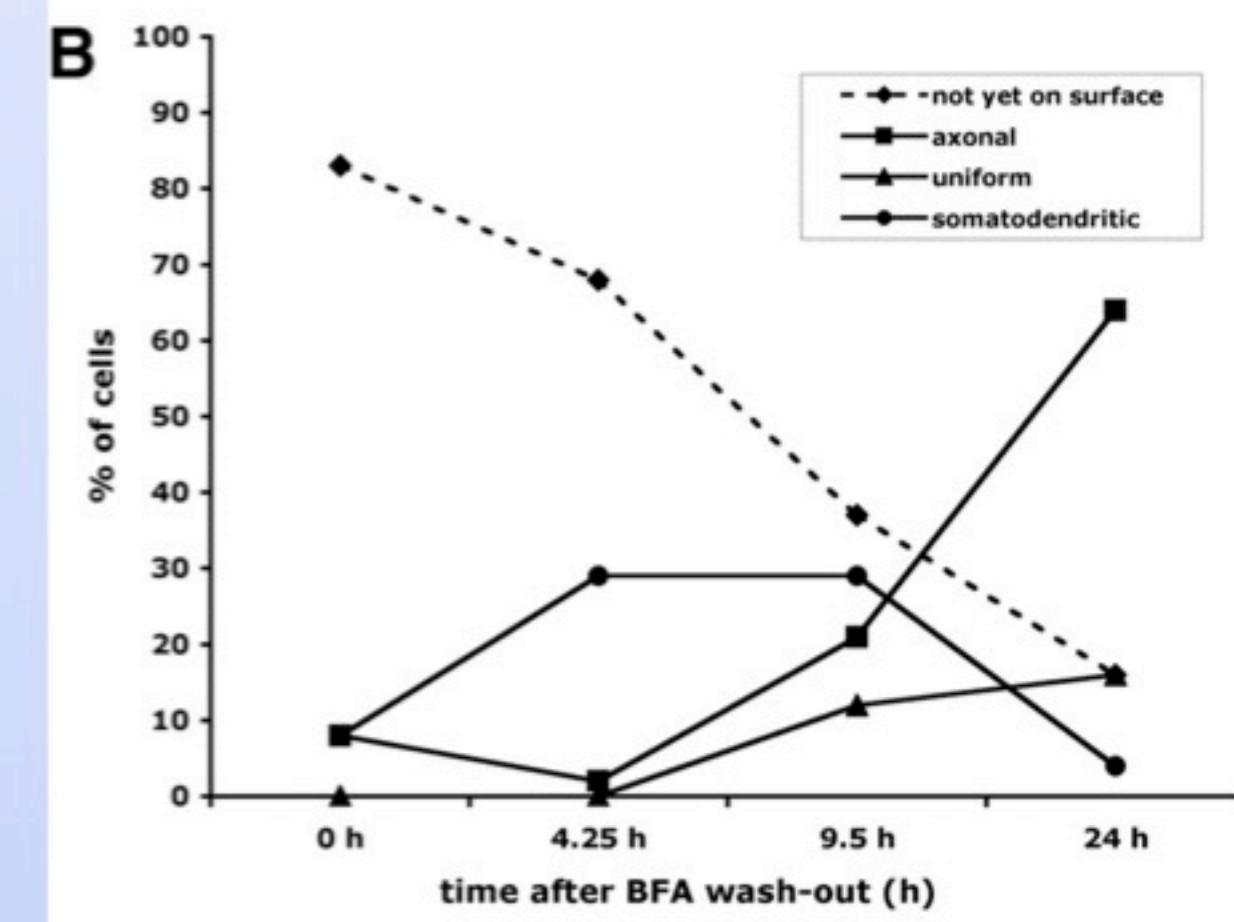
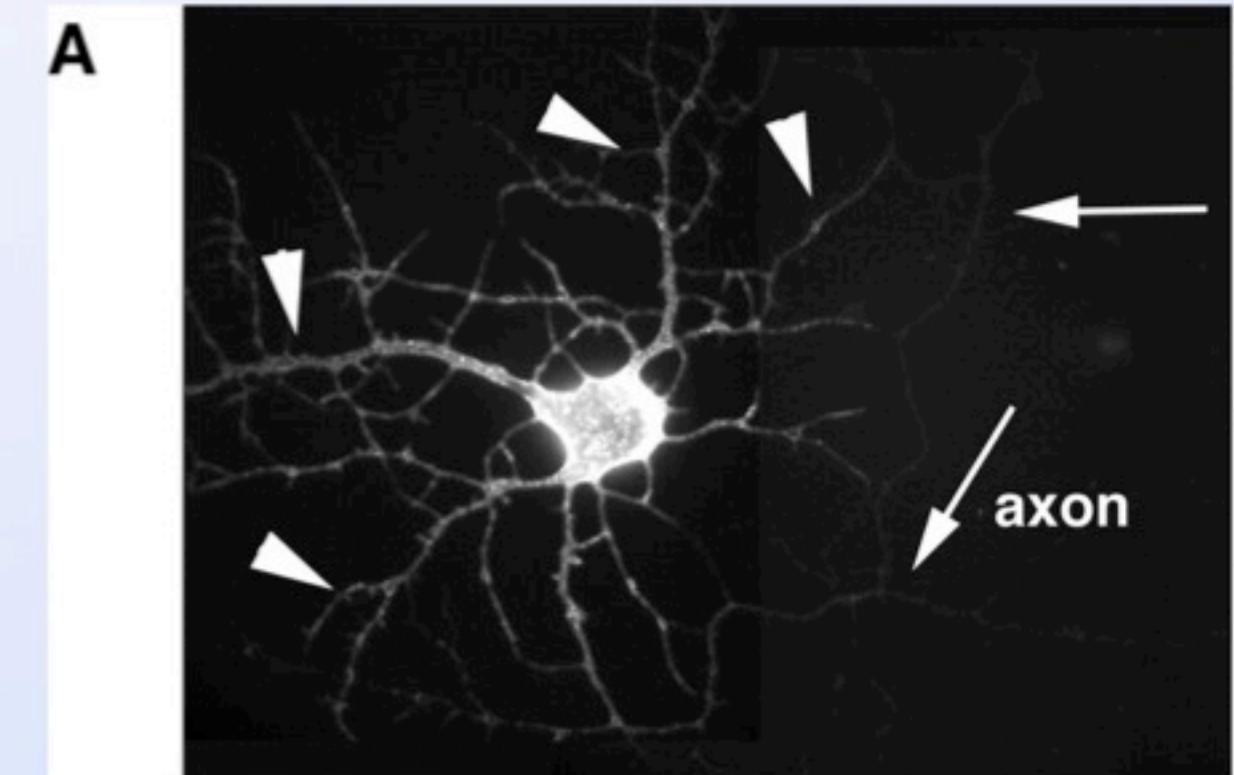
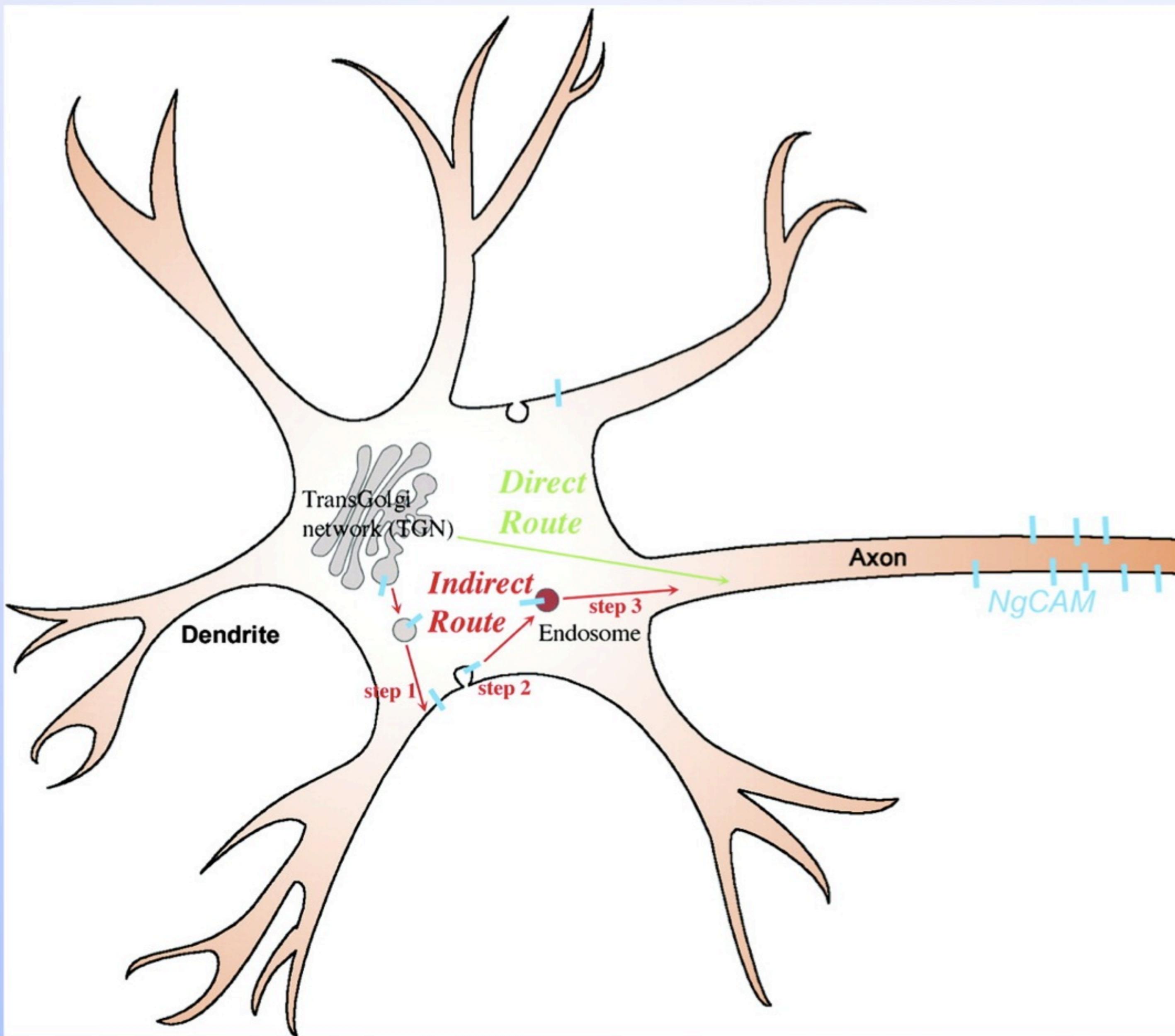


**NgCAM dendritique  
puis NgCAM axonal**

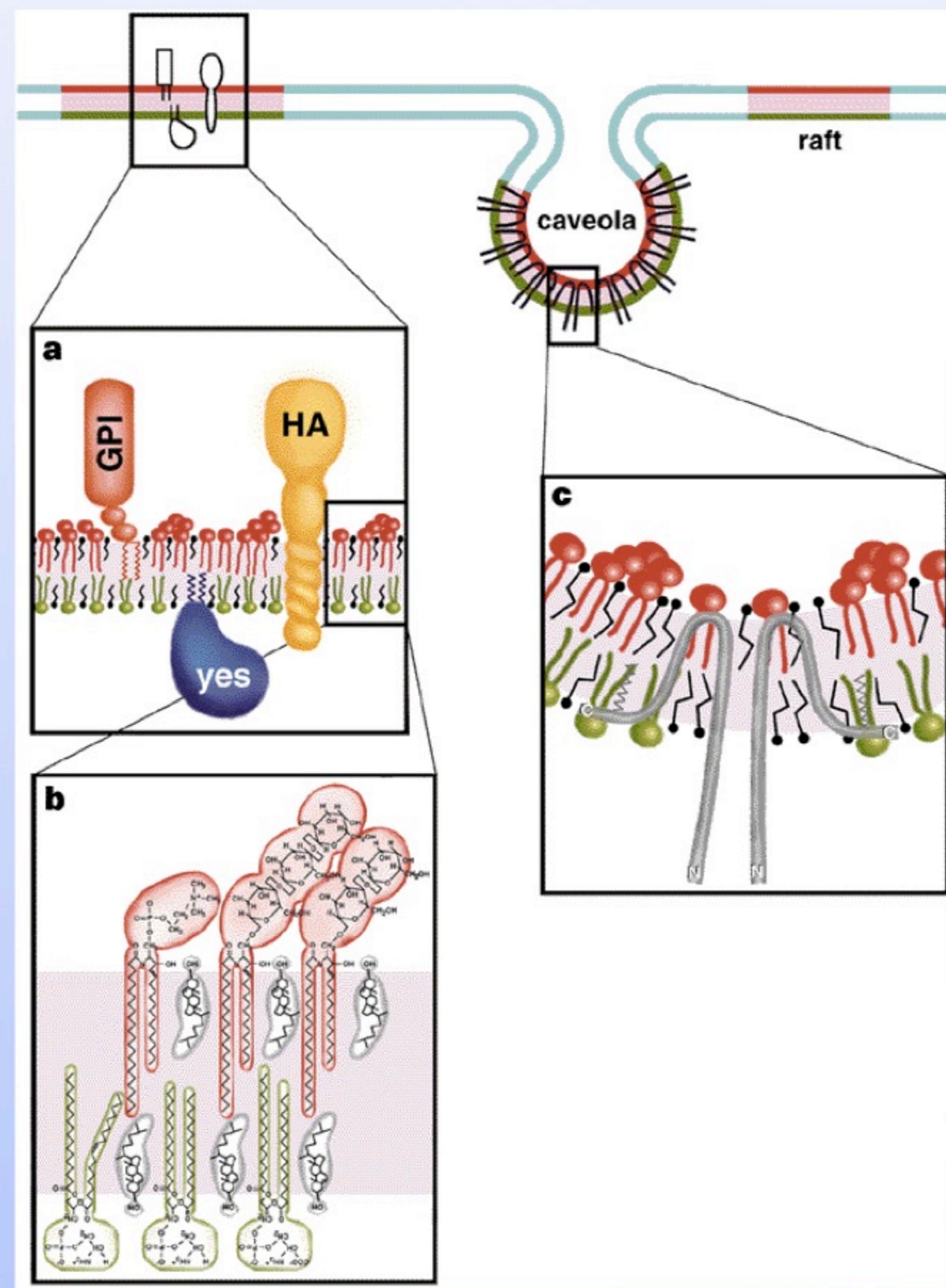
# Example de signal: pour rejoindre le segment initial de l’axone (AIS) par l’équipe de B. Dargent & coll.



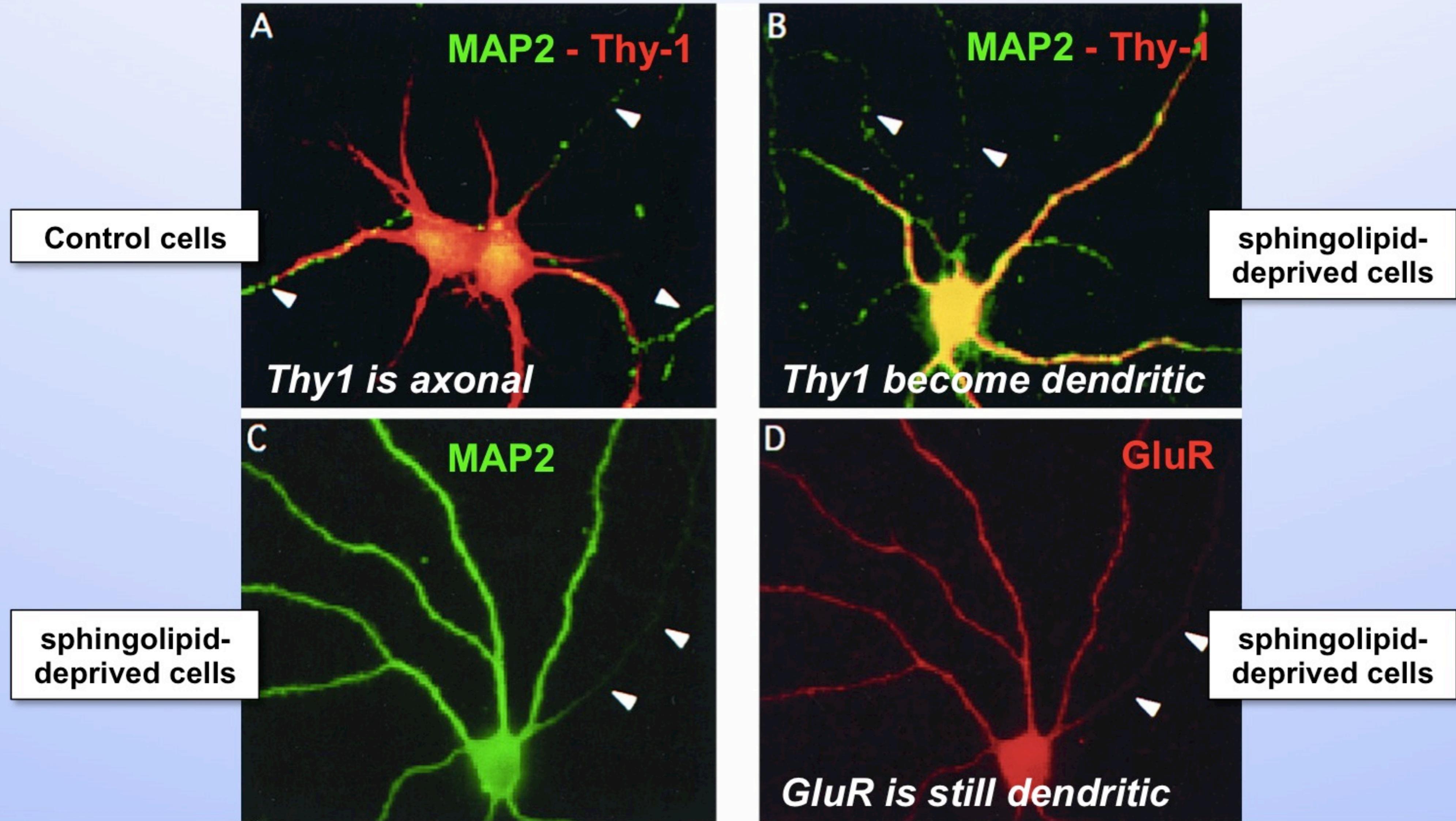
# Transcytosis of NgCAM in neurons



# Raft: axonal signals?

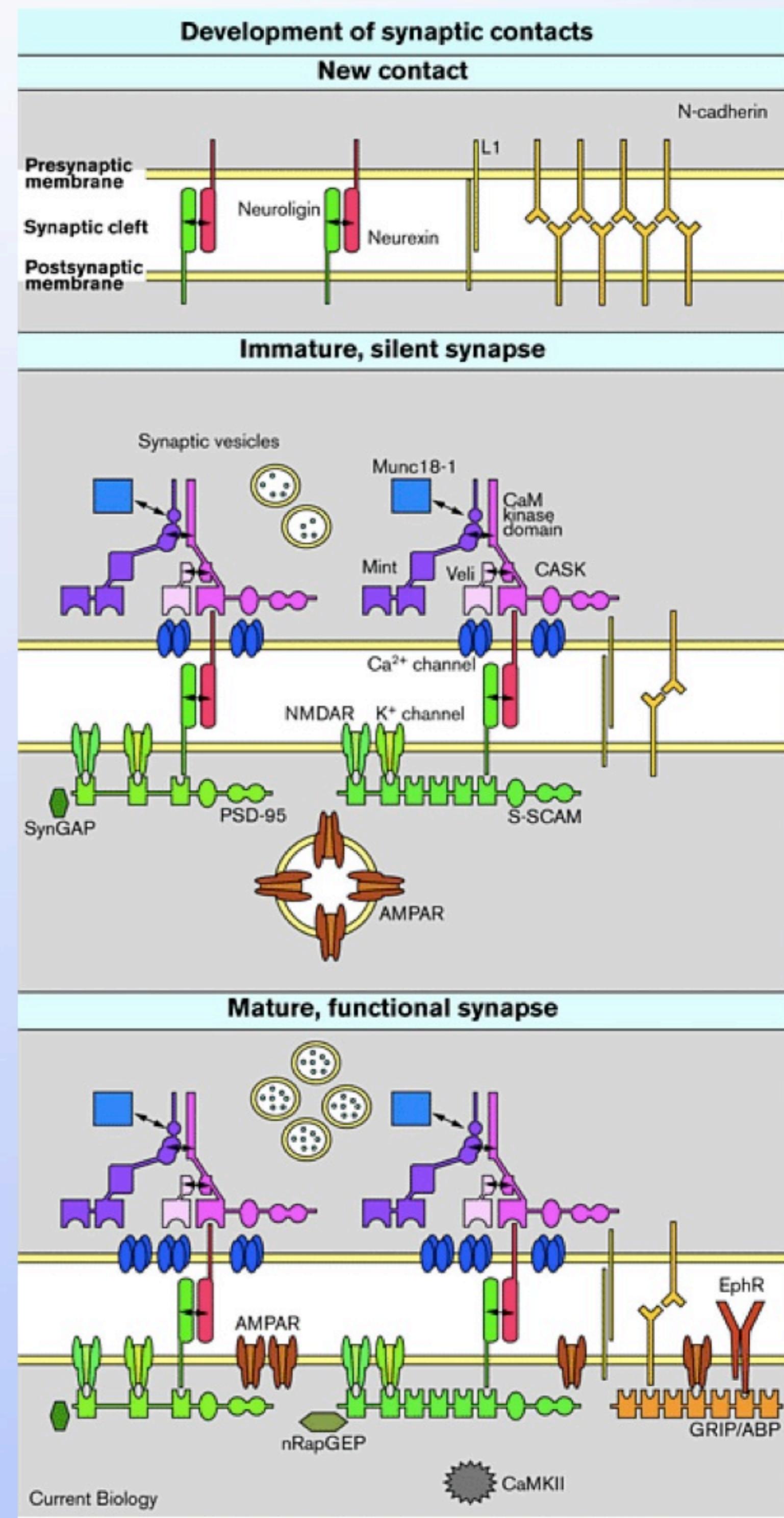


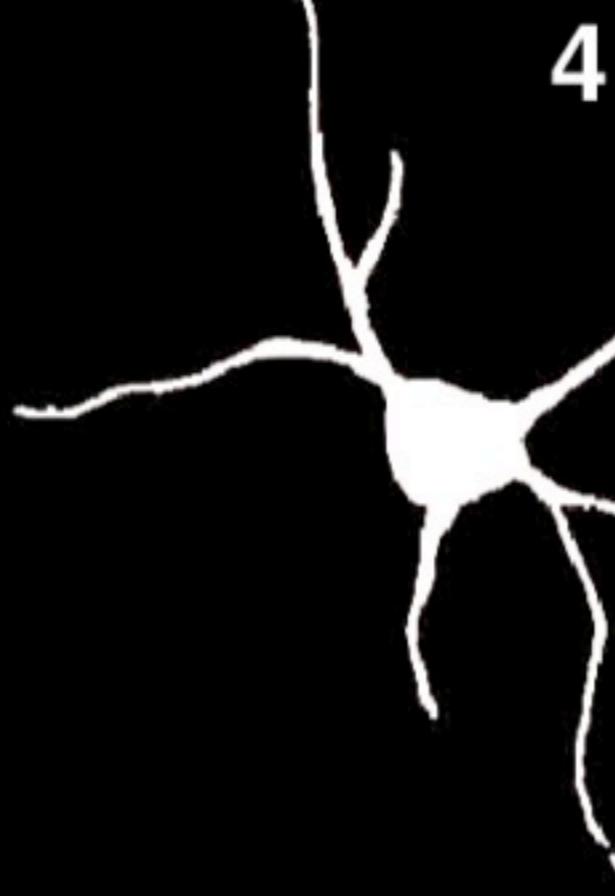
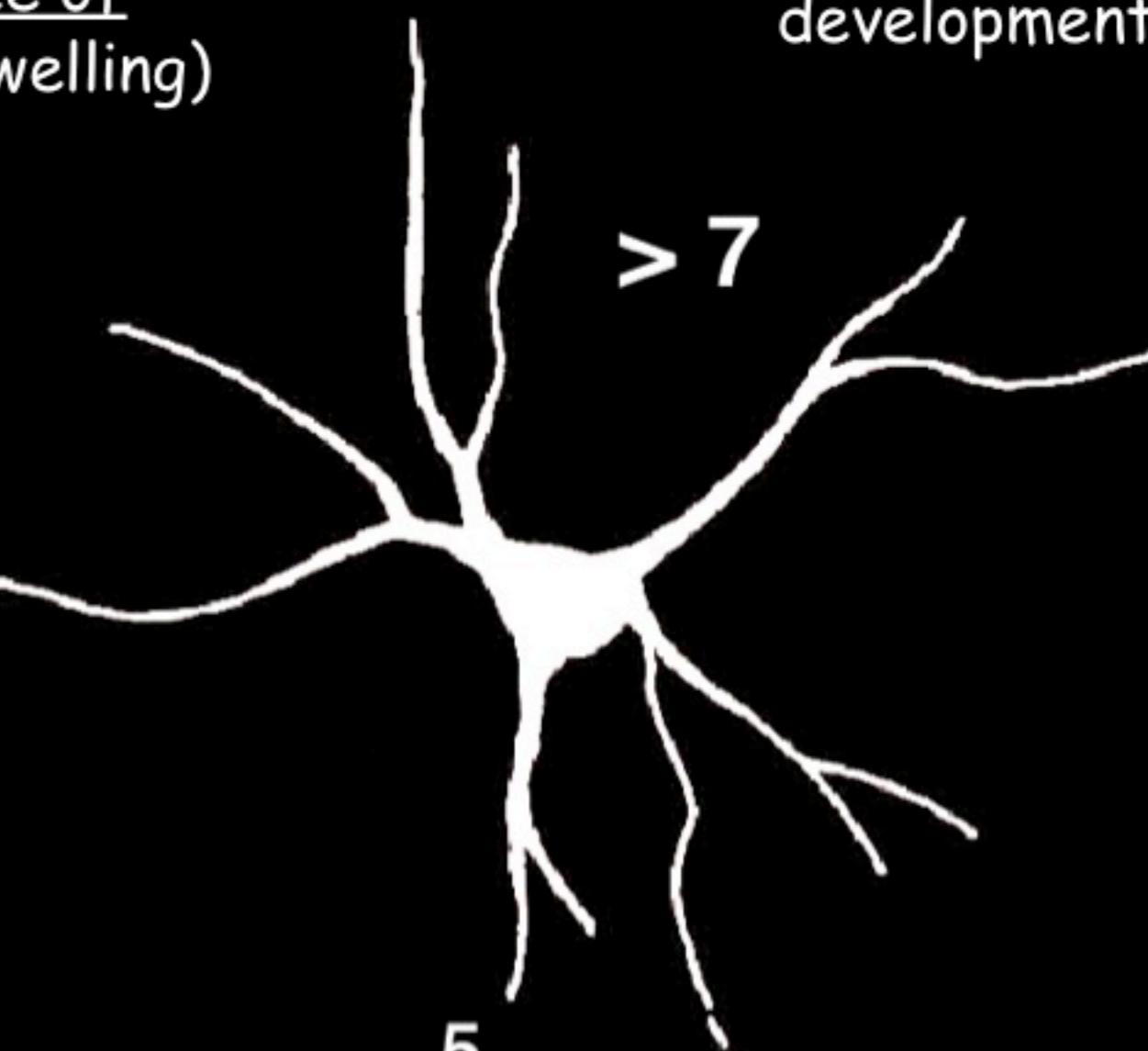
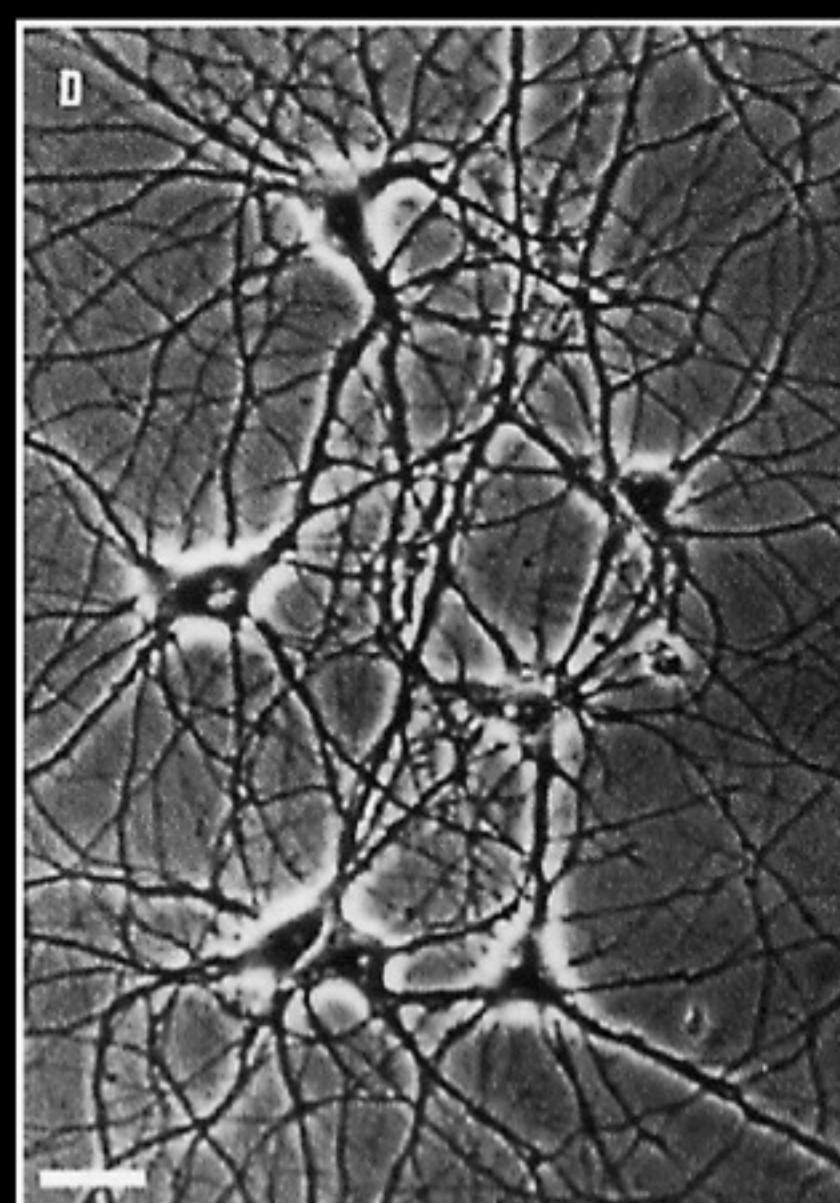
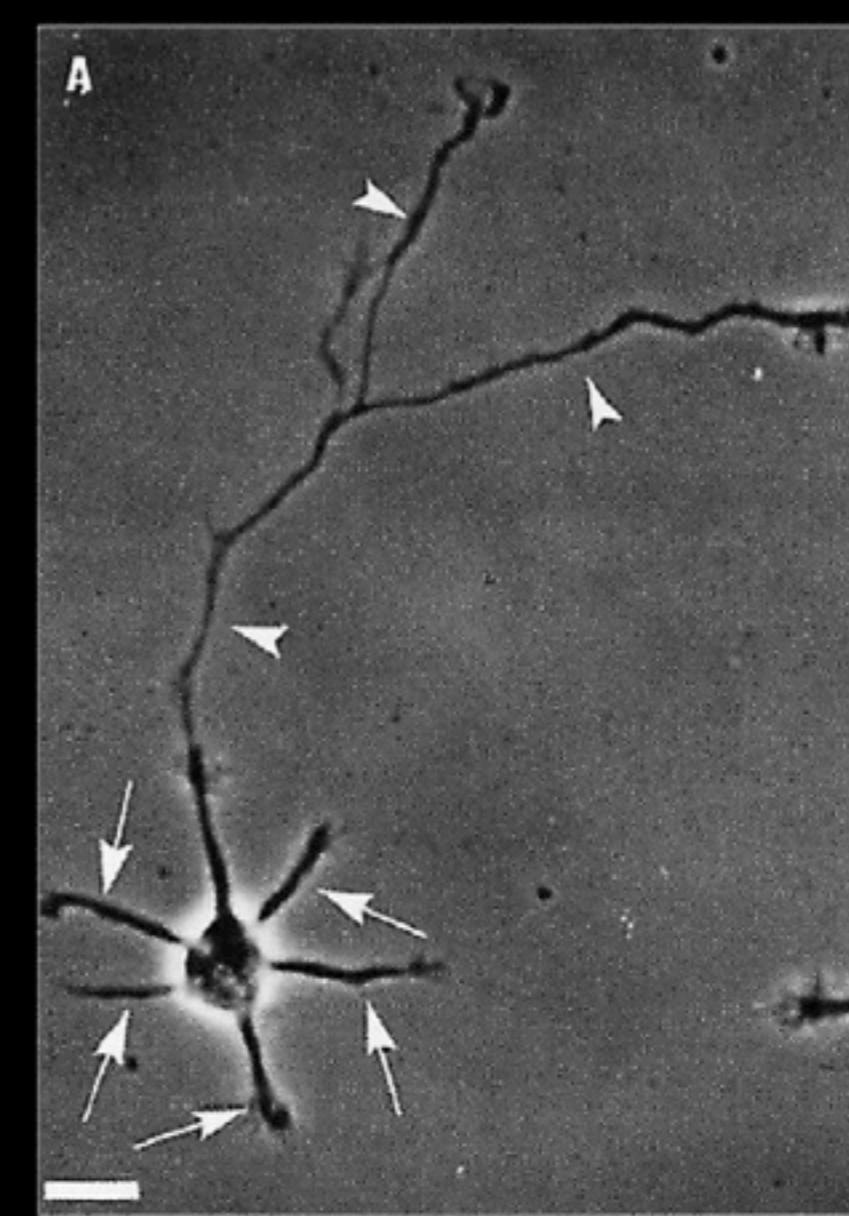
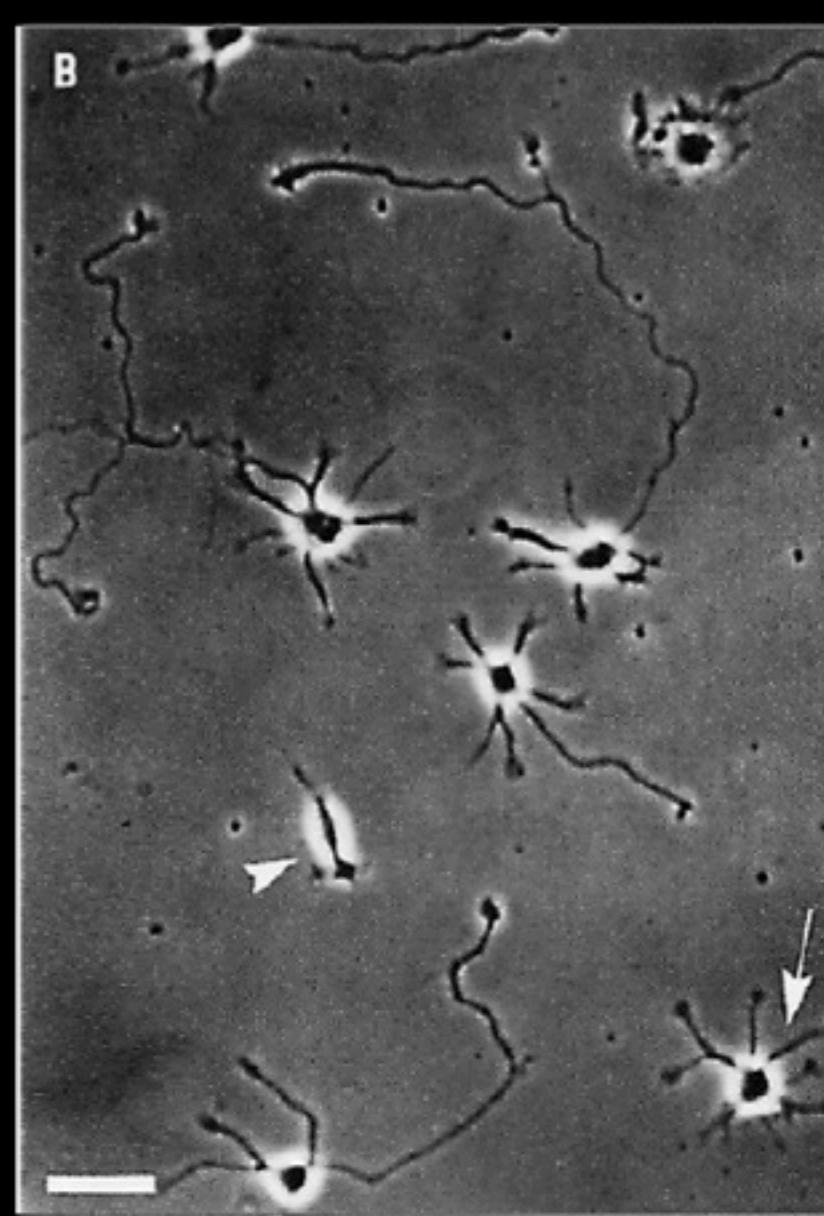
Misorting of the axonal Thy-1 but not of a dendritic membrane protein occurred in sphingolipid-deprived cells. These results indicate that neurons sort a subset of axolemmal proteins by a mechanism that requires the formation of protein-lipid rafts. The involvement of rafts in axonal membrane sorting may explain the neurological deficits observed in patients with certain types of Niemann-Pick disease.



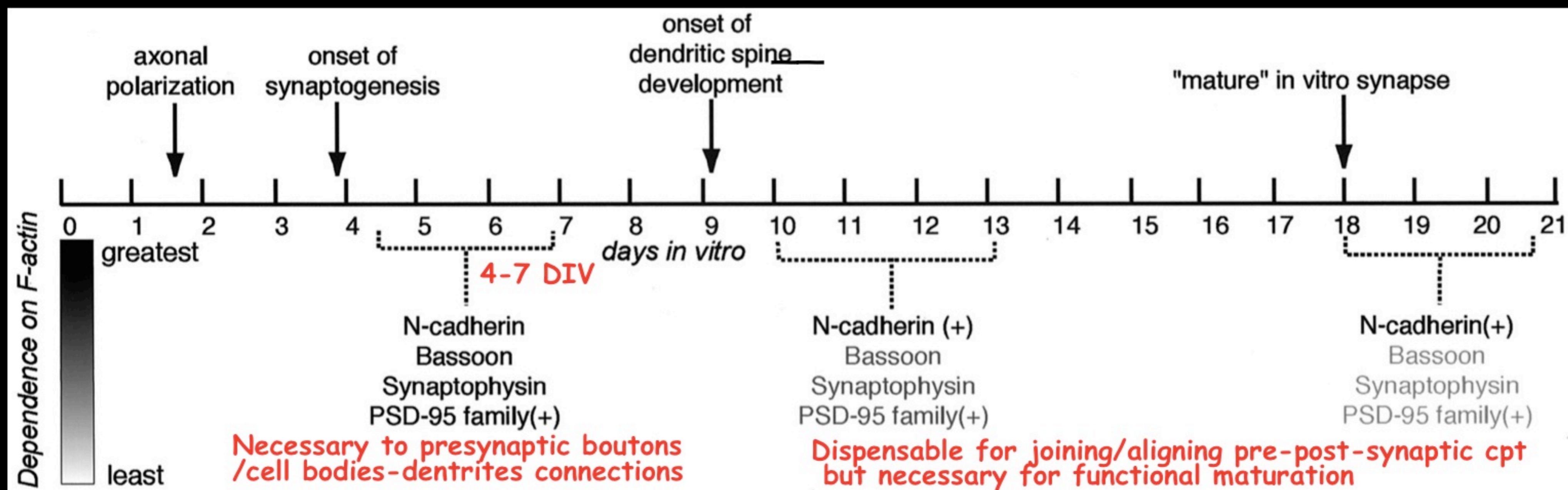
Ledesma, Maria Dolores et al. (1998) Proc. Natl. Acad. Sci. USA 95, 3966-3971

# 8. Formation of the synapse



**Jours en culture :****0,25****0,5****1,5**Axonal outgrowth  
and polarisation**DIV 4**Dendrites outgrowthSynaptogenesis with appearance of  
axonal pre-synaptic boutons (swelling)  
connecting dendrites/bodies**4****> 7**Onset of dendritic spines  
development**Stades :****1**  
Lamelipodes**2**  
Neurites mineurs**3**  
Croissance axonale**4**  
Croissance dendritique**5**  
Maturation**DIV 18-21**

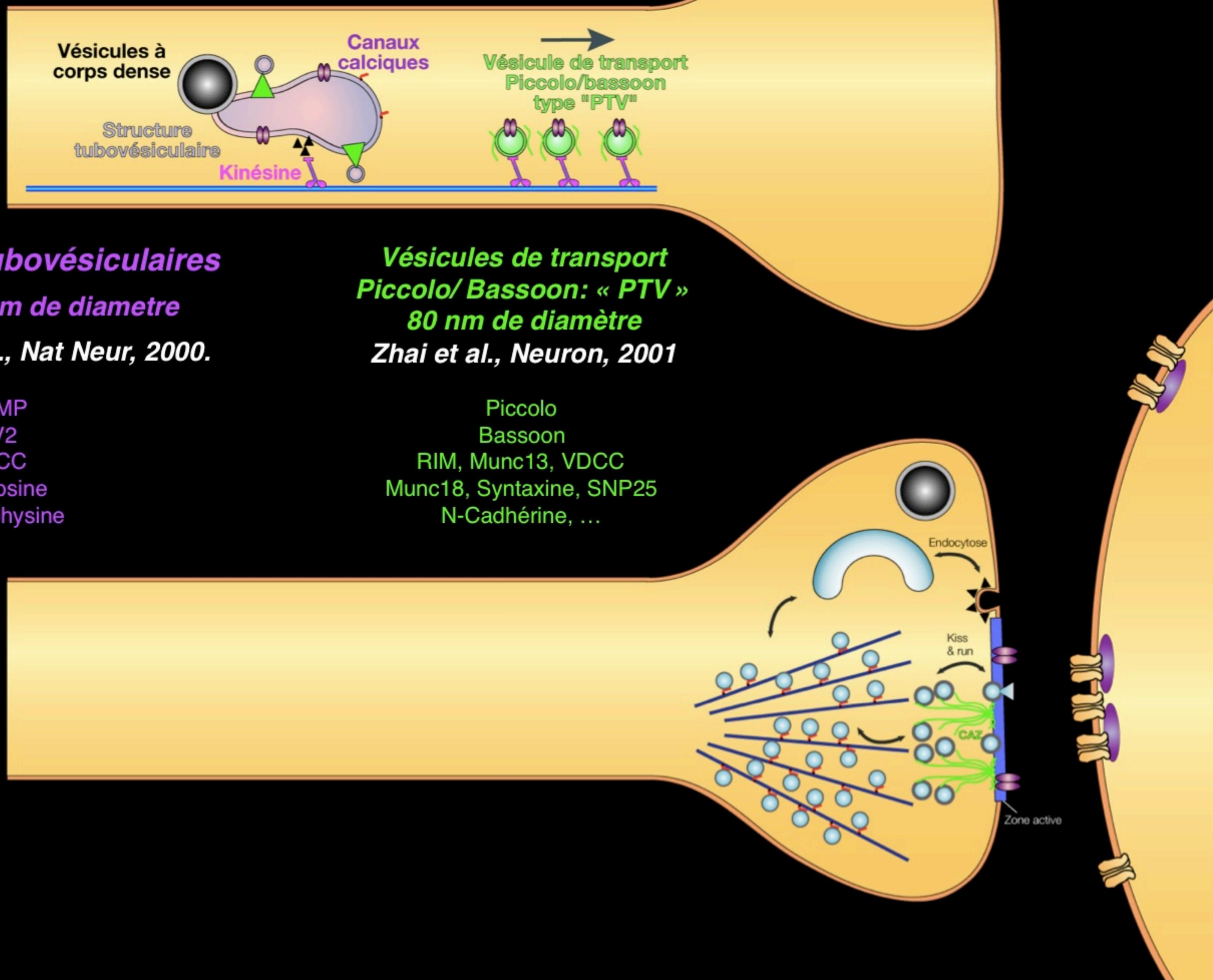
## Stages in the development of hippocampal neurons in culture (DIV = Days In Vitro)



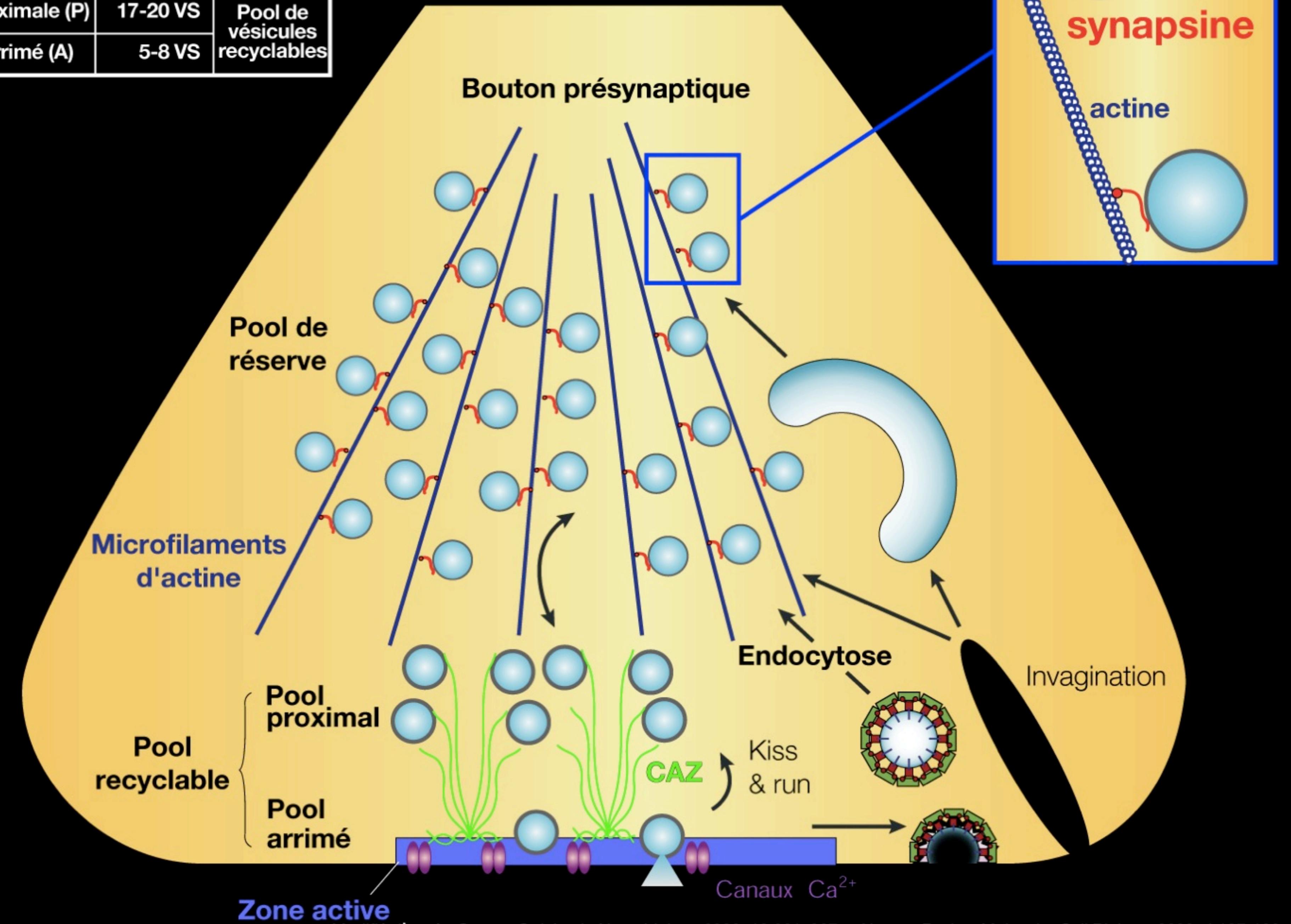
a role in directing pre- and postsynaptic specializations ?

a role in dendritic spine morphology ?

# Transport axonal des protéines présynaptiques



Type de pool	Nombre de vésicules	
de réserve (R)	180 VS	
proximale (P)	17-20 VS	Pool de vésicules recyclables
arrimé (A)	5-8 VS	



7 DIV

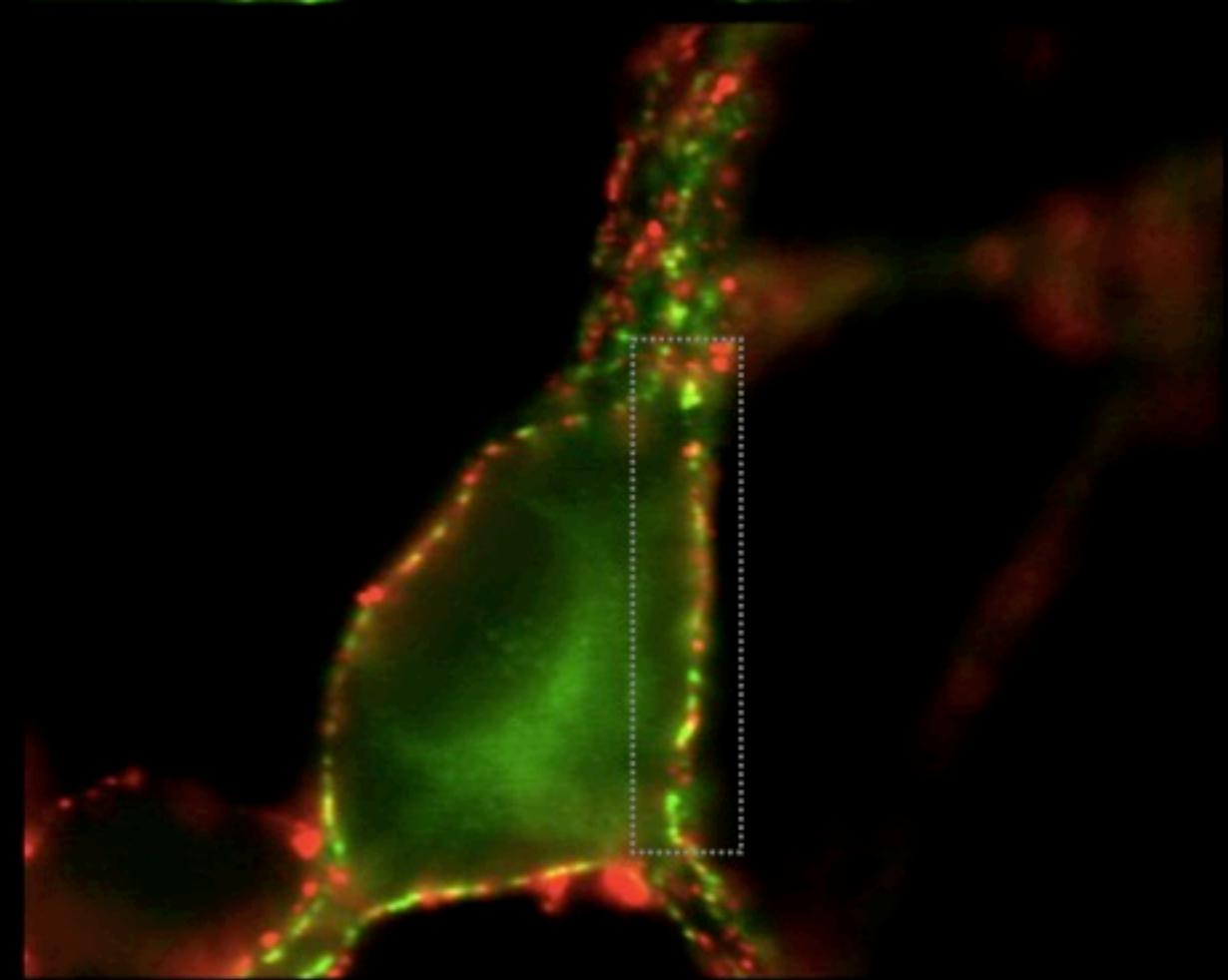
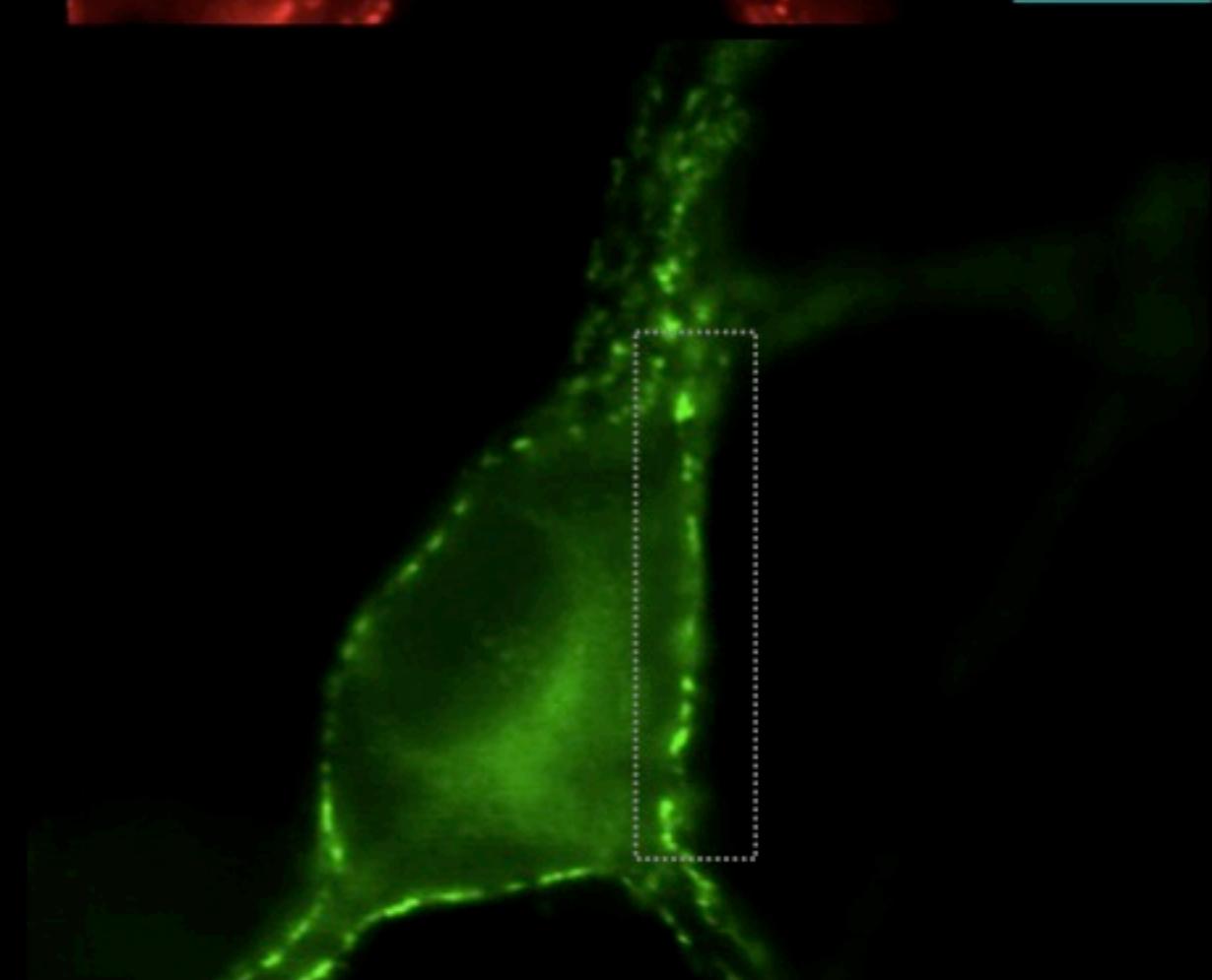
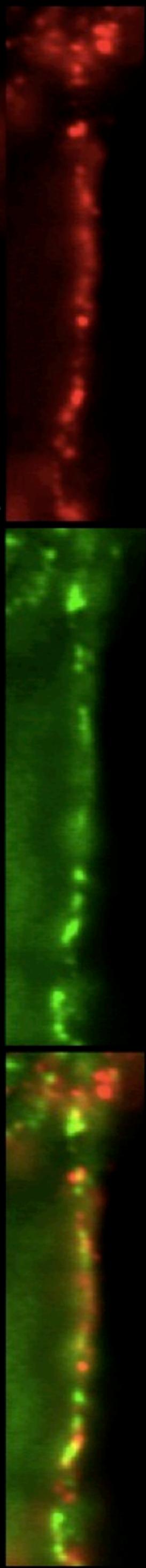
*Synapsine*

Tau

VIAAT

*Superposition*

21 DIV



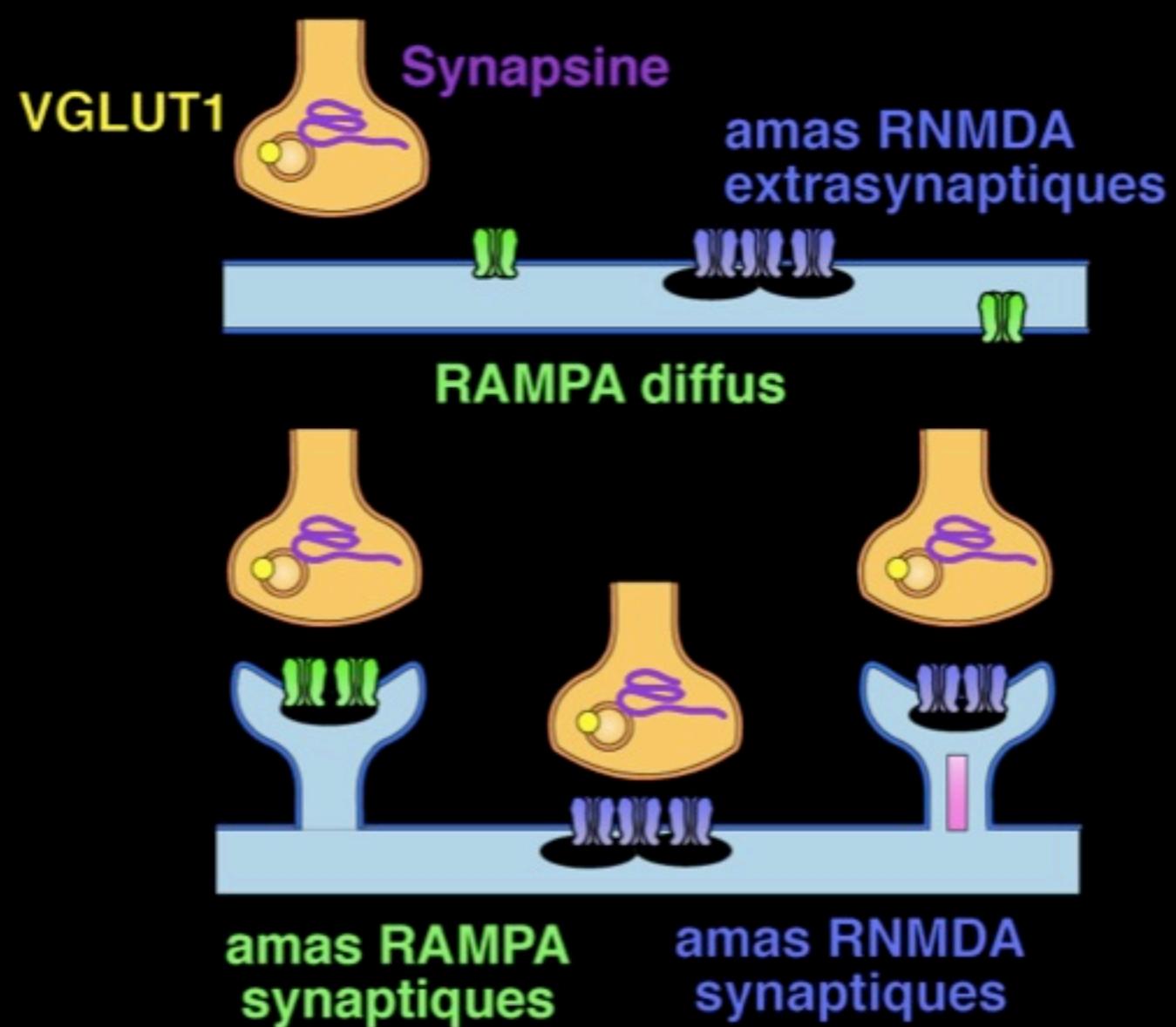
VIAAT: Expression diffuse dans l'axone puis formation d'amas dès 10 jours de culture.

# Ciblage des protéines post-synaptiques

## Synapse excitatrice hippocampe

Laboratoire A.M. Craig

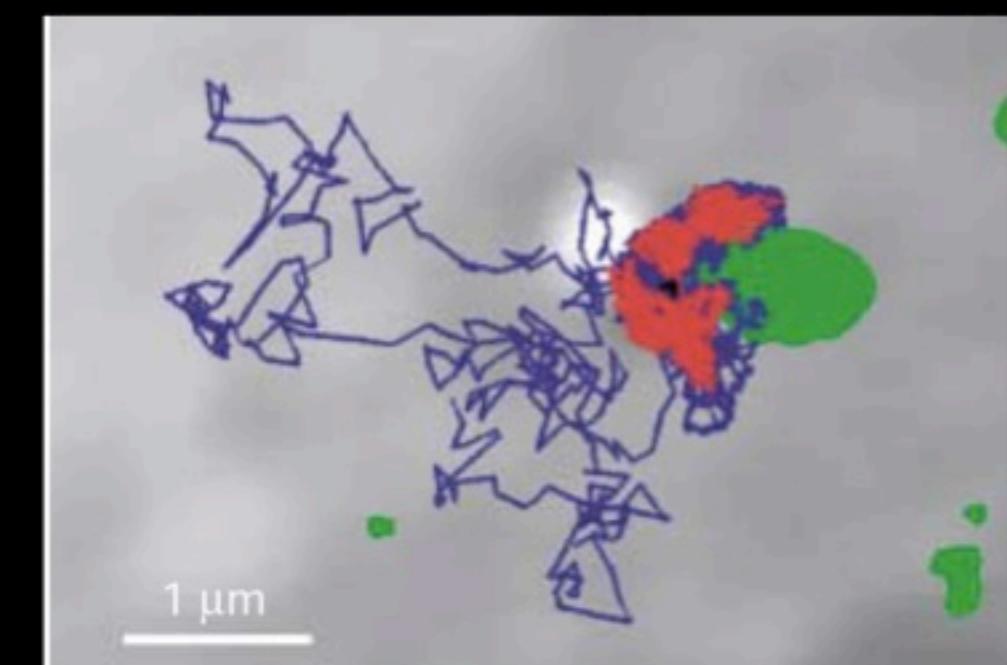
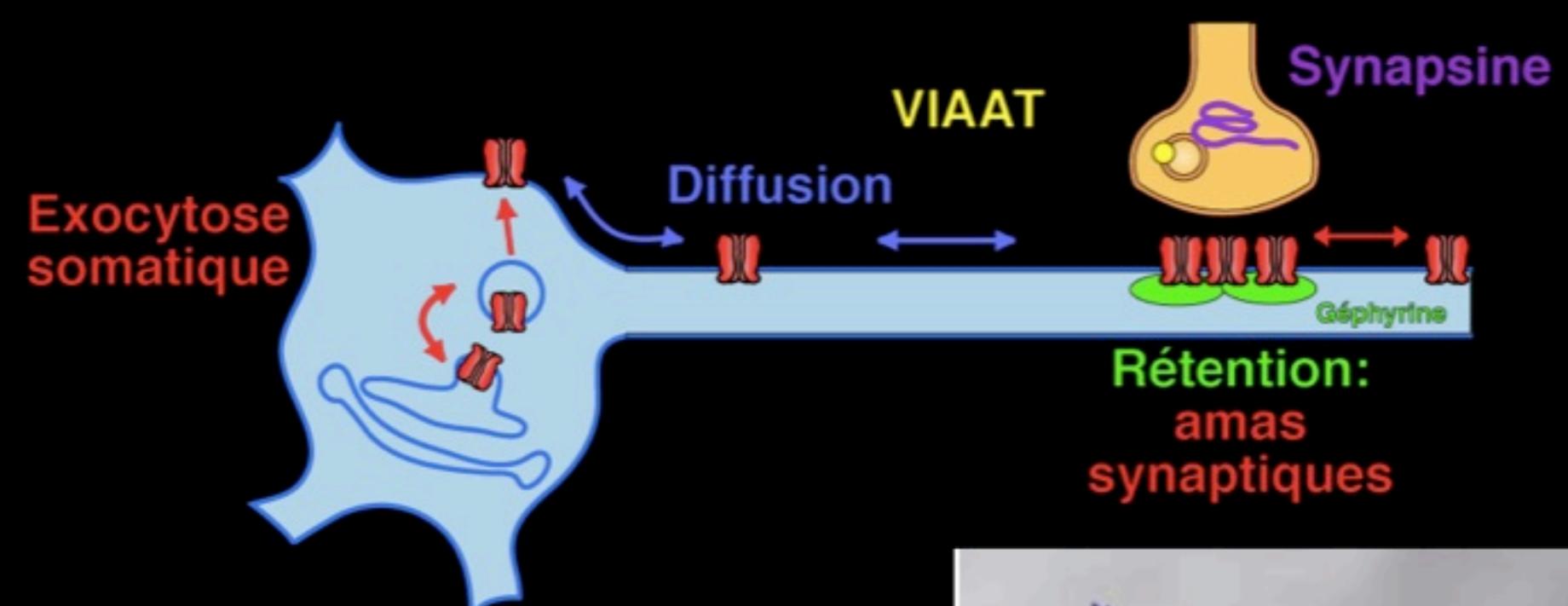
Rao et al., J. Neurosci., 2000  
Rao et al., J. Neurosci., 2000



## Synapse inhibitrice moelle épinière

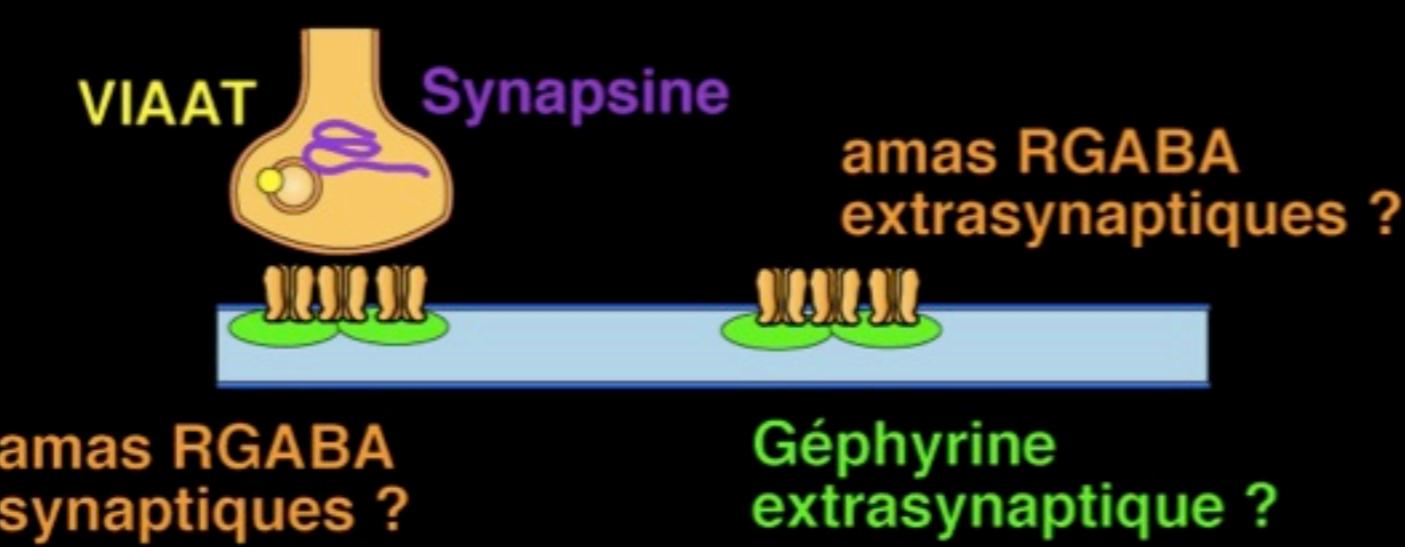
Laboratoire A.Triller

Rosenberg et al., J. Neurosci, 2000  
Meier et al, JCS, 2000  
Meier et al., Nat. Neur., 2001



## Synapse inhibitrice hippocampe

?



# Importance de l' établissement de 2 compartiments distincts: Axonal <> Somatodendritique

- Passage stade 2 à 3
- Instabilité de l' actine
- Assemblage des microtubules
- Activation de la voie PI3K-AKT-GSK3b-CRMP2
- Différents moyens d' atteindre le compartiment:  
transport sélectif, ou tri par endocytose
- Formation de contacts polarisés: les synapses....